

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
13 February 2003 (13.02.2003)

PCT

(10) International Publication Number
WO 03/012116 A2

(51) International Patent Classification⁷: **C12N 15/82**,
9/50, 15/57

1X8 (CA). **HUANG, Yafan** [CA/CA]; 935 Milford Drive, Kingston, Ontario K7P 1X8 (CA). **CAMPBELL, Mary-Jane, D.** [CA/CA]; 1066 King St. West, Apt. 505, Kingston, Ontario K7M 9CR (CA). **KUZMA, Monika, M.** [CA/CA]; 507 Aragon Road, Glenburnie, Ontario K0H 1S0 (CA). **GILLETT, Angela, P.** [CA/CA]; 1262 Sunbury Road, R.R.#2, Inverary, Ontario K0H 1X0 (CA).

(21) International Application Number: PCT/IB02/03887

(22) International Filing Date: 1 August 2002 (01.08.2002)

(25) Filing Language: English

(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW.

(26) Publication Language: English

(30) Priority Data:
US 60/309,396 1 August 2001 (01.08.2001) US
US 60/337,084 4 December 2001 (04.12.2001) US

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

(63) Related by continuation (CON) or continuation-in-part (CIP) to earlier applications:

US 60/309,396 (CIP)
Filed on 1 August 2001 (01.08.2001)
US 60/337,084 (CIP)
Filed on 4 December 2001 (04.12.2001)

Published:

— without international search report and to be republished upon receipt of that report

(71) Applicant (for all designated States except US): **PERFORMANCE PLANTS, INC.** [CA/CA]; c/o Queens University, Bioscience Complex, Kingston, Ontario K7L 3N6 (CA).

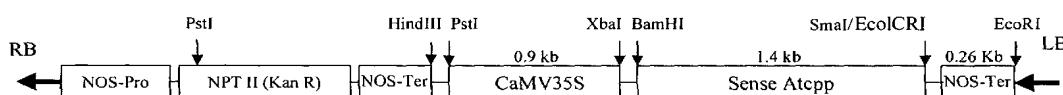
For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(72) Inventors; and

(75) Inventors/Applicants (for US only): **WAN, Jiangxin** [CA/CA]; 935 Milford Drive, Kingston, Ontario K7M

WO 03/012116 A2

(54) Title: CAAX PRENYL PROTEASE NUCLEIC ACIDS AND POLYPEPTIDES AND METHODS OF USE THEREOF



(57) Abstract: The present invention provides novel isolated prenyl protease polynucleotides and polypeptides encoded by the prenyl protease polynucleotides. Also provided are the antibodies that immunospecifically bind to a prenyl protease polypeptide or any derivative, variant, mutant or fragment of the prenyl protease polypeptide, polynucleotide or antibody. The invention additionally provides methods of constructing transgenic plants that have altered levels of prenyl protease polynucleotides and polypeptides. Methods for identifying prenyl protease enzymes substrates and inhibitors are also provided.

CaaX Prenyl Protease Nucleic Acids and Polypeptides and Methods of Use Thereof

FIELD OF THE INVENTION

The invention relates to novel plant CaaX prenyl protease polynucleotides and polypeptides. Also included are transgenic plants expressing the novel polynucleotides and polypeptides. Also included are transgenic plant cells, tissues and plants having novel phenotypes resulting from the expression of these polynucleotides in either the sense or antisense orientation.

10

BACKGROUND OF THE INVENTION

Most higher plants encounter at least transient decreases in relative water content at some stage of their life cycle and, as a result, have evolved a number of desiccation protection mechanisms. If however, the change in water deficit is prolonged the effects on the plants growth and development can be profound. Decreased water content due to drought, cold or salt stress can irreparably damage plant cells which in turn limits plant growth and crop productivity in agriculture.

Plants respond to adverse conditions of drought, salinity and cold with a variety of morphological and physiological changes. Although our understanding of plant tolerance mechanisms to these stresses is incomplete, the plant hormone abscisic acid (ABA) is believed to be an essential mediator between environmental stimulus and plant responses. ABA levels increase in response to water deficits and exogenously applied ABA mimics many of the responses induced by water-stress. Once ABA is synthesized it causes the closure of the leaf stomata thereby decreasing water loss through transpiration.

The identification of genes that transduce ABA into a cellular response opens the possibility of exploiting these regulators to enhance desiccation tolerance in crop species. In principle, these ABA signaling genes can be coupled with the appropriate controlling elements to allow optimal plant growth, development and productivity. Thus, not only would these genes allow the genetic tailoring of crops to withstand transitory environmental stresses, but they should also broaden the environments where traditional crops can be grown.

The recent isolation of an *Arabidopsis* mutant, *era1*, is hypersensitive to ABA and has been shown to also be tolerant to conditions of water deprivation. ERA1 has been identified as a β subunit of farnesyl transferase knockout mutant in. Farnesyl transferase is a heterodimeric enzyme that provides the specific addition of a farnesyl pyrophosphate moiety onto the substrate target sequence. The target sequence is defined as a sequence of four amino acids which are present at the carboxy terminus of the protein and is referred to as a CaaX motif in which the "C" is cysteine, "a" is any aliphatic amino acid and "X" is any amino acid. The α subunit is common with a second prenylation enzyme, geranylgeranyl transferase, that has a different β subunit and adds a geranylgeranyl isoprenyl pyrophosphate moiety to the target sequence.

Prenylation is a multistep pathway which includes prenylation of the cysteine residue of the CaaX site, cleavage of the -aaX tripeptide and methylation of the prenylcysteine residue. Potentially, each of these steps could represent a target for genetic manipulation of the prenylation process to generate a desired phenotype such as stress tolerance.

In plants, prenylation has been linked to cell cycle control, meristem development, and phytohormone signal transduction, however, few details of the role of prenylation, the substrate proteins or the extent to which the plant system will be analogous to the mammalian and yeast systems are known. The most characterized substrates for CaaX modification are the Ras and a-factor proteins of yeast. Although there are three steps to complete protein maturation, abolition or modification of any one step does not necessarily result in cessation of target biological activities. Ras function is attenuated if the -aaX tripeptide is not cleaved but not abolished and some proteins retain the -aaX tripeptide after farnesylation.

In *Arabidopsis*, more than 600 proteins contain a CaaX motif, suggesting a role for the post-translational modification by prenylation in numerous cellular processes. In *Arabidopsis*, it has been demonstrated that the loss-of-function of the β -subunit of farnesyl transferase will result in a ABA-hypersensitive phenotype. Although it is still not clear why plants lacking the functional β -subunit of farnesyl transferase become more sensitive to ABA, it clearly suggests that protein prenylation is involved in regulation of the homeostasis of ABA sensitivity. The balance of ABA cellular responses, whether more sensitive or less sensitive to ABA, is possibly regulated by the

relative activities of prenylated proteins. The changes in AtCPP expression and gene activity may affect the activity of two pools of genes, one pool acting as positive regulators (pool A) and the second pool (pool B) as negative regulators, which require prenylation in order to function properly. Pool A may contain genes that can promote 5 ABA sensitivity, and pool B genes that may reduce ABA sensitivity. The homeostasis of ABA sensitivity may therefore governed by the ratio of activity of pool A to pool B. For example, in the case of up-regulation of AtCPP in *Arabidopsis*, the activity ratio of pool A over pool B may be increased due to difference in substrate affinity of pool A proteins toward AtCPP, thus the homeostasis of ABA sensitivity is changed, and the AtCPP over-expression plants are more sensitive to ABA.

10 This invention is directed at the manipulation of the CaaX prenyl protease enzyme (CPP), which catalyses the proteolytic cleavage of the -aaX tripeptide in the second step of the prenylation process. Included in this invention are vector constructs containing CPP sequence under the control of appropriate regulatory sequences to 15 produce a water-stress tolerant phenotype.

SUMMARY OF THE INVENTION

The present invention is based in part upon the discovery of novel CaaX prenyl protease (CPP) nucleic acid sequences and polypeptides isolated from *Arabidopsis thaliana*, *Brassica napus* and *Glycine max*. The nucleic acids, polynucleotides, proteins 20 and polypeptides, or fragments thereof described herein are collectively referred to as CPP nucleic acids and polypeptides.

Accordingly, in one aspect, the invention provides an isolated nucleic acid molecule that includes the sequence of SEQ ID NO:1, SEQ ID NO:14, or SEQ ID NO:17 or fragment, homolog, analog or derivative thereof. The nucleic acid can include, e.g., a 25 nucleic acid sequence encoding a polypeptide at least 99% identical to a polypeptide that includes the amino acid sequences of SEQ ID NO:2, SEQ ID NO:15, or SEQ ID NO:18 or a nucleic acid sequence encoding a polypeptide at least 96% identical to a polypeptide that includes the amino acid sequences of SEQ ID NO:15. In yet another aspect, the invention provides a nucleic acid that includes the sequence of SEQ ID NO: 68, 70, 72 or 74. The nucleic acid can be, e.g., a genomic DNA fragment, or a cDNA molecule. 30 Preferably, the nucleic acid is naturally occurring. The invention also provides a nucleic

acid sequence that is complementary to the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:14, or SEQ ID NO:17. For example, SEQ ID NO: 16, 19 or 20.

Also included in the invention is a vector containing one or more of the nucleic acids described herein, and a cell containing the vectors or nucleic acids described
5 herein. In various aspects the vector comprises the nucleic acid sequences of SEQ ID NO: 4, 5, 36-53.

The invention is also directed to host cells transformed with a vector comprising any of the nucleic acid molecules described above.

10 The invention is also directed to plants and cells transformed with a CPP nucleic acid or a vector comprising a CPP nucleic acid. Also included in the invention is the seed, and progeny of the transformed plants or cells.

15 In a further aspect, the invention includes a substantially purified CPP polypeptide, *e.g.*, any of the CPP polypeptides encoded by an CPP nucleic acid, and fragments, homologs, analogs, and derivatives thereof. Accordingly, in one aspect, the invention provides an isolated polypeptide molecule that includes the sequence of SEQ ID NO:2, SEQ ID NO:15, or SEQ ID NO:18.

In yet another aspect the invention provides a polypeptides that includes the sequence of SEQ ID NO: 69, 71, 73 or 75.

20 In still a further aspect, the invention provides an antibody that binds specifically to an CPP polypeptide. The antibody can be, *e.g.*, a monoclonal or polyclonal antibody, and fragments, homologs, analogs, and derivatives thereof. The invention is also directed to isolated antibodies that bind to an epitope on a polypeptide encoded by any of the nucleic acid molecules described above.

25 The invention also includes a method of producing a transgenic plant which has an altered phenotype such as, but not limited to, increased tolerance to stress, delayed senescence, increased ABA sensitivity, increased yield, increased productivity and increased biomass compared to a wild type plant by introducing into one or more cells of a plant a compound that alters (*e.g.*, increases or decreases) CPP expression or activity in the plant. In one aspect the compound is a CPP nucleic acid or polypeptide. In one
30 embodiment the nucleic acid is an inhibitor or farnesylation. For example, the compound comprises SEQ ID NO: 1, 14, 17, 68, 70, 72, 74, 21, 23, 25, 27, 29, 31, 33, 2, 15, 18, 22, 24 26, 28, 30, 32 34, 69, 71, 73, or 75. Alternatively, the compound is a CPP double stranded RNA-inhibition hair-pin nucleic acid or CPP antisense nucleic acid, such as for example, SEQ ID NO: 16, 19, 20, 5, 35, 37, 42, 45, 46, 48, 49, 51 or 51.

The invention further provides a method for producing a CPP polypeptide by providing a cell containing an CPP nucleic acid, *e.g.*, a vector that includes a CPP nucleic acid, and culturing the cell under conditions sufficient to express the CPP polypeptide encoded by the nucleic acid. The expressed CPP polypeptide is then 5 recovered from the cell. Preferably, the cell produces little or no endogenous CPP polypeptide. The cell can be, *e.g.*, a prokaryotic cell or eukaryotic cell.

The invention is also directed to methods of identifying a CPP polypeptide or nucleic acid in a sample by contacting the sample with a compound that specifically binds to the polypeptide or nucleic acid, and detecting complex formation, if present. 10 The invention further provides methods of identifying a compound that modulates the activity of a CPP polypeptide by contacting a CPP polypeptide with a compound and determining whether the CPP polypeptide activity is modified.

The invention is also directed to compounds that modulate CPP polypeptide activity identified by contacting a CPP polypeptide with the compound and determining 15 whether the compound modifies activity of the CPP polypeptide, binds to the CPP polypeptide, or binds to a nucleic acid molecule encoding a CPP polypeptide.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those 20 described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In the case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not 25 intended to be limiting.

Other features and advantages of the invention will be apparent from the following detailed description and claims.

BRIEF DESCRIPTION OF THE DRAWINGS

30 Figure 1. is a schematic representation of the vector constructs; A) pBI121-AtCPP, B) pBI121-antisense-AtCPP, C) pBI121-HP-AtCPP.

Figure 2. is an illustration of (A) nucleic acid and (B) amino acid sequence identities as determined by ClustalW analysis.

Figure 3. is a scan of a typical Southern blot of transgenic *Arabidopsis* T1 lines carrying the pBI121-AtCPP construct.

5 Figure 4. is a scan of a typical Southern blot of transgenic *Arabidopsis* T3 lines carrying the pBI121-HP-AtCPP construct.

Figure 5. is a scan of a typical Southern blot of transgenic *Arabidopsis* lines carrying the pRD29A-AtCPP construct.

10 Figure 6. is a scan of a typical Southern blot of transgenic *Arabidopsis* lines carrying the pRD29A-HP-AtCPP construct.

Figure 7 is an illustration showing the relative expression of AtCPP mRNA transcript (solid bars) and AtCPP protein levels (stippled bars) in several pBI121-AtCPP transgenic lines.

15 Figure 8. is a histogram showing the percentage of lines which were categorized as ABA sensitive, moderately ABA sensitive or ABA insensitive. Seedlings were assessed on agar plates containing 1 µM ABA and scored at 21 days growth. Thirty-six lines of the pBI121-AtCPP over-expression construct were assessed at 21 days by leaf and seedling development. Thirty-two lines of the 35S-HP-AtCPP down-regulation construct were assessed at 21 days for leaf and seedling 20 development. Each line was assessed by plating approximately 100 seeds per plate and the seedlings scored and recorded as the percent insensitive seedlings per plate. Each line was then expressed as a percent of wild type (Wt). Lines were categorized as sensitive (less than 1% of Wt) solid bars, intermediate (1-50% of Wt) diagonally lined or insensitive (greater than 50% of Wt) stippled, based on 25 their relationship to Wt and the percentage of each category plotted as a histogram.

Figure 9. is an illustration showing the response of wild type and a pRD29A-HP-AtCPP transgenic line to various concentrations of ABA in two week old seedlings.

30 Figure 10. is a histogram showing the analysis of transgenic plants containing the pBI121-AtCPP over-expression construct, (SEQ ID NO:4). Water loss per gram shoot dry weight after four days of water stress treatment. Lines that are marked

with a star are those which were strongly ABA sensitive. Lines marked with a triangle are moderately ABA sensitive. Bars represent means of eight replicates. Lines marked with a filled dot above the bar represents lines which were significantly different from control at a p=0.05 value.

5 Figure 11. is a histogram showing seed yield in grams of transgenic *Arabidopsis* lines of pBI121-AtCPP grown under optimal water conditions

Figure 12. is a bar chart showing growth and yield of transgenic *Arabidopsis* lines of pBI121-AtCPP grown under optimal watering conditions plus a biotic stress condition. Yields as a % of wild type, rosette leaf number, rosette leaf fresh weight and shoot dry weight are plotted.
10

Figure 13. are photographs showing growth of transgenic *Arabidopsis* lines of pBI121-AtCPP grown on agar plates. Changes to root growth visible.

Figure 14. is a bar chart showing growth of transgenic *Arabidopsis* lines of pRD29A-HP-AtCPP grown under optimal watering conditions. Rosette leaf number, rosette leaf dry weight and shoot dry weight are plotted.
15

DETAILED DESCRIPTION OF INVENTION

The present invention provides novel CaaX prenyl protease (CPP) nucleic acid sequences (SEQ ID No:1, SEQ ID NO:14 and SEQ ID NO:17) the encoded polypeptides: SEQ ID NO:2, SEQ ID NO:15 and SEQ ID NO:18) isolated from *Arabidopsis thaliana* (At) *Brassica napus* (Bn) and *Glycine Max* (Gm) respectively. The invention also provides CaaX prenyl protease antisense nucleic acids. (SEQ ID NO: 16, SEQ ID NO:19 and SEQ ID NO:20). The sequences are collectively referred to as “CPP nucleic acids”, CPP polynucleotides” or “CPP antisense nucleic acids” and the corresponding encoded polypeptide is referred to as a “CPP polypeptide” or “CPP protein”. Unless indicated otherwise, “CPP” is meant to refer to any of the novel sequences disclosed herein. Table A below summarizes the nucleic acids and polypeptides according to the invention
25
30

Table A

SEQ ID NO.	SEQ	Type	Species Transformed
------------	-----	------	---------------------

1	AtCPP	NA	PCR	
2	AtCPP	AA	Translation	
3	At-AFC1	AA	Ref.	
4	pBI121-AtCPP	NA	Construct	At, Bn
5	pBI121-HP-AtCPP	NA	Construct	At
6	AtCPP BamFW	NA	Primer	
7	AtCPP SmaRV	NA	Primer	
8	AtCPP-HP-SacFW	NA	Primer	
9	AtCPP-HP-SacRV	NA	Primer	
10	pBI121-AtCPP Forward	NA	Primer	
11	pBI121-antiAtCPP-SmaFW	NA	Primer	
12	pBI121-antiAtCPP-BamRV	NA	Primer	
13	p35S-HP-AtCPP Reverse	NA	Primer	
14	BnCPP	NA	PCR	
15	BnCPP	AA	Translation	
16	BnCPP antisense	NA	PCR	
17	GmCPP	NA	PCR	
18	GmCPP	AA	Translation	
19	GmCPP antisense	NA	PCR	
20	AtCPP antisense	NA	PCR	
21	BASF-AT1	NA	Ref.	
22	BASF-AT1	AA	Ref.	
23	BASF-AT2	NA	Ref.	
24	BASF-AT2	AA	Ref.	
25	BASF-Corn	NA	Ref.	
26	BASF-Corn	AA	Ref.	
27	BASF-Soy	NA	Ref.	
28	BASF-Soy	AA	Ref.	
29	AFC1	NA	Ref.	
30	AFC1	AA	Ref.	
31	AT4g01320	NA	Ref.	
32	AT4g01320	AA	Ref.	
33	AF007269	NA	Ref.	
34	AF007269	AA	Ref.	
35	pBI121-antisense-AtCPP	NA	Construct	
36	pRD29A-AtCPP	NA	Construct	At, Bn
37	pRD29A-HP-AtCPP	NA	Construct	At
38	pRD29A-antisense-AtCPP	NA	Construct	
39	MuA-AtCPP	NA	Construct	Gm, Zm
40	MuA-GmCPP	NA	Construct	
41	pBI121-GmCPP		Construct	
42	pBI121-HP-GmCPP		Construct	
43	pBI121-antisense-GmCPP		Construct	
44	pRD29A-GmCPP		Construct	
45	pRD29A-HP-GmCPP		Construct	
46	pRD29A-antisense-GmCPP		Construct	
47	pBI121-BnCPP		Construct	
48	pBI121-HP-BnCPP		Construct	
49	pBI121-antisense-BnCPP		Construct	
50	pRD29A-BnCPP		Construct	
51	pRD29A-HP-BnCPP		Construct	
52	pRD29A-antisense-BnCPP		Construct	
53	MuA-BnCPP		Construct	
54	GmCPP SmaFW		Primer	
55	GmCPP SacRV		Primer	
56	BnCPP-anti-SmaFW		Primer	
57	BnCPP-anti-BamRV		Primer	
58	BnCPP-HP-Sac-FW		Primer	

59	BnCPP-HP-Sac-RV	Primer	
60	BnCPP-HP-BamFW	Primer	
61	BnCPP-HP-XbaRV	Primer	
62	GmCPP-HP-Sac-FW	Primer	
63	GmCPP-HP-Sac-RV	Primer	
64	GmCPP-HP-BamFW	Primer	
65	GmCPP-HP-XbaRV	Primer	
66	pRD29AP	Primer	
67	NoTerm-RV	Primer	
68	Consensus- BASF	NA	
69	Consensus- BASF	AA	
70	Consensus- Generic	NA	
71	Consensus- Generic	AA	
72	Consensus- PPI	NA	
73	Consensus- PPI	AA	
74	Consensus- PPI/Generic	NA	
75	Consensus- PPI/Generic	AA	

In a BLAST search of public sequence databases, it was found, for example, that the *Arabidopsis thaliana* nucleic acid sequence has 99.5 % identity to an *Arabidopsis thaliana* CaaX processing zinc-metallo endoprotease (AFC1) mRNA (Genbank Accesion No.: AF353722). The full amino acid sequence of the protein of the invention was found to be 98.8 % identical to *Arabidopsis thaliana* CaaX processing zinc-metallo endoprotease (AFC1) polypeptide (Genbank Accesion No.:AAK39514). A ClustalW alignment of the *Arabidopsis thaliana* CPP polypeptide (SEQ ID NO:2), the *Brassica napus* CPP polypeptide (SEQ ID NO:15), the *Glycine max* CPP polypeptide (SEQ ID NO:18) and seven other published CPP sequences is illustrated in Table 6B. ClustalW alignment of these polypeptides indicate that SEQ ID NO:2, SEQ ID NO:15 and SEQ ID NO:18 are 99%, 93% and 83% identical to the published AFC sequence (SEQ ID NO:30) respectively. The *Glycine max* CPP polypeptide (SEQ ID NO:18) is 99% identical to the published sequence shown as SEQ ID NO:28. Similarly, ClustalW alignment of the *Arabidopsis thaliana* CPP polynucleotide (SEQ ID NO:1), the *Brassica napus* CPP polynucleotide (SEQ ID NO:14), the *Glycine max* CPP polynucleotide (SEQ ID NO:17) and seven other published CPP sequences is illustrated in Table 6a indicate that SEQ ID NO:1, SEQ ID NO:14 and SEQ ID NO:17 are 99%, 93% and 77% identical to the published AFC sequence (SEQ ID NO:30) respectively. The *Glycine max* CPP polynucleotide (SEQ ID NO:17) is 93% identical to the published sequence shown as SEQ ID NO:27.

CaaX prenyl proteases belong to a family of putative membrane-bound proteins that are involved in protein and/or peptide modification (*i.e.*, prenylation) and secretion. Prenylation is a post translational modification of specific proteins and is required for the

proper localization of these polypeptides to the correct cellular site for functionality.

Prenylation is a three step process involving the addition of either a C15 farnesyl, or C20 geranylgeranyl group to the cysteine residue of the target 3' terminal CaaX sequence, where "C" is a cysteine, "a" is any aliphatic amino acid and "X" is any amino acid.

- 5 Secondly, a CaaX prenyl protease (CPP) cleaves the -aaX tripeptide from the protein and thirdly the exposed α -carboxyl group of the cysteine is methylated by a prenyl-cysteine methyltransferase.

Protein farnesylation, the addition of a C-terminal, 15 carbon chain to protein and subsequent processing is a three step enzymatic reaction including farnesylation,

- 10 proteolytic cleavage and methylation. First, a farnesyltransferase adds the C-terminal 15 carbon chain to a conserved cysteine residue of the CaaX terminal motif, where "C" is a Cystine, "a" is an aliphatic amino acid and "X" is any amino acid. Second, the last three amino acid residues (aaX) are cleaved by a prenyl protease. Lastly, the modified cysteine is methylated by a methylase to create the final active product of the
- 15 protein farnesylation pathway. The Applicant's have shown previously that over expression and down-regulation of the alpha or the beta farnesyl transferase gene in plant cells (i.e, the first step in farnesylation) results in plants with an altered phenotype such as but not limited to drought tolerance and delayed senescence. The present invention shows that over expression and down-regulation of the prenyl protease gene (i.e, the
- 20 second step in farnesylation) in plant cells also results in a plant displaying an altered phenotype including for example but not limited to drought tolerance and increased resistance to biotic and abiotic stress. These results taken together support the hypothesis that modification of the expression of any of the enzymes in the farnesylation pathway in a plant cell will result in a plant displaying an altered
- 25 phenotype

Based on their structural and functional relatedness to known CaaX prenyl protease proteins, the CPP proteins are novel members of the CaaX prenyl protease family of proteins. CPP nucleic acids, and their encoded polypeptides, according to the invention are useful in a variety of applications and contexts. For example, the nucleic acids (i.e., sense or antisense CPP nucleic acids) can be used to produce transgenic plants that have an increase resistance to biotic and abiotic stresses, *e.g.*, chilling stress, salt stress, water stress, wound healing, pathogen challenge, or herbicides. Additionally, the transgenic plants have an increased productivity during both optimal and suboptimal

growth conditions, increased yield, or increased biomass. Alternatively, the transgenic plants have an increased sensitivity to the phytohormone abscisic acid (ABA).

This invention includes methods to up-regulate the CPP enzyme activity in transgenic plants, cells and tissue cultures by using an over-expression vector construct
5 and methods to down-regulate the CPP enzyme activity in transgenic plants, cells and tissue cultures by using a double stranded RNA-inhibition, hairpin vector constructs or antisense constructs. Alteration (i.e., upregulation or downregulation) of CPP enzyme activity or expression results in transgenic plants with altered phenotypes as described below. These methods are by way of example to produce the up-regulation or down-
10 regulation effects and are not meant to be limiting as to the method of achieving this outcome.

Additionally, the nucleic acids and polypeptides according to the invention may be used as targets for the identification of small molecules that modulate or inhibit, CPP activity. Alternatively, the CPP nucleic acids and polypeptides can be used to identify
15 proteins that are members of the CaaX prenyl protease family of proteins.

Additional utilities for CPP nucleic acids and polypeptides according to the invention are disclosed herein.

CPP Nucleic Acids

The nucleic acids of the invention include those that encode a CPP polypeptide or protein. As used herein, the terms polypeptide and protein are interchangeable.
20

In some embodiments, a CPP nucleic acid encodes a mature CPP polypeptide. As used herein, a “mature” form of a polypeptide or protein described herein relates to the product of a naturally occurring polypeptide or precursor form or proprotein. The naturally occurring polypeptide, precursor or proprotein includes, by way of nonlimiting
25 example, the full length gene product, encoded by the corresponding gene.

Alternatively, it may be defined as the polypeptide, precursor or proprotein encoded by an open reading frame described herein. The product “mature” form arises, again by way of nonlimiting example, as a result of one or more naturally occurring processing steps that may take place within the cell in which the gene product arises. Examples of
30 such processing steps leading to a “mature” form of a polypeptide or protein include the cleavage of the N-terminal methionine residue encoded by the initiation codon of an open reading frame, or the proteolytic cleavage of a signal peptide or leader sequence.

Thus a mature form arising from a precursor polypeptide or protein that has residues 1 to N, where residue 1 is the N-terminal methionine, would have residues 2 through N remaining after removal of the N-terminal methionine. Alternatively, a mature form arising from a precursor polypeptide or protein having residues 1 to N, in which an N-terminal signal sequence from residue 1 to residue M is cleaved, would have the residues from residue M+1 to residue N remaining. Further as used herein, a "mature" form of a polypeptide or protein may arise from a step of post-translational modification other than a proteolytic cleavage event. Such additional processes include, by way of non-limiting example, glycosylation, myristoylation or phosphorylation. In general, a mature polypeptide or protein may result from the operation of only one of these processes, or a combination of any of them.

Among the CPP nucleic acids is the nucleic acid whose sequence is provided in SEQ ID NO: 1, SEQ ID NO:14 OR SEQ ID NO:17 or a fragment thereof. Additionally, the invention includes mutant or variant nucleic acids of SEQ ID NO: 1, SEQ ID NO:14 OR SEQ ID NO:17 or a fragment thereof, any of whose bases may be changed from the corresponding base shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17, while still encoding a protein that maintains at least one of its CPP-like activities and physiological functions. The invention further includes the complement of the nucleic acid sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17, including fragments, derivatives, analogs and homologs thereof. Complement nucleic acid CPP sequences include SEQ ID NO: 16, 19 or 20. The invention additionally includes nucleic acids or nucleic acid fragments, or complements thereto, whose structures include chemical modifications.

One aspect of the invention pertains to isolated nucleic acid molecules that encode CPP proteins or biologically active portions thereof. Also included are nucleic acid fragments sufficient for use as hybridization probes to identify CPP-encoding nucleic acids (*e.g.*, CPP mRNA) and fragments for use as polymerase chain reaction (PCR) primers for the amplification or mutation of CPP nucleic acid molecules. As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (*e.g.*, cDNA or genomic DNA), RNA molecules (*e.g.*, mRNA), analogs of the DNA or RNA generated using nucleotide analogs, and derivatives, fragments and homologs thereof. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

"Probes" refer to nucleic acid sequences of variable length, preferably between at least about 10 nucleotides (nt), 100 nt, or as many as about, e.g., 6,000 nt, depending on use. Probes are used in the detection of identical, similar, or complementary nucleic acid sequences. Longer length probes are usually obtained from a natural or recombinant source, are highly specific and much slower to hybridize than oligomers. Probes may be single- or double-stranded and designed to have specificity in PCR, membrane-based hybridization technologies, or ELISA-like technologies.

An "isolated" nucleic acid molecule is one that is separated from other nucleic acid molecules that are present in the natural source of the nucleic acid. Examples of isolated nucleic acid molecules include, but are not limited to, recombinant DNA molecules contained in a vector, recombinant DNA molecules maintained in a heterologous host cell, partially or substantially purified nucleic acid molecules, and synthetic DNA or RNA molecules. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (*i.e.*, sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated CPP nucleic acid molecule can contain less than about 50 kb, 25 kb, 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material or culture medium when produced by recombinant techniques, or of chemical precursors or other chemicals when chemically synthesized.

A nucleic acid molecule of the present invention, e.g., a nucleic acid molecule having the nucleotide sequence of SEQ ID NO: 1, SEQ ID NO:14, or SEQ ID NO:17 or a complement of any of this nucleotide sequence, can be isolated using standard molecular biology techniques and the sequence information provided herein. Using all or a portion of the nucleic acid sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 as a hybridization probe, CPP nucleic acid sequences can be isolated using standard hybridization and cloning techniques (*e.g.*, as described in Sambrook *et al.*, eds., 25 MOLECULAR CLONING: A LABORATORY MANUAL 2nd Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989; and Ausubel, *et al.*, eds., CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, New York, NY, 1993.)

A nucleic acid of the invention can be amplified using cDNA, mRNA or alternatively, genomic DNA, as a template and appropriate oligonucleotide primers

according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis.

Furthermore, oligonucleotides corresponding to CPP nucleotide sequences can be prepared by standard synthetic techniques, *e.g.*, using an automated DNA synthesizer.

- 5 As used herein, the term “oligonucleotide” refers to a series of linked nucleotide residues, which oligonucleotide has a sufficient number of nucleotide bases to be used in a PCR reaction. A short oligonucleotide sequence may be based on, or designed from, a genomic or cDNA sequence and is used to amplify, confirm, or reveal the presence of an identical, similar or complementary DNA or RNA in a particular cell or tissue.
- 10 Oligonucleotides comprise portions of a nucleic acid sequence having about 10 nt, 50 nt, or 100 nt in length, preferably about 15 nt to 30 nt in length. In one embodiment, an oligonucleotide comprising a nucleic acid molecule less than 100 nt in length would further comprise at least 6 contiguous nucleotides of SEQ ID NO: 1, 14 or 17, or a complement thereof. Oligonucleotides may be chemically synthesized and may be used
- 15 as probes.

In another embodiment, an isolated nucleic acid molecule of the invention includes a nucleic acid molecule that is a complement of the nucleotide sequence shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17. In another embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule that is a complement of the nucleotide sequence shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17, or a portion of these nucleotide sequence. A nucleic acid molecule that is complementary to the nucleotide sequence shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 is one that is sufficiently complementary to the nucleotide sequence shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 that it can hydrogen bond with little or no mismatches to the nucleotide sequence shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17, thereby forming a stable duplex. Exemplary complement nucleic acid sequences include the sequences of SEQ ID NO: 16, 19 or 20.

As used herein, the term “complementary” refers to Watson-Crick or Hoogsteen base pairing between nucleotide units of a nucleic acid molecule, and the term “binding” means the physical or chemical interaction between two polypeptides or compounds or associated polypeptides or compounds or combinations thereof. Binding includes ionic, non-ionic, Von der Waals, hydrophobic interactions, etc. A physical interaction can be either direct or indirect. Indirect interactions may be through or due to the effects of another polypeptide or compound. Direct binding refers to interactions that do not take

place through, or due to, the effect of another polypeptide or compound, but instead are without other substantial chemical intermediates.

Moreover, the nucleic acid molecule of the invention can comprise only a portion of the nucleic acid sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17, e.g.,

5 a fragment that can be used as a probe or primer, or a fragment encoding a biologically active portion of CPP. Fragments provided herein are defined as sequences of at least 6 (contiguous) nucleic acids or at least 4 (contiguous) amino acids, a length sufficient to allow for specific hybridization in the case of nucleic acids or for specific recognition of an epitope in the case of amino acids, respectively, and are at most some portion less

10 than a full length sequence. Fragments may be derived from any contiguous portion of a nucleic acid or amino acid sequence of choice. Derivatives are nucleic acid sequences or amino acid sequences formed from the native compounds either directly or by modification or partial substitution. Analogs are nucleic acid sequences or amino acid sequences that have a structure similar to, but not identical to, the native compound but

15 differs from it in respect to certain components or side chains. Analogs may be synthetic or from a different evolutionary origin and may have a similar or opposite metabolic activity compared to wild type.

Derivatives and analogs may be full length or other than full length, if the derivative or analog contains a modified nucleic acid or amino acid, as described below.

20 Derivatives or analogs of the nucleic acids or proteins of the invention include, but are not limited to, molecules comprising regions that are substantially homologous to the nucleic acids or proteins of the invention, in various embodiments, by at least about 70%, 80%, 85%, 90%, 95%, 98%, or even 99% identity (with a preferred identity of 80-99%) over a nucleic acid or amino acid sequence of identical size or when compared to an

25 aligned sequence in which the alignment is done by a computer homology program known in the art, or whose encoding nucleic acid is capable of hybridizing to the complement of a sequence encoding the aforementioned proteins under stringent, moderately stringent, or low stringent conditions. See e.g. Ausubel, *et al.*, CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, New York, NY, 1993, and

30 below. An exemplary program is the Gap program (Wisconsin Sequence Analysis Package, Version 8 for UNIX, Genetics Computer Group, University Research Park, Madison, WI) using the default settings, which uses the algorithm of Smith and Waterman (Adv. Appl. Math., 1981, 2: 482-489, which is incorporated herein by reference in its entirety). A "homologous nucleic acid sequence" or "homologous amino

acid sequence,” or variations thereof, refer to sequences characterized by a homology at the nucleotide level or amino acid level as discussed above. Homologous nucleotide sequences encode those sequences coding for isoforms of a CPP polypeptide. Isoforms can be expressed in different tissues of the same organism as a result of, for example, 5 alternative splicing of RNA. Alternatively, isoforms can be encoded by different genes. Homologous nucleotide sequences also include, but are not limited to, naturally occurring allelic variations and mutations of the nucleotide sequences set forth herein. Exemplary homologous nucleic acid sequences include the nucleic acid sequences of SEQ ID NO: 68, 70, 72 and 74. Homologous nucleic acid sequences include those 10 nucleic acid sequences that encode conservative amino acid substitutions (see below) in SEQ ID NO:2, SEQ ID NO:15 and SEQ ID NO:18 , as well as a polypeptide having CPP activity, e.g. substrate binding.

The nucleotide sequence determined from the cloning of the *Arabidopsis thaliana*, *Brassica napus* or *Glycine max* CPP gene allows for the generation of probes 15 and primers designed for use in identifying and/or cloning CPP homologues in other cell types, e.g., from other tissues, as well as CPP homologues from other plants. The probe/primer typically comprises a substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12, 25, 50, 100, 150, 200, 250, 300, 350 or 20 400 or more consecutive sense strand nucleotide sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17; or an anti-sense strand nucleotide sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17; or of a naturally occurring mutant of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17.

Probes based on the *Arabidopsis thaliana*, *Brassica napus* or *Glycine max* CPP 25 nucleotide sequence can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. In various embodiments, the probe further comprises a label group attached thereto, e.g., the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells or tissue which misexpress a CPP protein, such as 30 by measuring a level of a CPP-encoding nucleic acid in a sample of cells from a subject e.g., detecting CPP mRNA levels or determining whether a genomic CPP gene has been mutated or deleted.

A “polypeptide having a biologically active portion of CPP” refers to polypeptides exhibiting activity similar, but not necessarily identical to, an activity of a

polypeptide of the present invention, including mature forms, as measured in a particular biological assay, with or without dose dependency. A nucleic acid fragment encoding a "biologically active portion of CPP" can be prepared by isolating a portion of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 that encodes a polypeptide having a CPP 5 biological activity (biological activities of the CPP proteins are described below), expressing the encoded portion of CPP protein (*e.g.*, by recombinant expression *in vitro*) and assessing the activity of the encoded portion of CPP. In another embodiment, a nucleic acid fragment encoding a biologically active portion of CPP includes one or more regions.

10

CPP Variants

The invention further encompasses nucleic acid molecules that differ from the nucleotide sequences shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 due to the degeneracy of the genetic code. These nucleic acids thus encode the same CPP 15 protein as that encoded by the nucleotide sequence shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17, *e.g.*, the polypeptide of SEQ ID NO: 2, SEQ ID NO:15, SEQ ID NO: 18. In another embodiment, an isolated nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO: 2, SEQ ID NO:15, SEQ ID NO: 18.

20 In addition to the *Arabidopsis thaliana*, *Brassica napus* or *Glycine max* CPP nucleotide sequence shown in SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17, it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of CPP may exist within a population (*e.g.*, the plant). Such genetic polymorphism in the CPP gene may exist among individuals within 25 a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules comprising an open reading frame encoding a CPP protein, preferably a plant CPP protein. Such natural allelic variations can typically result in 1-5% variance in the nucleotide sequence of the CPP gene. Any and all such nucleotide variations and resulting amino acid polymorphisms in CPP that 30 are the result of natural allelic variation and that do not alter the functional activity of CPP are intended to be within the scope of the invention.

Moreover, nucleic acid molecules encoding CPP proteins from other species, and thus that have a nucleotide sequence that differs from the sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 are intended to be within the scope of the invention.

Nucleic acid molecules corresponding to natural allelic variants and homologues of the CPP cDNAs of the invention can be isolated based on their homology to the *Arabidopsis thaliana*, *Brassica napus* or *Glycine max* CPP nucleic acids disclosed herein using the cDNAs, or a portion thereof, as a hybridization probe according to standard
5 hybridization techniques under stringent hybridization conditions.

Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 6 nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17. In another embodiment, the nucleic acid is at least 10, 25,
10 50, 100, 250, 500 or 750 nucleotides in length. In another embodiment, an isolated nucleic acid molecule of the invention hybridizes to the coding region. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% homologous to each other typically remain hybridized to each other.

15 Homologs (*i.e.*, nucleic acids encoding CPP proteins derived from species other than *Arabidopsis thaliana*, *Brassica napus* or *Glycine max*) or other related sequences (*e.g.*, paralogs) can be obtained by low, moderate or high stringency hybridization with all or a portion of the particular sequence as a probe using methods well known in the art for nucleic acid hybridization and cloning.

20 As used herein, the phrase "stringent hybridization conditions" refers to conditions under which a probe, primer or oligonucleotide will hybridize to its target sequence, but to no other sequences. Stringent conditions are sequence-dependent and will be different depending upon circumstances. Longer sequences hybridize specifically at higher temperatures than shorter sequences. Generally, stringent
25 conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength, pH and nucleic acid concentration) at which 50% of the probes complementary to the target sequence hybridize to the target sequence at equilibrium. Since the target sequences are generally present at excess, at T_m , 50% of the probes are
30 occupied at equilibrium. Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes, primers or oligonucleotides (*e.g.*, 10 nt to 50 nt) and at least about 60°C for

longer probes, primers and oligonucleotides. Stringent conditions may also be achieved with the addition of destabilizing agents, such as formamide.

Stringent conditions are known to those skilled in the art and can be found in CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, N.Y. (1989), 5 6.3.1-6.3.6. Preferably, the conditions are such that sequences at least about 65%, 70%, 75%, 85%, 90%, 95%, 98%, or 99% homologous to each other typically remain hybridized to each other. A non-limiting example of stringent hybridization conditions is hybridization in a high salt buffer comprising 6X SSC, 50 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.02% BSA, and 500 mg/ml denatured salmon sperm 10 DNA at 65°C. This hybridization is followed by one or more washes in 0.2X SSC, 0.01% BSA at 50°C. An isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to the sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 corresponds to a naturally occurring nucleic acid molecule. As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a 15 nucleotide sequence that occurs in nature (*e.g.*, encodes a natural protein).

In a second embodiment, a nucleic acid sequence that is hybridizable to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17, or fragments, analogs or derivatives thereof, under conditions of moderate stringency is provided. A non-limiting example of moderate stringency 20 hybridization conditions are hybridization in 6X SSC, 5X Denhardt's solution, 0.5% SDS and 100 mg/ml denatured salmon sperm DNA at 55°C, followed by one or more washes in 1X SSC, 0.1% SDS at 37°C. Other conditions of moderate stringency that may be used are well known in the art. See, *e.g.*, Ausubel *et al.* (eds.), 1993, CURRENT 25 PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, NY, and Kriegler, 1990, GENE TRANSFER AND EXPRESSION, A LABORATORY MANUAL, Stockton Press, NY.

In a third embodiment, a nucleic acid that is hybridizable to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO: 1 or SEQ ID NO:14, or SEQ ID NO: 17 or fragments, analogs or derivatives thereof, under conditions of low stringency, is provided. A non-limiting example of low stringency hybridization 30 conditions are hybridization in 35% formamide, 5X SSC, 50 mM Tris-HCl (pH 7.5), 5 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.2% BSA, 100 mg/ml denatured salmon sperm DNA, 10% (wt/vol) dextran sulfate at 40°C, followed by one or more washes in 2X SSC, 25 mM Tris-HCl (pH 7.4), 5 mM EDTA, and 0.1% SDS at 50°C. Other conditions of low stringency that may be used are well known in the art (*e.g.*, as employed for

cross-species hybridizations). See, e.g., Ausubel *et al.* (eds.), 1993, CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, NY, and Kriegler, 1990, GENE TRANSFER AND EXPRESSION, A LABORATORY MANUAL, Stockton Press, NY; Shilo and Weinberg, 1981, *Proc Natl Acad Sci USA* 78: 6789-6792.

5

Conservative mutations

In addition to naturally-occurring allelic variants of the CPP sequence that may exist in the population, the skilled artisan will further appreciate that changes can be introduced by mutation into the nucleotide sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO: 17, thereby leading to changes in the amino acid sequence of the encoded CPP protein, without altering the functional ability of the CPP protein. For example, nucleotide substitutions leading to amino acid substitutions at "non-essential" amino acid residues can be made in the sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO: 17. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of CPP without altering the biological activity, whereas an "essential" amino acid residue is required for biological activity. For example, amino acid residues that are conserved among the CPP proteins of the present invention, are predicted to be particularly unamenable to alteration.

Another aspect of the invention pertains to nucleic acid molecules encoding CPP proteins that contain changes in amino acid residues that are not essential for activity. Such CPP proteins differ in amino acid sequence from SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18, yet retain biological activity. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a protein, wherein the protein comprises an amino acid sequence at least about 75% homologous to the amino acid sequence of SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18. Preferably, the protein encoded by the nucleic acid is at least about 80% homologous to SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18 more preferably at least about 90%, 95%, 98%, and most preferably at least about 99% homologous to SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18.

An isolated nucleic acid molecule encoding a CPP protein homologous to the protein of SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18 can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of SEQ ID NO: 1 or SEQ ID NO:14, or SEQ ID NO:17 such that

one or more amino acid substitutions, additions or deletions are introduced into the encoded protein.

Mutations can be introduced into the nucleotide sequence of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (*e.g.*, lysine, arginine, histidine), acidic side chains (*e.g.*, aspartic acid, glutamic acid), uncharged polar side chains (*e.g.*, glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (*e.g.*, alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (*e.g.*, threonine, valine, isoleucine) and aromatic side chains (*e.g.*, tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in CPP is replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of a CPP coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for CPP biological activity to identify mutants that retain activity. Following mutagenesis of SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17 the encoded protein can be expressed by any recombinant technology known in the art and the activity of the protein can be determined.

In one embodiment, a mutant CPP protein can be assayed for (1) the ability to form protein:protein interactions with other CPP proteins, other cell-surface proteins, or biologically active portions thereof, (2) complex formation between a mutant CPP protein and a CPP receptor; (3) the ability of a mutant CPP protein to bind to an intracellular target protein or biologically active portion thereof; (*e.g.*, avidin proteins); (4) the ability to bind CPP protein; or (5) the ability to specifically bind an anti-CPP protein antibody.

30

Antisense CPP Nucleic Acids

Another aspect of the invention pertains to isolated antisense nucleic acid molecules that are hybridizable to or complementary to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO: 1 or SEQ ID NO:14 or SEQ ID

NO:17, or fragments, analogs or derivatives thereof. An "antisense" nucleic acid comprises a nucleotide sequence that is complementary to a "sense" nucleic acid encoding a protein, *e.g.*, complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. In specific aspects, antisense
5 nucleic acid molecules are provided that comprise a sequence complementary to at least about 10, 25, 50, 100, 250 or 500 nucleotides or an entire CPP coding strand, or to only a portion thereof. Nucleic acid molecules encoding fragments, homologs, derivatives and analogs of a CPP protein of SEQ ID NO: 2 or SEQ ID NO:15 or SEQ ID NO:18 or antisense nucleic acids complementary to a CPP nucleic acid sequence of SEQ ID NO: 1,
10 SEQ ID NO:14 or SEQ ID NO:17 are additionally provided. Exemplary CPP anti-sense nucleic acid include the nucleic acid sequences of SEQ ID NO:16, 19, and 20.

In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding CPP. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are
15 translated into amino acid residues (*e.g.*, the protein coding region of *Arabidopsis thaliana*, *Brassica napus* or *Glycine max* CPP corresponds to SEQ ID NO: 2 or SEQ ID NO:15 or SEQ ID NO:18). In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding CPP. The term "noncoding region" refers to 5' and 3' sequences which flank
20 the coding region that are not translated into amino acids (*i.e.*, also referred to as 5' and 3' untranslated regions).

Given the coding strand sequences encoding CPP disclosed herein (*e.g.*, SEQ ID NO: 1 or SEQ ID NO:14 or SEQ ID NO:17), antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick or Hoogsteen base pairing. The
25 antisense nucleic acid molecule can be complementary to the entire coding region of CPP mRNA, but more preferably is an oligonucleotide that is antisense to only a portion of the coding or noncoding region of CPP mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of CPP mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20,
30 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis or enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (*e.g.*, an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the

molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used.

Examples of modified nucleotides that can be used to generate the antisense nucleic acid include: 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 10 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 15 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of 20 an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are generated *in situ* such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding a CPP protein to thereby inhibit expression of the protein, *e.g.*, by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule that binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention includes direct injection at a tissue site. 25 Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, *e.g.*, by linking the antisense nucleic acid molecules to peptides or antibodies that bind to cell surface receptors or antigens. The antisense

nucleic acid molecules can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

5 In yet another embodiment, the antisense nucleic acid molecule of the invention is an α -anomeric nucleic acid molecule. An α -anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β -units, the strands run parallel to each other (Gaultier *et al.* (1987) *Nucleic Acids Res* 15: 6625-6641). The antisense nucleic acid molecule can also comprise a
10 2'-o-methylribonucleotide (Inoue *et al.* (1987) *Nucleic Acids Res* 15: 6131-6148) or a chimeric RNA -DNA analogue (Inoue *et al.* (1987) *FEBS Lett* 215: 327-330).

Such modifications include, by way of nonlimiting example, modified bases, and nucleic acids whose sugar phosphate backbones are modified or derivatized. These modifications are carried out at least in part to enhance the chemical stability of the
15 modified nucleic acid, such that they may be used, for example, as antisense binding nucleic acids in applications.

Double Stranded RNA Inhibition (RNAi) by Hairpin Nucleic Acids

Another aspect of the invention pertains to the use of post transcriptional gene silencing (PTGS) to repress gene expression. Double stranded RNA can initiate the sequence specific repression of gene expression in plants and animals. Double stranded RNA is processed to short duplex oligomers of 21-23 nucleotides in length. These small interfering RNA's suppress the expression of endogenous and heterologous genes in a sequence specific manner (Fire *et al.* *Nature* 391:806-811, Carthew, *Curr. Opin. in Cell Biol.*, 13:244-248, Elbashir *et al.*, *Nature* 411:494-498). A RNAi suppressing construct can be designed in a number of ways, for example, transcription of a inverted repeat which can form a long hair pin molecule, inverted repeats separated by a spacer sequence that could be an unrelated sequence such as GUS or an intron sequence. Transcription of sense and antisense strands by opposing promoters or cotranscription of sense and
30 antisense genes.

CPP Ribozymes and PNA moieties

In still another embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity that are capable of cleaving a single-stranded nucleic acid, such as a mRNA, to which they have a complementary region. Thus, ribozymes (*e.g.*, hammerhead ribozymes (described in 5 Haselhoff and Gerlach (1988) *Nature* 334:585-591)) can be used to catalytically cleave CPP mRNA transcripts to thereby inhibit translation of CPP mRNA. A ribozyme having specificity for a CPP-encoding nucleic acid can be designed based upon the nucleotide sequence of a CPP DNA disclosed herein (*i.e.*, SEQ ID NO: 1, SEQ ID NO:14 or SEQ ID NO:17). For example, a derivative of a Tetrahymena L-19 IVS RNA can be 10 constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in a CPP-encoding mRNA. See, *e.g.*, Cech *et al.* U.S. Pat. No. 4,987,071; and Cech *et al.* U.S. Pat. No. 5,116,742. Alternatively, CPP mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, *e.g.*, Bartel *et al.*, (1993) *Science* 261:1411-1418.

15 Alternatively, CPP gene expression can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the CPP (*e.g.*, the CPP promoter and/or enhancers) to form triple helical structures that prevent transcription of the CPP gene in target cells. See generally, Helene. (1991) *Anticancer Drug Des.* 6: 569-84; Helene. *et al.* (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and Maher (1992) *Bioassays* 14: 20 807-15.

In various embodiments, the nucleic acids of CPP can be modified at the base moiety, sugar moiety or phosphate backbone to improve, *e.g.*, the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acids can be modified to generate peptide nucleic acids (see Hyrup *et al.* (1996) 25 *Bioorg Med Chem* 4: 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, *e.g.*, DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of 30 PNAs oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup *et al.* (1996) above; Perry-O'Keefe *et al.* (1996) *PNAS* 93: 14670-675.

PNAs of CPP can be used in therapeutic and diagnostic applications. For example, PNAs can be used as antisense or antigene agents for sequence-specific

modulation of gene expression by, *e.g.*, inducing transcription or translation arrest or inhibiting replication. PNAs of CPP can also be used, *e.g.*, in the analysis of single base pair mutations in a gene by, *e.g.*, PNA directed PCR clamping; as artificial restriction enzymes when used in combination with other enzymes, *e.g.*, S1 nucleases (Hyrup B. 5 (1996) above); or as probes or primers for DNA sequence and hybridization (Hyrup *et al.* (1996), above; Perry-O'Keefe (1996), above).

In another embodiment, PNAs of CPP can be modified, *e.g.*, to enhance their stability or cellular uptake, by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug 10 delivery known in the art. For example, PNA-DNA chimeras of CPP can be generated that may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, *e.g.*, RNase H and DNA polymerases, to interact with the DNA portion while the PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms 15 of base stacking, number of bonds between the nucleobases, and orientation (Hyrup (1996) above). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup (1996) above and Finn *et al.* (1996) *Nucl Acids Res* 24: 3357-63. For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite 20 coupling chemistry, and modified nucleoside analogs, *e.g.*, 5'-(4-methoxytrityl) amino-5'-deoxy-thymidine phosphoramidite, can be used between the PNA and the 5' end of DNA (Mag *et al.* (1989) *Nucl Acid Res* 17: 5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn *et al.* (1996) above). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment. See, Petersen *et al.* 25 (1975) *Bioorg Med Chem Lett* 5: 1119-1124.

CPP Polypeptides

A CPP polypeptide of the invention includes the protein whose sequence is provided in SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18. The invention also includes a mutant or variant protein any of whose residues may be changed from the 30 corresponding residue shown in SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18 while still encoding a protein that maintains its CPP-like activities and physiological functions, or a functional fragment thereof. In some embodiments, up to 20% or more of the residues may be so changed in the mutant or variant protein. In some embodiments, the CPP polypeptide according to the invention is a mature polypeptide.

In general, a CPP -like variant that preserves CPP-like function includes any variant in which residues at a particular position in the sequence have been substituted by other amino acids, and further include the possibility of inserting an additional residue or residues between two residues of the parent protein as well as the possibility of deleting 5 one or more residues from the parent sequence. Any amino acid substitution, insertion, or deletion is encompassed by the invention. In favorable circumstances, the substitution is a conservative substitution as defined above.

One aspect of the invention pertains to isolated CPP proteins, and biologically active portions thereof, or derivatives, fragments, analogs or homologs thereof. Also 10 provided are polypeptide fragments suitable for use as immunogens to raise anti-CPP antibodies. In one embodiment, native CPP proteins can be isolated from cells or tissue sources by an appropriate purification scheme using standard protein purification techniques. In another embodiment, CPP proteins are produced by recombinant DNA techniques. Alternative to recombinant expression, a CPP protein or polypeptide can be 15 synthesized chemically using standard peptide synthesis techniques.

An "isolated" or "purified" protein or biologically active portion thereof is substantially free of cellular material or other contaminating proteins from the cell or tissue source from which the CPP protein is derived, or substantially free from chemical precursors or other chemicals when chemically synthesized. The language "substantially 20 free of cellular material" includes preparations of CPP protein in which the protein is separated from cellular components of the cells from which it is isolated or recombinantly produced. In one embodiment, the language "substantially free of cellular material" includes preparations of CPP protein having less than about 30% (by dry weight) of non-CPP protein (also referred to herein as a "contaminating protein"), more 25 preferably less than about 20% of non-CPP protein, still more preferably less than about 10% of non-CPP protein, and most preferably less than about 5% non-CPP protein. When the CPP protein or biologically active portion thereof is recombinantly produced, it is also preferably substantially free of culture medium, *i.e.*, culture medium represents less than about 20%, more preferably less than about 10%, and most preferably less than 30% about 5% of the volume of the protein preparation.

The language "substantially free of chemical precursors or other chemicals" includes preparations of CPP protein in which the protein is separated from chemical precursors or other chemicals that are involved in the synthesis of the protein. In one embodiment, the language "substantially free of chemical precursors or other chemicals"

includes preparations of CPP protein having less than about 30% (by dry weight) of chemical precursors or non-CPP chemicals, more preferably less than about 20% chemical precursors or non-CPP chemicals, still more preferably less than about 10% chemical precursors or non-CPP chemicals, and most preferably less than about 5%
5 chemical precursors or non-CPP chemicals.

Biologically active portions of a CPP protein include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequence of the CPP protein, *e.g.*, the amino acid sequence shown in SEQ ID NO: 2 that include fewer amino acids than the full length CPP proteins, and exhibit at least one activity of a
10 CPP protein, *e.g.* substrate binding. Typically, biologically active portions comprise a domain or motif with at least one activity of the CPP protein. A biologically active portion of a CPP protein can be a polypeptide which is, for example, 10, 25, 50, 100 or more amino acids in length.

A biologically active portion of a CPP protein of the present invention may
15 contain at least one of the above-identified domains conserved between the CPP proteins,
e.g.. Moreover, other biologically active portions, in which other regions of the protein are deleted, can be prepared by recombinant techniques and evaluated for one or more of the functional activities of a native CPP protein.

A biologically active portion or a CPP protein can be the N-terminal domain of
20 the CPP polypeptide. Alternatively, a biologically active portion or a CPP protein can be the C-terminal domain of the CPP polypeptide. Preferably, the biologically active portion comprises at least 75 amino acids of the C-terminal domain. More preferably, the biologically active portion comprises at least 25 amino acids of the C-terminal domain. Most preferably, the biologically active portion comprises at least 10 amino
25 acids of the C-terminal.

In an embodiment, the CPP protein has an amino acid sequence shown in SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18. In other embodiments, the CPP protein is substantially homologous to SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18 and retains the functional activity of the protein of SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID
30 NO:18, yet differs in amino acid sequence due to natural allelic variation or mutagenesis, as described in detail below. Accordingly, in another embodiment, the CPP protein is a protein that comprises an amino acid sequence at least about 45% homologous to the amino acid sequence of S SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18 and retains

the functional activity of the CPP proteins of SEQ ID NO: 2, SEQ ID NO:15 or SEQ ID NO:18.

Exemplary homologous CPP polypeptides include for example the polypeptide sequences of SEQ ID NO: 69, 71, 73 and 75.

5 **Determining homology between two or more sequence**

To determine the percent homology of two amino acid sequences or of two nucleic acids, the sequences are aligned for optimal comparison purposes (*e.g.*, gaps can be introduced in either of the sequences being compared for optimal alignment between the sequences). The amino acid residues or nucleotides at corresponding amino acid 10 positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are homologous at that position (*i.e.*, as used herein amino acid or nucleic acid "homology" is equivalent to amino acid or nucleic acid "identity").

15 The nucleic acid sequence homology may be determined as the degree of identity between two sequences. The homology may be determined using computer programs known in the art, such as GAP software provided in the GCG program package. See, *Needleman and Wunsch 1970 J Mol Biol 48: 443-453*. Using GCG GAP software with the following settings for nucleic acid sequence comparison: GAP creation penalty of 5.0 20 and GAP extension penalty of 0.3, the coding region of the analogous nucleic acid sequences referred to above exhibits a degree of identity preferably of at least 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, with the CDS (encoding) part of the DNA sequence shown in SEQ ID NO:1 or SEQ ID NO:14 or SEQ ID NO:17.

The term "sequence identity" refers to the degree to which two polynucleotide or 25 polypeptide sequences are identical on a residue-by-residue basis over a particular region of comparison. The term "percentage of sequence identity" is calculated by comparing two optimally aligned sequences over that region of comparison, determining the number of positions at which the identical nucleic acid base (*e.g.*, A, T, C, G, U, or I, in the case of nucleic acids) occurs in both sequences to yield the number of matched positions, 30 dividing the number of matched positions by the total number of positions in the region of comparison (*i.e.*, the window size), and multiplying the result by 100 to yield the percentage of sequence identity. The term "substantial identity" as used herein denotes a characteristic of a polynucleotide sequence, wherein the polynucleotide comprises a sequence that has at least 80 percent sequence identity, preferably at least 85 percent

identity and often 90 to 95 percent sequence identity, more usually at least 99 percent sequence identity as compared to a reference sequence over a comparison region. The term "percentage of positive residues" is calculated by comparing two optimally aligned sequences over that region of comparison, determining the number of positions at which 5 the identical and conservative amino acid substitutions, as defined above, occur in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the region of comparison (*i.e.*, the window size), and multiplying the result by 100 to yield the percentage of positive residues.

10 Chimeric and fusion proteins

The invention also provides CPP chimeric or fusion proteins. As used herein, a CPP "chimeric protein" or "fusion protein" comprises a CPP polypeptide operatively linked to a non-CPP polypeptide. An "CPP polypeptide" refers to a polypeptide having an amino acid sequence corresponding to CPP, whereas a "non-CPP polypeptide" refers 15 to a polypeptide having an amino acid sequence corresponding to a protein that is not substantially homologous to the CPP protein, *e.g.*, a protein that is different from the CPP protein and that is derived from the same or a different organism. Within a CPP fusion protein the CPP polypeptide can correspond to all or a portion of a CPP protein. In one embodiment, a CPP fusion protein comprises at least one biologically active 20 portion of a CPP protein. In another embodiment, a CPP fusion protein comprises at least two biologically active portions of a CPP protein. Within the fusion protein, the term "operatively linked" is intended to indicate that the CPP polypeptide and the non-CPP polypeptide are fused in-frame to each other. The non-CPP polypeptide can be fused to the N-terminus or C-terminus of the CPP polypeptide.

25 A CPP chimeric or fusion protein of the invention can be produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, *e.g.*, by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive 30 ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers that give rise to complementary overhangs between two consecutive gene fragments that can

subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, Ausubel et al. (eds.) CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (*e.g.*, a GST polypeptide). A CPP-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the CPP protein.

CPP agonists and antagonists

The present invention also pertains to variants of the CPP proteins that function as either CPP agonists (mimetics) or as CPP antagonists. Variants of the CPP protein can be generated by mutagenesis, *e.g.*, discrete point mutation or truncation of the CPP protein. An agonist of the CPP protein can retain substantially the same, or a subset of, the biological activities of the naturally occurring form of the CPP protein. An antagonist of the CPP protein can inhibit one or more of the activities of the naturally occurring form of the CPP protein by, for example, competitively binding to a downstream or upstream member of a cellular signaling cascade which includes the CPP protein. Thus, specific biological effects can be elicited by treatment with a variant of limited function.

Variants of the CPP protein that function as either CPP agonists (mimetics) or as CPP antagonists can be identified by screening combinatorial libraries of mutants, *e.g.*, truncation mutants, of the CPP protein for CPP protein agonist or antagonist activity. In one embodiment, a variegated library of CPP variants is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene library. A variegated library of CPP variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential CPP sequences is expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (*e.g.*, for phage display) containing the set of CPP sequences therein. There are a variety of methods which can be used to produce libraries of potential CPP variants from a degenerate oligonucleotide sequence.

Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding the desired set of potential CPP sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (see, *e.g.*, Narang (1983)

Tetrahedron 39:3; Itakura *et al.* (1984) *Annu Rev Biochem* 53:323; Itakura *et al.* (1984) *Science* 198:1056; Ike *et al.* (1983) *Nucl Acid Res* 11:477.

Polyptide libraries

5 In addition, libraries of fragments of the CPP protein coding sequence can be used to generate a variegated population of CPP fragments for screening and subsequent selection of variants of a CPP protein. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of a CPP coding sequence with a nuclease under conditions wherein nicking occurs only about once per
10 molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA that can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclelease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal and internal
15 fragments of various sizes of the CPP protein.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of CPP proteins. The most widely used techniques, which are amenable to high throughput analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates isolation of the vector encoding the gene whose product was
20 detected. Recursive ensemble mutagenesis (REM), a new technique that enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify CPP variants (Arkin and Yourvan (1992) PNAS 89:7811-7815; Delgrave *et al.* (1993) Protein Engineering 6:327-331).

30 **CPP Antibodies**

CPP polypeptides, including chimeric polypeptides, or derivatives, fragments, analogs or homologs thereof, may be utilized as immunogens to generate antibodies that immunospecifically-bind these peptide components. Such antibodies include, *e.g.*, polyclonal, monoclonal, chimeric, single chain, Fab fragments and a Fab expression

library. In a specific embodiment, fragments of the CPP polypeptides are used as immunogens for antibody production. Various procedures known within the art may be used for the production of polyclonal or monoclonal antibodies to a CPP polypeptides, or derivative, fragment, analog or homolog thereof.

- 5 For the production of polyclonal antibodies, various host animals may be immunized by injection with the native peptide, or a synthetic variant thereof, or a derivative of the foregoing. Various adjuvants may be used to increase the immunological response and include, but are not limited to, Freund's (complete and incomplete), mineral gels (e.g., aluminum hydroxide), surface active substances (e.g., lysolecithin, pluronic polyols, 10 polyanions, peptides, oil emulsions, dinitrophenol, etc.) and human adjuvants such as *Bacille Calmette-Guerin* and *Corynebacterium parvum*.

For preparation of monoclonal antibodies directed towards a CPP polypeptides, or derivatives, fragments, analogs or homologs thereof, any technique that provides for the production of antibody molecules by continuous cell line culture may be utilized.

- 15 Such techniques include, but are not limited to, the hybridoma technique (see, Kohler and Milstein, 1975. *Nature* 256: 495-497); the trioma technique; the human B-cell hybridoma technique (see, Kozbor, et al., 1983. *Immunol Today* 4: 72) and the EBV hybridoma technique to produce human monoclonal antibodies (see, Cole, et al., 1985. In: *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96). Human 20 monoclonal antibodies may be utilized in the practice of the present invention and may be produced by the use of human hybridomas (see, Cote, et al., 1983. *Proc Natl Acad Sci USA* 80: 2026-2030) or by transforming human B-cells with Epstein Barr Virus *in vitro* (see, Cole, et al., 1985. In: *Monoclonal Antibodies and Cancer Therapy* (Alan R. Liss, Inc., pp. 77-96).

- 25 According to the invention, techniques can be adapted for the production of single-chain antibodies specific to a CPP polypeptides (see, e.g., U.S. Patent No. 4,946,778). In addition, methodologies can be adapted for the construction of Fab expression libraries (see, e.g., Huse, et al., 1989. *Science* 246: 1275-1281) to allow rapid and effective identification of monoclonal Fab fragments with the desired specificity for 30 a CPP polypeptides or derivatives, fragments, analogs or homologs thereof. Antibody fragments that contain the idiotypes to a CPP polypeptides may be produced by techniques known in the art including, e.g., (i) an F(ab')₂ fragment produced by pepsin digestion of an antibody molecule; (ii) an Fab fragment generated by reducing the

disulfide bridges of an F(ab')₂ fragment; (iii) an Fab fragment generated by the treatment of the antibody molecule with papain and a reducing agent and (iv) Fv fragments.

In one embodiment, methodologies for the screening of antibodies that possess the desired specificity include, but are not limited to, enzyme-linked immunosorbent assay

- 5 (ELISA) and other immunologically-mediated techniques known within the art. In a specific embodiment, selection of antibodies that are specific to a particular domain of a CPP polypeptides is facilitated by generation of hybridomas that bind to the fragment of a CPP polypeptides possessing such a domain. Antibodies that are specific for a domain within a CPP polypeptides, or derivative, fragments, analogs or homologs thereof, are
10 also provided herein. The anti-CPP polypeptide antibodies may be used in methods known within the art relating to the localization and/or quantitation of a CPP polypeptide(*e.g.*, for use in measuring levels of the peptide within appropriate physiological samples, for use in diagnostic methods, for use in imaging the peptide, and the like).

15

CPP Recombinant Expression Vectors and Host Cells

Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding a CPP protein, or derivatives, fragments, analogs or homologs thereof. As used herein, the term "vector" refers to a nucleic acid molecule
20 capable of transporting another nucleic acid to which it has been linked. Exemplary expression vector constructs include for example the constructs of SEQ ID NO: 4,, 5, 36, 37, 39, 40, 441, 42, 44, 45, 47, 48, 50 , 51 and 53. Additional exemplary expression vector constructs include constructs comprising CPP anti-sense nucleic acid such as SEQ ID NO: 38. 43., 46, 49, 52. One type of vector is a "plasmid", which refers to a circular
25 double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (*e.g.*, bacterial vectors having a bacterial origin of replication). Other vectors are integrated into the genome of a host cell upon introduction into the host
30 cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively-linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the

plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors or plant transformation vectors, binary or otherwise, which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, that is operatively-linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably-linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner that allows for expression of the nucleotide sequence (*e.g.*, in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell).

The term "regulatory sequence" is intended to includes promoters, enhancers and other expression control elements (*e.g.*, polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel, GENE EXPRESSION TECHNOLOGY: METHODS IN ENZYMOLOGY 185, Academic Press, San Diego, Calif. (1990). Regulatory sequences include those that direct constitutive expression of a nucleotide sequence in many types of host cell and those that direct expression of the nucleotide sequence only in certain host cells (*e.g.*, tissue-specific regulatory sequences). Examples of suitable promoters include for example constitutive promoters, ABA inducible promoters, tissue specific promoters or guard cell specific promoters. It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, etc. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (*e.g.*, CPP proteins, mutant forms of CPP proteins, fusion proteins, etc.).

The recombinant expression vectors of the invention can be designed for expression of CPP proteins in prokaryotic or eukaryotic cells. For example, CPP proteins can be expressed in bacterial cells such as *Escherichia coli*, insect cells (using baculovirus expression vectors) yeast cells, plant cells or mammalian cells. Suitable host cells are discussed further in Goeddel, GENE EXPRESSION TECHNOLOGY: METHODS IN ENZYMOLOGY 185, Academic Press, San Diego, Calif. (1990). Alternatively, the

recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

- Expression of proteins in prokaryotes is most often carried out in *Escherichia coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: (i) to increase expression of recombinant protein; (ii) to increase the solubility of the recombinant protein; and (iii) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification.
- Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase. Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith and Johnson, 1988. *Gene* 67: 31-40), pMAL (New England Biolabs, Beverly, Mass.) and pRIT5 (Pharmacia, Piscataway, N.J.) that fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amrann *et al.*, (1988) *Gene* 69:301-315) and pET 11d (Studier *et al.*, GENE EXPRESSION TECHNOLOGY: METHODS IN ENZYMOLOGY 185, Academic Press, San Diego, Calif. (1990) 60-89).

One strategy to maximize recombinant protein expression in *E. coli* is to express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein. See, e.g., Gottesman, GENE EXPRESSION TECHNOLOGY: METHODS IN ENZYMOLOGY 185, Academic Press, San Diego, Calif. (1990) 119-128. Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *E. coli* (see, e.g., Wada, *et al.*, 1992. *Nucl. Acids Res.* 20: 2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the CPP expression vector is a yeast expression vector. Examples of vectors for expression in yeast *Saccharomyces cerevisiae* include pYEpSec1 (Baldari, *et al.*, 1987. *EMBO J.* 6: 229-234), pMFA (Kurjan and Herskowitz, 1982. *Cell*

30: 933-943), pJRY88 (Schultz *et al.*, 1987. *Gene* 54: 113-123), pYES2 (Invitrogen Corporation, San Diego, Calif.), and picZ (InVitrogen Corp, San Diego, Calif.). Alternatively, CPP can be expressed in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (*e.g.*, SF9

5 cells) include the pAc series (Smith, *et al.*, 1983. *Mol. Cell. Biol.* 3: 2156-2165) and the pVL series (Lucklow and Summers, 1989. *Virology* 170: 31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors include pCDM8 (Seed, 1987. *Nature* 329: 840) and pMT2PC (Kaufman, *et al.*, 1987.

10 *EMBO J.* 6: 187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, adenovirus 2, cytomegalovirus, and simian virus 40. For other suitable expression systems for both prokaryotic and eukaryotic cells see, *e.g.*, Chapters 16 and 17 of Sambrook, *et al.*, MOLECULAR CLONING: A LABORATORY

15 MANUAL. 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989.

In yet another embodiment, a nucleic acid of the invention is expressed in plants cells using a plant expression vector. Examples of plant expression vectors systems include tumor inducing (Ti) plasmid or portion thereof found in *Agrobacterium*, 20 cauliflower mosaic virus (CaMV) DNA and vectors such as pBI121 .

For expression in plants, the recombinant expression cassette will contain in addition to the CPP nucleic acids, a plant promoter region, a transcription initiation site (if the coding sequence to transcribed lacks one), and a transcription termination/polyadenylation sequence. The termination/polyadenylation region may be obtained from the same gene as the promoter sequence or may be obtained from different genes. Unique restriction enzyme sites at the 5' and 3' ends of the cassette are typically included to allow for easy insertion into a pre-existing vector.

Examples of suitable promoters include promoters from plant viruses such as the 35S promoter from cauliflower mosaic virus (CaMV). Odell, *et al.*, *Nature*, 313: 810-812 (1985). and promoters from genes such as rice actin (McElroy, *et al.*, *Plant Cell*, 163-171 (1990)); ubiquitin (Christensen, *et al.*, *Plant Mol. Biol.*, 12: 619-632 (1992); and Christensen, *et al.*, *Plant Mol. Biol.*, 18: 675-689 (1992)); pEMU (Last, *et al.*, *Theor. Appl. Genet.*, 81: 581-588 (1991)); MAS (Velten, *et al.*, *EMBO J.*, 3: 2723-2730 (1984)); maize H3 histone (Lepetit, *et al.*, *Mol. Gen. Genet.*, 231: 276-285 (1992); and

Atanassvoa, et al., Plant Journal, 2(3): 291-300 (1992)), the 5'- or 3'-promoter derived from T-DNA of *Agrobacterium tumefaciens*, the Smas promoter, the cinnamyl alcohol dehydrogenase promoter (U.S. Pat. No. 5,683,439), the Nos promoter, the rubisco promoter, the GRP1-8 promoter, ALS promoter, (WO 96/30530), a synthetic promoter, such as, Rsyn7, SCP and UCP promoters, ribulose-1,3-diphosphate carboxylase, fruit-specific promoters, heat shock promoters, seed-specific promoters and other transcription initiation regions from various plant genes, for example, include the various opine initiation regions, such as for example, octopine, mannopine, and nopaline.

Additional regulatory elements that may be connected to a CPP encoding nucleic acid sequence for expression in plant cells include terminators, polyadenylation sequences, and nucleic acid sequences encoding signal peptides that permit localization within a plant cell or secretion of the protein from the cell. Such regulatory elements and methods for adding or exchanging these elements with the regulatory elements CPP gene are known, and include, but are not limited to, 3' termination and/or polyadenylation regions such as those of the *Agrobacterium tumefaciens* nopaline synthase (nos) gene (Bevan, et al., Nucl. Acids Res., 12: 369-385 (1983)); the potato proteinase inhibitor II (PINII) gene (Keil, et al., Nucl. Acids Res., 14: 5641-5650 (1986) and hereby incorporated by reference); and An,, et al., Plant Cell, 1: 115-122 (1989)); and the CaMV 19S gene (Mogen, et al., Plant Cell, 2: 1261-1272 (1990)).

Plant signal sequences, including, but not limited to, signal-peptide encoding DNA/RNA sequences which target proteins to the extracellular matrix of the plant cell (Dratewka-Kos, et al., J. Biol. Chem., 264: 4896-4900 (1989)) and the *Nicotiana plumbaginifolia* extension gene (DeLoose, et al., Gene, 99: 95-100 (1991)), or signal peptides which target proteins to the vacuole like the sweet potato sporamin gene (Matsuka, et al., Proc. Nat'l Acad. Sci. (USA), 88: 834 (1991)) and the barley lectin gene (Wilkins, et al., Plant Cell, 2: 301-313 (1990)), or signals which cause proteins to be secreted such as that of PR1b (Lind, et al., Plant Mol. Biol., 18: 47-53 (1992)), or those which target proteins to the plastids such as that of rapeseed enoyl-ACP reductase (Verwaert, et al., Plant Mol. Biol., 26: 189-202 (1994)) are useful in the invention.

In another embodiment, the recombinant expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (e.g., tissue-specific regulatory elements are used to express the nucleic acid). Tissue-specific regulatory elements are known in the art. Especially useful in connection with the nucleic acids of

the present invention are expression systems which are operable in plants. These include systems which are under control of a tissue-specific promoter, as well as those which involve promoters that are operable in all plant tissues.

Organ-specific promoters are also well known. For example, the patatin class I promoter is transcriptionally activated only in the potato tuber and can be used to target gene expression in the tuber (Bevan, M., 1986, *Nucleic Acids Research* 14:4625-4636). Another potato-specific promoter is the granule-bound starch synthase (GBSS) promoter (Visser, R.G.R., et al., 1991, *Plant Molecular Biology* 17:691-699). Other organ-specific promoters appropriate for a desired target organ can be isolated using known procedures. These control sequences are generally associated with genes uniquely expressed in the desired organ. In a typical higher plant, each organ has thousands of mRNAs that are absent from other organ systems (reviewed in Goldberg, P., 1986, *Trans. R. Soc. London* B314:343).

For in situ production of the antisense mRNA of GST, those regions of the GST gene which are transcribed into GST mRNA, including the untranslated regions thereof, are inserted into the expression vector under control of the promoter system in a reverse orientation. The resulting transcribed mRNA is then complementary to that normally produced by the plant.

The resulting expression system or cassette is ligated into or otherwise constructed to be included in a recombinant vector which is appropriate for plant transformation. The vector may also contain a selectable marker gene by which transformed plant cells can be identified in culture. Usually, the marker gene will encode antibiotic resistance. These markers include resistance to G418, hygromycin, bleomycin, kanamycin, and gentamicin. After transforming the plant cells, those cells having the vector will be identified by their ability to grow on a medium containing the particular antibiotic. Replication sequences, of bacterial or viral origin, are generally also included to allow the vector to be cloned in a bacterial or phage host, preferably a broad host range prokaryotic origin of replication is included. A selectable marker for bacteria should also be included to allow selection of bacterial cells bearing the desired construct. Suitable prokaryotic selectable markers also include resistance to antibiotics such as kanamycin or tetracycline.

Other DNA sequences encoding additional functions may also be present in the vector, as is known in the art. For instance, in the case of *Agrobacterium*

transformations, T-DNA sequences will also be included for subsequent transfer to plant chromosomes.

- Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but also to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.
- Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and "transfection" are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (*e.g.*, DNA) into a host cell.

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (*i.e.*, express) a polypeptide of the invention encoded in an open reading frame of a polynucleotide of the invention. Accordingly, the invention further provides methods for producing a polypeptide using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding a polypeptide of the invention has been introduced) in a suitable medium such that the polypeptide is produced. In another embodiment, the method further comprises isolating the polypeptide from the medium or the host cell.

A number of types of cells may act as suitable host cells for expression of a polypeptide encoded by an open reading frame in a polynucleotide of the invention.

Plant host cells include, for example, plant cells that could function as suitable hosts for the expression of a polynucleotide of the invention include epidermal cells, mesophyll and other ground tissues, and vascular tissues in leaves, stems, floral organs, and roots from a variety of plant species, such as *Arabidopsis thaliana*, *Nicotiana tabacum*, *Brassica napus*, *Zea mays*, *Oryza sativa*, *Gossypium hirsutum* and *Glycine max*.

Alternatively, it may be possible to produce a polypeptide in lower eukaryotes such as yeast or in prokaryotes such as bacteria. Potentially suitable yeast strains include *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Kluyveromyces* strains, *Candida*, or any yeast strain capable of expressing heterologous proteins. Potentially suitable bacterial strains include *Escherichia coli*, *Bacillus subtilis*, *Salmonella*

typhimurium, or any bacterial strain capable of expressing heterologous polypeptides. If the polypeptide is made in yeast or bacteria, it may be necessary to modify the polypeptide produced therein, for example by phosphorylation or glycosylation of the appropriate sites, in order to obtain a functional polypeptide, if the polypeptide is of sufficient length and conformation to have activity. Such covalent attachments may be accomplished using known chemical or enzymatic methods.

A polypeptide may be prepared by culturing transformed host cells under culture conditions suitable to express the recombinant protein. The resulting expressed polypeptide or protein may then be purified from such culture (e.g., from culture medium or cell extracts) using known purification processes, such as gel filtration and ion exchange chromatography. The purification of the polypeptide or protein may also include an affinity column containing agents which will bind to the protein; one or more column steps over such affinity resins as concanavalin A-agarose, heparin-toyopearl® or Cibacrom blue 3GA Sepharose®; one or more steps involving hydrophobic interaction chromatography using such resins as phenyl ether, butyl ether, or propyl ether; or immunoaffinity chromatography.

Alternatively, a polypeptide or protein may also be expressed in a form which will facilitate purification. For example, it may be expressed as a fusion protein containing a six-residue histidine tag. The histidine-tagged protein will then bind to a Ni-affinity column. After elution of all other proteins, the histidine-tagged protein can be eluted to achieve rapid and efficient purification. One or more reverse-phase high performance liquid chromatography (RP-HPLC) steps employing hydrophobic RP-HPLC media, e.g., silica gel having pendant methyl or other aliphatic groups, can be employed to further purify a polypeptide. Some or all of the foregoing purification steps, in various combinations, can also be employed to provide a substantially homogeneous isolated recombinant polypeptide. The protein or polypeptide thus purified is substantially free of other plant proteins or polypeptides and is defined in accordance with the present invention as "isolated."

Transformed Plants Cells and Transgenic Plants

The invention includes protoplast, plants cells, plant tissue and plants (e.g., monocots and dicots transformed with a CPP nucleic acid (*i.e.*, sense or antisense), a vector containing a CPP nucleic acid (*i.e.*, sense or antisense) or an expression vector containing a CPP nucleic acid (*i.e.*, sense or antisense). As used herein, "plant" is meant

to include not only a whole plant but also a portion thereof (*i.e.*, cells, and tissues, including for example, leaves, stems, shoots, roots, flowers, fruits and seeds).

The plant can be any plant type including, for example, species from the genera *Cucurbita*, *Rosa*, *Vitis*, *Juglans*, *Fragaria*, *Lotus*, *Medicago*, *Onobrychis*, *Trifolium*,
5 *Trigonella*, *Vigna*, *Citrus*, *Linum*, *Geranium*, *Manihot*, *Daucus*, *Arabidopsis*, *Brassica*,
Raphanus, *Sinapis*, *Atropa*, *Capsicum*, *Datura*, *Hyoscyamus*, *Lycopersicon*, *Nicotiana*,
Solanum, *Petunia*, *Digitalis*, *Majorana*, *Ciahorium*, *Helianthus*, *Lactuca*, *Bromus*,
10 *Asparagus*, *Antirrhinum*, *Heterocallis*, *Nemesis*, *Pelargonium*, *Panieum*, *Pennisetum*,
Ranunculus, *Senecio*, *Salpiglossis*, *Cucumis*, *Browalia*, *Glycine*, *Pisum*, *Phaseolus*,
15 *Lolium*, *Oryza*, *Zea*, *Avena*, *Hordeum*, *Secale*, *Triticum*, *Sorghum*, *Gossypium*, *Picea*,
Caco, and *Populus*.

In some aspects of the invention, the transformed plant is resistant to biotic and abiotic stresses, *e.g.*, chilling stress, salt stress, water stress (*e.g.*, drought), disease, grazing pests and wound healing. Additionally, the invention also includes a transgenic
15 plant that is resistant to pathogens such as for example fungi, bacteria, nematodes, viruses and parasitic weeds. Alternatively, the transgenic plant is resistant to herbicides or has delayed senescence. The transgenic plant has an increase in yield, productivity, biomass or ABA sensitivity. By resistant is meant the plant grows under stress conditions (*e.g.*, high salt, decreased water, low temperatures) or under conditions that
20 normally inhibit, to some degree, the growth of an untransformed plant. Methodologies to determine plant growth or response to stress include for example, height measurements, weight measurements, leaf area, ability to flower, water use, transpiration rates and yield.

The invention also includes cells, tissues, including for example, leaves, stems,
25 shoots, roots, flowers, fruits and seeds and the progeny derived from the transformed plant.

Numerous methods for introducing foreign genes into plants are known and can be used to insert a gene into a plant host, including biological and physical plant transformation protocols. See, for example, Miki et al., (1993) "Procedure for
30 Introducing Foreign DNA into Plants", In: Methods in Plant Molecular Biology and Biotechnology, Glick and Thompson, eds., CRC Press, Inc., Boca Raton, pages 67-88 and Andrew Bent in, Clough SJ and Bent AF, 1998. Floral dipping: a simplified method for *Agrobacterium*-mediated transformation of *Arabidopsis thaliana*.. The methods

chosen vary with the host plant, and include chemical transfection methods such as calcium phosphate, polyethylene glycol (PEG) transformation, microorganism-mediated gene transfer such as *Agrobacterium* (Horsch, et al., Science, 227: 1229-31 (1985)), electroporation, protoplast transformation, micro-injection, flower dipping and biotic bombardment.

Agrobacterium-mediated Transformation

The most widely utilized method for introducing an expression vector into plants is based on the natural transformation system of *Agrobacterium*. *A. tumefaciens* and *A. rhizogenes* are plant pathogenic soil bacteria which genetically transform plant cells. The Ti and Ri plasmids of *A. tumefaciens* and *A. rhizogenes*, respectfully, carry genes responsible for genetic transformation of plants. See, for example, Kado, Crit. Rev. Plant Sci., 10: 1-32 (1991). Descriptions of the *Agrobacterium* vector systems and methods for *Agrobacterium*-mediated gene transfer are provided in Gruber et al., supra; and Moloney, et al, Plant Cell Reports, 8: 238-242 (1989).

Transgenic *Arabidopsis* plants can be produced easily by the method of dipping flowering plants into an *Agrobacterium* culture, based on the method of Andrew Bent in, Clough SJ and Bent AF, 1998. Floral dipping: a simplified method for *Agrobacterium*-mediated transformation of *Arabidopsis thaliana*. Wild type plants are grown until the plant has both developing flowers and open flowers. The plant are inverted for 1 minute into a solution of *Agrobacterium* culture carrying the appropriate gene construct. Plants are then left horizontal in a tray and kept covered for two days to maintain humidity and then righted and bagged to continue growth and seed development. Mature seed is bulk harvested.

25 Direct Gene Transfer

A generally applicable method of plant transformation is microprojectile-mediated transformation, where DNA is carried on the surface of microprojectiles measuring about 1 to 4 mu.m. The expression vector is introduced into plant tissues with a biotic device that accelerates the microprojectiles to speeds of 300 to 600 m/s which is sufficient to penetrate the plant cell walls and membranes. (Sanford, et al., Part. Sci. Technol., 5: 27-37 (1987); Sanford, Trends Biotech, 6: 299-302 (1988); Sanford, Physiol. Plant, 79: 206-209 (1990); Klein, et al., Biotechnology, 10: 286-291 (1992)).

Another method for physical delivery of DNA to plants is sonication of target cells as described in Zang, et al., BioTechnology, 9: 996-996 (1991). Alternatively, liposome or spheroplast fusions have been used to introduce expression vectors into plants. See, for example, Deshayes, et al., EMBO J., 4: 2731-2737 (1985); and Christou, et al., Proc. 5 Nat'l. Acad. Sci. (USA), 84: 3962-3966 (1987). Direct uptake of DNA into protoplasts using CaCl₂ precipitation, polyvinyl alcohol or poly-L-ornithine have also been reported. See, for example, Hain, et al., Mol. Gen. Genet., 199: 161 (1985); and Draper, et al., Plant Cell Physiol., 23: 451-458 (1982).

10 Electroporation of protoplasts and whole cells and tissues has also been described. See, for example, Donn, et al., (1990) In: Abstracts of the VIIth Int'l. Congress on Plant Cell and Tissue Culture IAPTC, A2-38, page 53; D'Halluin et al., Plant Cell, 4: 1495-1505 (1992); and Spencer et al., Plant Mol. Biol., 24: 51-61 (1994).

Particle Wounding/*Agrobacterium* Delivery

15 Another useful basic transformation protocol involves a combination of wounding by particle bombardment, followed by use of *Agrobacterium* for DNA delivery, as described by Bidney, et al., Plant Mol. Biol., 18: 301-31 (1992). Useful plasmids for plant transformation include Bin 19. See Bevan, Nucleic Acids Research, 12: 8711-8721 (1984), and hereby incorporated by reference.

20 In general, the intact meristem transformation method involves imbibing seed for 24 hours in the dark, removing the cotyledons and root radical, followed by culturing of the meristem explants. Twenty-four hours later, the primary leaves are removed to expose the apical meristem. The explants are placed apical dome side up and bombarded, e.g., twice with particles, followed by co-cultivation with *Agrobacterium*. To start the 25 co-cultivation for intact meristems, *Agrobacterium* is placed on the meristem. After about a 3-day co-cultivation period the meristems are transferred to culture medium with cefotaxime plus kanamycin for the NPTII selection.

The split meristem method involves imbibing seed, breaking of the cotyledons to produce a clean fracture at the plane of the embryonic axis, excising the root tip and then 30 bisecting the explants longitudinally between the primordial leaves. The two halves are placed cut surface up on the medium then bombarded twice with particles, followed by co-cultivation with *Agrobacterium*. For split meristems, after bombardment, the meristems are placed in an *Agrobacterium* suspension for 30 minutes. They are then removed from the suspension onto solid culture medium for three day co-cultivation.

After this period, the meristems are transferred to fresh medium with cefotaxime plus kanamycin for selection.

Transfer by Plant Breeding

5 Alternatively, once a single transformed plant has been obtained by the foregoing recombinant DNA method, conventional plant breeding methods can be used to transfer the gene and associated regulatory sequences via crossing and backcrossing. Such intermediate methods will comprise the further steps of: (1) sexually crossing the transgenic plant with a plant from a second taxon; (2) recovering reproductive material
10 from the progeny of the cross; and (3) growing transgenic plants from the reproductive material. Where desirable or necessary, the agronomic characteristics of the second taxon can be substantially preserved by expanding this method to include the further steps of repetitively: (1) backcrossing the transgenic progeny with non-transgenic plants from the second taxon; and (2) selecting for expression of an associated marker gene among the
15 progeny of the backcross, until the desired percentage of the characteristics of the second taxon are present in the progeny along with the gene or genes imparting marker gene trait.

By the term "taxon" herein is meant a unit of botanical classification. It thus includes, genus, species, cultivars, varieties, variants and other minor taxonomic groups which
20 lack a consistent nomenclature.

Regeneration of Transformants

The development or regeneration of plants from either single plant protoplasts or various explants is well known in the art (Weissbach and Weissbach, 1988). This regeneration and growth process typically includes the steps of selection of transformed
25 cells, culturing those individualized cells through the usual stages of embryonic development through the rooted plantlet stage. Transgenic embryos and seeds are similarly regenerated. The resulting transgenic rooted shoots are thereafter planted in an appropriate plant growth medium such as soil.

The development or regeneration of plants containing the foreign, exogenous
30 gene that encodes a polypeptide of interest introduced by *Agrobacterium* from leaf explants can be achieved by methods well known in the art such as described (Horsch et al., 1985). In this procedure, transformants are cultured in the presence of a selection agent and in a medium that induces the regeneration of shoots in the plant strain being transformed as described (Fraley et al., 1983). In particular, U.S. Pat. No. 5,349,124

(specification incorporated herein by reference) details the creation of genetically transformed lettuce cells and plants resulting therefrom which express hybrid crystal proteins conferring insecticidal activity against Lepidopteran larvae to such plants.

This procedure typically produces shoots within two to four months and those
5 shoots are then transferred to an appropriate root-inducing medium containing the selective agent and an antibiotic to prevent bacterial growth. Shoots that rooted in the presence of the selective agent to form plantlets are then transplanted to soil or other media to allow the production of roots. These procedures vary depending upon the particular plant strain employed, such variations being well known in the art.

10 Preferably, the regenerated plants are self-pollinated to provide homozygous transgenic plants, or pollen obtained from the regenerated plants is crossed to seed-grown plants of agronomically important, preferably inbred lines. Conversely, pollen from plants of those important lines is used to pollinate regenerated plants. A transgenic plant of the present invention containing a desired polypeptide is cultivated using
15 methods well known to one skilled in the art.

A preferred transgenic plant is an independent segregant and can transmit the CPP gene and its activity to its progeny. A more preferred transgenic plant is homozygous for the gene, and transmits that gene to all of its offspring on sexual mating. Seed from a transgenic plant may be grown in the field or greenhouse, and resulting
20 sexually mature transgenic plants are self-pollinated to generate true breeding plants. The progeny from these plants become true breeding lines that are evaluated for increased expression of the CPP transgene.

Method of Producing Transgenic Plants

Also included in the invention are methods of producing a transgenic plant. The
25 method includes introducing into one or more plant cells a compound that alters CaaX prenyl protease expression or activity in the plant to generate a transgenic plant cell and regenerating a transgenic plant from the transgenic cell. In some aspects the compound increases alters CaaX prenyl protease expression or activity. Alternatively, the compound decrease alters CaaX prenyl protease expression or activity. The compound
30 can be, e.g., (i) a CaaX prenyl protease polypeptide; (ii) a nucleic acid encoding a CaaX prenyl protease polypeptide; (iii) a nucleic acid that increases expression of a nucleic acid that encodes a CaaX prenyl protease polypeptide ; (iv) a nucleic acid that decreases the expression of a nucleic acid that encodes a CaaX prenyl protease polypeptide; (v) a

CaaX prenyl protease antisense nucleic acid and derivatives, fragments, analogs and homologs thereof. A nucleic acid that increases expression of a nucleic acid that encodes a CaaX prenyl protease polypeptide includes, *e.g.*, promoters, enhancers. The nucleic acid can be either endogenous or exogenous. Preferably, the compound is a CaaX prenyl 5 protease polypeptide or a nucleic acid encoding a CaaX prenyl protease polypeptide. For example the compound comprises the nucleic acid sequence of SEQ ID NO:1, 14, or 17 or fragment thereof. Alternatively, the compound is a CaaX prenyl protease antisense nucleic acid. For example the compound comprises the nucleic acid sequence of SEQ ID NO: 16, 19 or 20.

10 In various aspects the transgenic plant has an altered phenotype as compared to a wild type plant (*i.e.*, untransformed). By altered phenotype is meant that the plant has a one or more characteristic that is different from the wild type plant. For example, the transgenic plant has an increased resistance to stress. Increased stress resistance is meant that the transgenic plant can grow under stress conditions (*e.g.*, high salt, decreased 15 water, low temperatures, high temperatures) or under conditions that normally inhibit the growth of an untransformed. Stresses include, for example, chilling stress, heat stress, heat shock, salt stress, water stress (*i.e.*, drought), nutritional stress, disease, grazing pests, wound healing, pathogens such as for example fungi, bacteria, nematodes, viruses or parasitic weed and herbicides. Methodologies to determine plant growth or response 20 to stress include for example, height measurements, weight or biomass measurements, leaf area or number, ability to flower, water use, transpiration rates and yield. Alternatively, the transformed plant has an increased (*i.e.*, enhanced) ABA sensitivity. The enhanced ABA sensitivity is at the seedling growth stage. Alternatively, the enhanced ABA sensitivity is at the mature plant stage. Additional altered phenotypes 25 include for example, enhanced vegetative growth (*e.g.*, increased leaf number, thickness and overall biomass), delayed reproductive growth (*e.g.*, flowering later); enhanced seedling vigor (*e.g.*, increased root biomass and length), enhanced lateral root formation and therefore soil penetration more extensive vascular system resulting in an enhanced transport system.

30 The plant can be any plant type including, for example, species from the genera *Cucurbita*, *Rosa*, *Vitis*, *Juglans*, *Fragaria*, *Lotus*, *Medicago*, *Onobrychis*, *Trifolium*, *Trigonella*, *Vigna*, *Citrus*, *Linum*, *Geranium*, *Manihot*, *Daucus*, *Arabidopsis*, *Brassica*, *Raphanus*, *Sinapis*, *Atropa*, *Capsicum*, *Datura*, *Hyoscyamus*, *Lycopersicon*, *Nicotiana*,

Solanum, Petunia, Digitalis, Majorana, Ciahorium, Helianthus, Lactuca, Bromus, Asparagus, Antirrhinum, Heterocallis, Nemesis, Pelargonium, Panieum, Pennisetum, Ranunculus, Senecio, Salpiglossis, Cucumis, Browalia, Glycine, Pisum, Phaseolus, Lolium, Oryza, Zea, Avena, Hordeum, Secale, Triticum, Sorghum, Gossypium, Picea, 5 Caco, and Populus.

Screening Methods

The isolated nucleic acid molecules of the invention can be used to express CPP protein (e.g., via a recombinant expression vector in a host cell), to detect CPP mRNA (e.g., in a biological sample) or a genetic lesion in a CPP gene, and to modulate CPP 10 activity, as described further, below. In addition, the CPP proteins can be used to screen compounds that modulate the CPP protein activity or expression. In addition, the anti-CPP antibodies of the invention can be used to detect and isolate CPP proteins and modulate CPP activity.

The invention provides a method (also referred to herein as a "screening assay") 15 for identifying modulators, *i.e.*, candidate or test compounds or agents (e.g., peptides, peptidomimetics, small molecules or other drugs) that bind to CPP proteins or have a stimulatory or inhibitory effect on, *e.g.*, CPP protein expression or CPP protein activity. The invention also includes compounds identified in the screening assays described herein.

In one embodiment, the invention provides assays for screening candidate or test 20 compounds which bind to a CPP protein or polypeptide or biologically-active portion thereof. The test compounds of the invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic 25 library methods requiring deconvolution; the "one-bead one-compound" library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds. See, *e.g.*, Lam, 1997. *Anticancer Drug Design* 12: 145.

30 A "small molecule" as used herein, is meant to refer to a composition that has a molecular weight of less than about 5 kD and most preferably less than about 4 kD. Small molecules can be, *e.g.*, nucleic acids, peptides, polypeptides, peptidomimetics,

carbohydrates, lipids or other organic or inorganic molecules. Libraries of chemical and/or biological mixtures, such as fungal, bacterial, or algal extracts, are known in the art and can be screened with any of the assays of the invention.

Examples of methods for the synthesis of molecular libraries can be found in the art, for 5 example in: DeWitt, *et al.*, 1993. *Proc. Natl. Acad. Sci. U.S.A.* 90: 6909; Erb, *et al.*, 1994. *Proc. Natl. Acad. Sci. U.S.A.* 91: 11422; Zuckermann, *et al.*, 1994. *J. Med. Chem.* 37: 2678; Cho, *et al.*, 1993. *Science* 261: 1303; Carrell, *et al.*, 1994. *Angew. Chem. Int. Ed. Engl.* 33: 2059; Carell, *et al.*, 1994. *Angew. Chem. Int. Ed. Engl.* 33: 2061; and Gallop, *et al.*, 1994. *J. Med. Chem.* 37: 1233.

10 Libraries of compounds may be presented in solution (e.g., Houghten, 1992. *Biotechniques* 13: 412-421), or on beads (Lam, 1991. *Nature* 354: 82-84), on chips (Fodor, 1993. *Nature* 364: 555-556), bacteria (Ladner, U.S. Patent No. 5,223,409), spores (Ladner, U.S. Patent 5,233,409), plasmids (Cull, *et al.*, 1992. *Proc. Natl. Acad. Sci. USA* 89: 1865-1869) or on phage (Scott and Smith, 1990. *Science* 249: 386-390; 15 Devlin, 1990. *Science* 249: 404-406; Cwirla, *et al.*, 1990. *Proc. Natl. Acad. Sci. U.S.A.* 87: 6378-6382; Felici, 1991. *J. Mol. Biol.* 222: 301-310; Ladner, U.S. Patent No. 5,233,409.).

In one embodiment, an assay is a cell-based assay in which a cell which expresses a CPP protein, or a biologically-active portion thereof, is contacted with a test compound and 20 the ability of the test compound to bind to a CPP protein determined. The cell, for example, can be of mammalian origin, plant cell or a yeast cell. Determining the ability of the test compound to bind to the CPP protein can be accomplished, for example, by coupling the test compound with a radioisotope or enzymatic label such that binding of the test compound to the CPP protein or biologically-active portion thereof can be 25 determined by detecting the labeled compound in a complex. For example, test compounds can be labeled with ^{125}I , ^{35}S , ^{14}C , or ^3H , either directly or indirectly, and the radioisotope detected by direct counting of radioemission or by scintillation counting. Alternatively, test compounds can be enzymatically-labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label 30 detected by determination of conversion of an appropriate substrate to product. In one embodiment, the assay comprises contacting a cell which expresses a CPP protein, or a biologically-active portion thereof, with a known compound which binds CPP to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with a CPP protein, wherein determining the

ability of the test compound to interact with a CPP protein comprises determining the ability of the test compound to preferentially bind to CPP protein or a biologically-active portion thereof as compared to the known compound.

In another embodiment, an assay is a cell-based assay comprising contacting a 5 cell expressing a CPP protein, or a biologically-active portion thereof, with a test compound and determining the ability of the test compound to modulate (*e.g.*, stimulate or inhibit) the activity of the CPP protein or biologically-active portion thereof.

Determining the ability of the test compound to modulate the activity of CPP or a biologically-active portion thereof can be accomplished, for example, by determining the 10 ability of the CPP protein to bind to or interact with a CPP target molecule. As used herein, a "target molecule" is a molecule with which a CPP protein binds or interacts in nature, for example, a molecule on the surface of a cell which expresses a CPP interacting protein, a molecule on the surface of a second cell, a molecule in the extracellular milieu, a molecule associated with the internal surface of a cell membrane 15 or a cytoplasmic molecule. A CPP target molecule can be a non-CPP molecule or a CPP protein or polypeptide of the invention. In one embodiment, a CPP target molecule is a component of a signal transduction pathway that facilitates transduction of an extracellular signal (*e.g.* a signal generated by binding of a compound to a membrane-bound molecule) through the cell membrane and into the cell. The target, for 20 example, can be a second intercellular protein that has catalytic activity or a protein that facilitates the association of downstream signaling molecules with CPP.

Determining the ability of the CPP protein to bind to or interact with a CPP target molecule can be accomplished by one of the methods described above for determining direct binding. In one embodiment, determining the ability of the CPP protein to bind to or interact with a CPP target molecule can be accomplished by determining the activity 25 of the target molecule. For example, the activity of the target molecule can be determined by detecting induction of a cellular second messenger of the target (*i.e.* intracellular Ca^{2+} , diacylglycerol, IP_3 , etc.), detecting catalytic/enzymatic activity of the target an appropriate substrate, detecting the induction of a reporter gene (comprising a 30 CPP-responsive regulatory element operatively linked to a nucleic acid encoding a detectable marker, *e.g.*, luciferase), or detecting a cellular response, for example, cell survival, cellular differentiation, or cell proliferation.

In yet another embodiment, an assay of the invention is a cell-free assay comprising contacting a CPP protein or biologically-active portion thereof with a test

compound and determining the ability of the test compound to bind to the CPP protein or biologically-active portion thereof. Binding of the test compound to the CPP protein can be determined either directly or indirectly as described above. In one such embodiment, the assay comprises contacting the CPP protein or biologically-active portion thereof
5 with a known compound which binds CPP to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with a CPP protein, wherein determining the ability of the test compound to interact with a CPP protein comprises determining the ability of the test compound to preferentially bind to CPP or biologically-active portion thereof as compared to the
10 known compound.

In still another embodiment, an assay is a cell-free assay comprising contacting CPP protein or biologically-active portion thereof with a test compound and determining the ability of the test compound to modulate (*e.g.* stimulate or inhibit) the activity of the CPP protein or biologically-active portion thereof. Determining the ability of the test compound to modulate the activity of CPP can be accomplished, for example, by
15 determining the ability of the CPP protein to bind to a CPP target molecule by one of the methods described above for determining direct binding. In an alternative embodiment, determining the ability of the test compound to modulate the activity of CPP protein can be accomplished by determining the ability of the CPP protein further modulate a CPP
20 target molecule. For example, the catalytic/enzymatic activity of the target molecule on an appropriate substrate can be determined as described above.

In yet another embodiment, the cell-free assay comprises contacting the CPP protein or biologically-active portion thereof with a known compound which binds CPP protein to form an assay mixture, contacting the assay mixture with a test compound, and
25 determining the ability of the test compound to interact with a CPP protein, wherein determining the ability of the test compound to interact with a CPP protein comprises determining the ability of the CPP protein to preferentially bind to or modulate the activity of a CPP target molecule.

The cell-free assays of the invention are amenable to use of both the soluble form
30 or the membrane-bound form of CPP protein. In the case of cell-free assays comprising the membrane-bound form of CPP protein, it may be desirable to utilize a solubilizing agent such that the membrane-bound form of CPP protein is maintained in solution. Examples of such solubilizing agents include non-ionic detergents such as n-octylglucoside, n-dodecylglucoside, n-dodecylmaltoside,

- octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, Triton® X-100, Triton® X-114, Thesit®, Isotridecypoly(ethylene glycol ether)_n, N-dodecyl--N,N-dimethyl-3-ammonio-1-propane sulfonate, 3-(3-cholamidopropyl)dimethylamminiol-1-propane sulfonate (CHAPS), or
- 5 3-(3-cholamidopropyl)dimethylamminiol-2-hydroxy-1-propane sulfonate (CHAPSO).

In more than one embodiment of the above assay methods of the invention, it may be desirable to immobilize either CPP protein or its target molecule to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to CPP protein, 10 or interaction of CPP protein with a target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtiter plates, test tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided that adds a domain that allows one or both of the proteins to be bound to a matrix. For example, 15 GST-CPP fusion proteins or GST-target fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtiter plates, that are then combined with the test compound or the test compound and either the non-adsorbed target protein or CPP protein, and the mixture is incubated under conditions conducive to complex formation (*e.g.*, at physiological conditions for salt and 20 pH). Following incubation, the beads or microtiter plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described, *supra*. Alternatively, the complexes can be dissociated from the matrix, and the level of CPP protein binding or 25 activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either the CPP protein or its target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated CPP protein or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques well-known within the art (*e.g.*, biotinylation 30 kit, Pierce Chemicals, Rockford, Ill.), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with CPP protein or target molecules, but which do not interfere with binding of the CPP protein to its target molecule, can be derivatized to the wells of the plate, and unbound target or CPP protein trapped in the wells by antibody conjugation. Methods for

detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the CPP protein or target molecule, as well as enzyme-linked assays that rely on detecting an enzymatic activity associated with the CPP protein or target molecule.

5 In another embodiment, modulators of CPP protein expression are identified in a method wherein a cell is contacted with a candidate compound and the expression of CPP mRNA or protein in the cell is determined. The level of expression of CPP mRNA or protein in the presence of the candidate compound is compared to the level of expression of CPP mRNA or protein in the absence of the candidate compound. The 10 candidate compound can then be identified as a modulator of CPP mRNA or protein expression based upon this comparison. For example, when expression of CPP mRNA or protein is greater (*i.e.*, statistically significantly greater) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of CPP mRNA or protein expression. Alternatively, when expression of CPP 15 mRNA or protein is less (statistically significantly less) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of CPP mRNA or protein expression. The level of CPP mRNA or protein expression in the cells can be determined by methods described herein for detecting CPP mRNA or protein.

20 In yet another aspect of the invention, the CPP proteins can be used as "bait proteins" in a two-hybrid assay or three hybrid assay (*see, e.g.*, U.S. Patent No. 5,283,317; Zervos, *et al.*, 1993. *Cell* 72: 223-232; Madura, *et al.*, 1993. *J. Biol. Chem.* 268: 12046-12054; Bartel, *et al.*, 1993. *Biotechniques* 14: 920-924; Iwabuchi, *et al.*, 1993. *Oncogene* 8: 1693-1696; and Brent WO 94/10300), to identify other proteins that 25 bind to or interact with CPP ("CPP-binding proteins" or "CPP-bp") and modulate CPP activity. Such CPP-binding proteins are also likely to be involved in the propagation of signals by the CPP proteins as, for example, upstream or downstream elements of the CPP pathway.

30 The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. In one construct, the gene that codes for CPP is fused to a gene encoding the DNA binding domain of a known transcription factor (*e.g.*, GAL-4). In the other construct, a DNA sequence, from a library of DNA sequences, that encodes an unidentified protein ("prey" or "sample") is fused to a gene

that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact, *in vivo*, forming a CPP-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (*e.g.*, LacZ) that is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene that encodes the protein which interacts with CPP.

In yet another aspect of the invention are methods which utilize the transgenic plants of the invention to identify CPP-interacting components via genetic screening protocols. These components can be for example, regulatory elements which modify CPP-gene expression, interacting proteins which directly modify CPP activity or interacting proteins which modify components of the same signal transduction pathway and thereby exert an effect on the expression or activity of CPP. Briefly, genetic screening protocols are applied to the transgenic plants of the invention and in so doing identify related genes which are not identified using a wild type background for the screen. For example an activation tagged library (Weigel, *et al.*, 2000. *Plant Physiol.* 122: 1003-1013), can be produced using the transgenic plants of the invention as the genetic background. Plants are then screened for altered phenotypes from that displayed by the parent plants. Alternative methods of generating libraries from the transgenic plants of the invention can be used, for example, chemical or irradiation induced mutations, insertional inactivation or insertional activation methods.

The invention further pertains to novel agents identified by the aforementioned screening assays and uses thereof.

25

EXAMPLES

Example 1: RT-PCR amplification and cloning of CaaX prenyl proteases

Total RNA was isolated from leaf tissue of *Arabidopsis thaliana*, *Brassica napus* and *Glycine max*, using the Qiagen RNeasy kit and used as template to amplify the CPP genes by RT-PCR. Reaction conditions were as follows; 1X reaction buffer (10mM Tris-HCl pH 8.8, 1.5mM MgCl₂, 50mM KCl), dNTP's at 200μM, 1pM AtCPP BamFW and

AtCPP SmaRV primers, 2.5U. Pfu DNA polymerase, and template plus water to a final volume of 100µL. Reactions were run at 1 minute 94°C, 1 minute 60°C, 1 minute 72°C, for 30 cycles. Primers used to PCR amplify *Arabidopsis* and *Brassica* sequences were those identified by SEQ ID NO:6 and SEQ ID NO:7. Primers used to PCR amplify the 5 *Glycine* sequence were those identified by SEQ ID NO:54 and SEQ ID NO:55. PCR products were separated from the RT-PCR reaction mixture using the Qiagen PCR column spin kit and ligated into the prepared cloning vector, pBluescript KS+. The vector had been prepared by digestion with *EcoRV* and treated with *Taq* polymerase in the presence of dTTP to produce a 3' overhand suitable for ligation with the PCR 10 products. The ligation products were transformed into *E. coli* DH5α cells, positive colonies selected and the resulting inserts sequenced. The above methodology is applicable to obtain homologous sequences and may require alternative primers.

Table 1.

15	AtCPP BamFW:	5'-AAAGGATCCATGGCGATTCTTCATGG-3' (SEQ ID NO:6)
	AtCPP SmaRV:	5'-AAACCCGGGTTAACCTGTCTTGTCTTCTCCA-3' (SEQ ID NO:7)
20	GmCPP SmaFW:	5'-AAACCCGGGATGGCGTTCCCTACATGGAAGCC - 3' (SEQ ID NO:54)
	GmCPP SacRV:	5'-AAAGAGCTCTTAGTCTCCTTATCCGGTTCG -3' (SEQ ID NO:55)

Example 2: Vector Construction

Construction of the pBI121-AtCPP construct (SEQ ID NO: 4) was prepared as 25 follows. The pBI121 vector was digested with *BamHI* and *SmaI*. The AtCPP, 1.4 kb DNA fragment from RT-PCR (SEQ ID NO: 1) was digested with *BamHI* and *SmaI* and ligated into the pBI121 vector. The GUS sequence was then removed by digestion with *SmaI* and *EcoICRI* and the vector ligated after purification of the vector from the GUS insert to produce the pBI121-AtCPP vector (Figure 1A). This construct was used to 30 further generate constructs expressing the CPP gene from *Brassica* and *Glycine*. To produce the pBI121-BnCPP construct (SEQ ID NO:47) primer pairs identified by SEQ ID NO:6 and SEQ ID NO:7 are used to PCR amplify the appropriate fragment which is ligated into the prepared parent vector. To produce the pBI121-GmCPP construct (SEQ

ID NO:41) primer pairs identified by SEQ ID NO:54 and SEQ ID NO:55 are used to PCR amplify the appropriate fragment which is ligated into the prepared parent vector.

Construction of the pBI121-antisense-AtCPP construct (SEQ ID NO:35). The antisense fragment was produced using PCR amplification with SEQ ID NO:1 as template and primers identified as SEQ ID NO:11 and SEQ ID NO:12, listed in Table 2. This fragment was digested with *Bam*HI and *Sma*I and used to replace the sense fragment of the pBI121-AtCPP construct (SEQ ID NO: 4), to yield SEQ ID NO:35 (Figure 1B) . This construct, SEQ ID NO:35, was used to further generate constructs expressing the antisense CPP gene from *Brassica* and *Glycine*. To produce the pBI121-antisense-BnCPP construct (SEQ ID NO:49) primer pairs identified by SEQ ID NO:56 and SEQ ID NO:57 are used to PCR amplify the appropriate fragment which is ligated into the prepared parent vector. To produce the pBI121-antisense-GmCPP construct (SEQ ID NO:43) primer pairs identified by SEQ ID NO:58 and SEQ ID NO:59 are used to PCR amplify the appropriate fragment which is ligated into the prepared parent vector.

Construction of the pBI121-HP-AtCPP construct (SEQ ID NO: 5). The cloning strategy involved truncating the GUS gene of pBI121 and flanking the GUS sequence with a AtCPP fragment in the antisense orientation upstream of the GUS and in the sense orientation on the downstream side of GUS. The pBI121 vector was digested with *Sma*I and *Sac*I, the GUS sequence and the vector fragments were purified from one another. The isolated GUS fragment was digested using *Eco*RV and the 1079 bp. blunt ended *Eco*RV/*Sac*I fragment isolated. This was ligated back into the digested parent vector at the *Sma*I/*Sac*I sites. This intermediate vector was used in the subsequent production of the hair-pin vectors. The AtCPP fragment to be used as the gene specific hair-pin sequence was isolated by PCR. Primers identified as SEQ ID NO:8 and SEQ ID NO:9, listed in Table 2, were used to generate a 596 bp fragment. Cloning of the sense orientation fragment was achieved by digesting the PCR AtCPP fragment with *Sac*I and ligation into the *Sac*I site at the 3' end of GUS. To insert the same fragment upstream of GUS, the *Bam*HI site was opened and the ends blunted with Klenow. The PCR amplified AtCPP fragment was digested with *Eco*CRI, which is an isoschizomer of *Sac*I but leaves blunt ends, and ligated into the blunted *Bam*HI site of the vector to yield the final construct (Figure 1C). The intermediate construct used to produce SEQ ID NO:5 above contained only the truncated GUS gene and no CPP sequences this intermediate vector was used to further generate constructs expressing hair-pin CPP gene constructs from

- Brassica* and *Glycine*. To produce the pBI121-HP-BnCPP construct (SEQ ID NO:48) primer pairs identified by SEQ ID NO:58 and SEQ ID NO:59 are used to PCR amplify the sense fragment and primer pairs identified by SEQ ID NO:60 and SEQ ID NO:61 are used to PCR amplify the antisense fragment. These fragments are cloned into the
- 5 prepared intermediate vector described above. To produce the pBI121-HP-GmCPP construct (SEQ ID NO:42) primer pairs identified by SEQ ID NO:62 and SEQ ID NO:63 are used to PCR amplify the sense fragment and primer pairs identified by SEQ ID NO:64 and SEQ ID NO:65 are used to PCR amplify the antisense fragment. These fragments are cloned into the prepared intermediate vector described above.
- 10 The above vector constructs were modified to place the genes under the control of alternative promoters, such as, but not limited to, the RD29A or MuA . This was accomplished by excising the 35S promoter sequence and replacing it with an appropriate promoter sequence. In this way SEQ ID NO's:39 and 40 were generated and SEQ ID NO's:38, 41-53 can be constructed.

15 **Table 2**

	AtCPP-HP-SacFW 5'-CTGGAGCTTTACCGAGGTTGGGCCTTGATCC-3' (SEQ ID NO:8)
	AtCPP-HP-SacRV 5'-ATTGAGCTCCAATGTCCAAGCTCGTGTGCAATA-3' (SEQ ID NO:9)
20	AtCPP-anti-SmaFW 5'-AAACCCGGGATGGCGATTCTTCATGG-3' (SEQ ID NO:11)
	AtCPP-anti-BamRV 5'-AAAGGATCCTTAATCTGTCTTGTCTCTCCA-3' (SEQ ID NO:12)
25	BnCPP-anti-SmaFW 5'-AAACCCGGGATGGCGATTCTTCATGG -3' (SEQ ID NO:56)
	BnCPP-anti-BamRV 5'-AAAGGATCCTTAATCTGTCTTGTCTCTCC -3' (SEQ ID NO:57)
30	BnCPP-HP-Sac-FW 5'- AAAGAGCTCTTCTACCAATGGTGGGACTCG -3' (SEQ ID NO:58)
	BnCPP-HP-Sac-RV 5'- AAAGAGCTCCCAGTGTCCCAGCTCGTGTG -3' (SEQ ID NO:59)
35	BnCPP-HP-BamFW 5'- AAAGGATCCTTCTACCAATGGTGGGACTCG -3' (SEQ ID NO:60)
	BnCPP-HP-XbaRV 5'- AAATCTAGACCAGTGTCCCAGCTCGTGTG -3' (SEQ ID NO:61)
	GmCPP-HP-Sac-FW 5'-GATGAGCTACAAGATCAAGTCACAGCAATGCCT -3'

(SEQ ID NO:62)

GmCPP-HP-Sac-RV 5'- AAAGAGCTCCGGTTCGTCCAGCGCGGCC -3'
(SEQ ID NO:63)

GmCPP-HP-BamFW

5 5'- GATGGATCCACAAGATCAAGTCACAGCAATGCCT -3'
(SEQ ID NO:64)

GmCPP-HP-XbaRV 5'- CCTTCTAGACCGGTTCGTCCAGCGCGGCC -3'
(SEQ ID NO:65)

10 Example 3: Sequence Analysis

Arabidopsis thaliana CPP (AtCPP)

A disclosed nucleic acid of 1275 nucleotides (SEQ ID NO:1) and also referred to as AtCPP, is shown in Table 3.

Table 3A. AtCPP Nucleotide Sequence (SEQ ID NO:1).

ATGGCGATTCTTCAATGGAAACCGTGTGGGTTTATGATAGTGATGTACATTTGAG ACGTATTGGATCTGAGGCAACTCACTGCTCTCAAGCTCCAACCTCTCCGAAAACCTTG GTTGGTGAATTAGCCAAGAGAAGTTGAGAAATCAGCAGCATACAGTCTTGACAAAAGC TATTTCACTTGTTCATGAGTTGTAACTATACTTATGGACTCTGCAATTTGTTCTT GGGATCTGCCTGGTTTGAAGATGTCTGGAGCTGTTACCGAGGTTGGCCTTGAT CCGGAGAACATGAAACTGCATACTCTTCATTCTGGCTGGTGTATGACATGGCACAG ATCACTGATTGCCATTTCCTTGACTCAACTTCTGATCGAGTCTGGCATGGTTC AACAAACAAACAATATGGATGTTCAATTAGGGACATGATCAAAGGAACATTCCCTCTGTC ATACTAGGCCACCATTGTGCTGCGATAATTTCATAGTCCAGAAAGGAGGTCTTAT CTTGCATCTATCTGTGGCATTCACTGTTATCCTGTCTAGTGATGACTATATAC CCGGTCTTGATAGCACCCTTCAACAAATTCACTCCTCCAGATGGAGACCTCCGG GAGAAGATTGAGAAACTGCTCTCCCTAAAGTTCTTGAGAAGAGCTGTTGTC GATGGATCTACAAGGTCAAGCCATAGCAATGCTACATGTATGGTTCTTAAGAACAAA AGGATTGTTCTTATGATACGTTGATTGAGCTGCAAGAACGAGGATGAAATTGTGGCG GTTATTGCACACGAGCTGGACATTGAAACTGAATCACACTACATACTGTTCATGCA GTTCAAATCCTGCCTTCTTACAATTGGAGGATACACTCTCAGAAACTCCACTGAT CTCTCAGGAGTTGGATTGATACACAGCCTGTTCTATTGGTTGATCATATTCAG CACACTGTAATACCACTGCAACATCTAGTAAGCTTGGCCTGAACCTCGTTAGTCGAGCG TTTGAGTTCAAGGCTGATGCTTGTGAAGCTGACTATGCAAAAGATCTCGTCCT GCTCTAGTGAACACTACAGGAAGAGAACCTATCAACAATGAACACTGATCCATTGACTCA GCTTATCACTACTCACATCCTCCTTGTGAAGGCTCGAGCCACTGATGGAGAACAC AAGAACAGATTTAA

A disclosed CPP polypeptide (SEQ ID NO:2) encoded by SEQ ID NO:1 has 424 amino acid residues and is presented in Table 3B using the one-letter amino acid code.

Table 3B. Encoded CPP protein sequence (SEQ ID NO:2).

MAIPFMETVVGFMIVMYIFETYLDLRQLTALKLPLPKTLVGVISQEKFESRAYSLDKS YFHFVHEFVTILMDSAILFFGILPWFWKMSGAVLPRLGLDPENEILHTLSFLAGVMTWSQ ITDLPFSLYSTFVIESRHGFNKQTIWMFIRDMIKGTFLSVILGPPIVAIIIFIVQKGGPY LAIYLWAFMFILSLVMMTIYPVLIAPLFNKFTPPLPDGDLREKIEKLASSLKPLKKLFVV DGSTRSSHNSAYMYGFFKNKRIVLYDTLIQQCKNEDEIVAVIAHELGHWKLNHTTYSFIA VQILAFLQFGGYTLLRNSTDLFRSFGFDTQPVLIGLIIFQHTVIPLQHLVSFGNLVSRA FEFQADAFAVKLDYAKDLRPALVKLQEENLSTMNTDPLYSAYHYSHPPLVERLRATGED KKTD
--

The present invention also includes a nucleic acid sequence complimentary to the 5 *Arabidopsis thaliana* CaaX prenyl protease of SEQ ID NO:1. The disclosed complimentary sequence is shown as SEQ ID NO:20.

SEQ ID NO:20

TTAATCTGTCTTCTTGCTTCCATCAGTGGCTCGAACGCCTTCAACAAGAGGAGGAT 10 GTGAGTAGTGATAAGCTGAGTACAATGGATCAGTGGTCAATTGTTGATAAGTTCTTCC TGTAGTTCACTAGAGCAGGACGAAGATCTTGCATAGTCAGCTCACAGCAAAGC ATCAGCCTGAAACTCAAACGCTCGACTAACGAGGTTAGGCCAAAGCTTACTAGATGTT GCAGTGGTATTACAGTGTGCTGAAATATGATCAAACCAATGAGAACAGGCTGTATCA AATCCGAAACTCCTGAAGAGATCAGTGGAGTTCTGAGAAGAGTGTATCCTCCAAATTG 15 TAAGAAGGCAAGGATTGAACTGCAATGAACGAGTATGTAGTGTGATTCAAGTTCCAAT GTCCAAGCTCGTGTCAATAACCGCCACAATTTCATCCTCATTTGCAGTGTGAATC AACGTATCATAAAGAACAAATCCTTTGTTCTAAAGAAACCATACTGTAAGCATTGCT ATGGCTTGACCTTGTAGATCCATCGACAACAAACAGCTTCTCAAAGGAAACTTAGGG AAGAAGCAAGTTCTCAATCTTCTCCGGAGGTCTCCATCTGGAAAGAGGAGTGAATTG 20 TTGAAGAGCGGTGCTATCAAGACCGGGTATATAGTCATCATCACTAGAGACAGGATAAA CATGAATGCCACAGATAGATGGCAAGATAAGGACCTCCTTCTGGACTATGAAAATTA TCGCAGCAACAATGGGTGGCCTAGTATGACAGAGAGGAATGTCCTTGATCATGTCC CTAATGAACATCCATATTGTTGTTGTAACCCATGCCAGACTCGATCACGAAAGT TGAGTACAAAGAAATGGCAAATCAGTGTGACCATGTATAACACCAGCCAAGA 25 ATGAAAGAGTATGCAGTATTTCATTCTCCGGATCAAGGCCAACCTCGTAAACAGCT CCAGACATCTCCAAAACCAAGGCAAGATCCAAAGAACAAAATTGCAGAGTCCATAAG

TATAGTTACAAACTCATGAACAAAGTGAAAATAGCTTGTCAAGACTGTATGCTCGTG
ATTCTCAAACCTCTGGCTAATTACACCAACCAAGGTTTCGGGAGAGTTGGAAGC
TTGAGAGCAGTGAGTTGCCTCAGATCCAAATACGTCTCAAAAATGTACATCACTATCAT
AAAACCCACGACGGTTCCATGAAAGGAATGCCAT

5

Due to the nature of the cloning strategy the sequence presented is not full length but is missing the 5' and 3' non-translated regions. The percent identities of the *Arabidopsis thaliana* nucleotide sequence and its encoded amino acid sequence to that of other CPP sequences as determined by ClustalW analysis are shown in Figure 2.

- 10 Using the sequences disclosed herein as hybridization probes, one is able to screen and isolate full length sequences from cDNA or genomic libraries or use the rapid amplification of cDNA ends (RACE) technology or other such PCR techniques.

***Brassica napus* CPP (BnCPP)**

- 15 A disclosed nucleic acid of 1275 nucleotides (SEQ ID NO:14) and also referred to as BnCPP, is shown in Table 4.

Table 4A. BnCPP Nucleotide Sequence (SEQ ID NO:14).

ATGGCGATTCTTCAATGGAAACCGTGTGGTTATGATAGTGTACGTTTGAGACGTA TTGGATCTGAGGCAACATACTGCTCTCAAGCTTCCACTCTCCAAAGACTTGGTGGAGTCA TTAGCCAAGAGAAGTTGAGAAATCTGAGCTTACAGTCTGACAAAAGCCATTTCACTTGTT CATGAGTTGTTACTATACTTATGGACTCTGCGATTCTGTTCTGGATCTGCCTGGTTTG GAAGATATCTGGGGCTTCTACCAATGGTGGACTCGATCCAGAGAATGAAATCTGCACACTC TTTCATTCTGGCTGGTCTTATGACATGGTCACAGATCACTGATTGCCATTTCTTGTACTCA ACTTCGTGATCGAGTCTGGCATGGTCAACAAACAAACATGGATGTTCTAGGGACAT GATCAAAGGAATACTCCTCTGTACACCTGCCCTATCGTGGCAATTATTGTTATAG TTCAGAAAGGAGGTCTTACCTGCCATCTATCTGTTGCGATTGTTATCCTGTCTAGTG ATGATGACTATATACCCGTGGATTGACCTCTTCAACAAGTTCACTCCTCTGATGG AGACCTCCGGGAGAAGATTGAGAAACTGCTTCTCTAAAGTTCTGAAGAAGCTGTTG TTGTCGATGGATCTACAAGGTCAAGCCATAGTAATGCTTACATGTATGGTTCTCAAGAACAAA AGGATTGTTCTTATGACACATTGATTGAGCAGTGCAGAATGAGAATGAAATTGCGGTTAT TGCACACGAGCTGGGACACTGGAAGCTGAATCACACTACATCGTTCATGCTGTTCAAATCC TTGCCTTCTGCAATTGGAGGATACACTCTGTCAGAAACTCCACTGATCTTCAGGAGTTT GGTTTGATACACAACCAGTCTCATGGTTGATCATATTTCAGCACACTGTAATACCACCTCA ACACCTAGTAAGCTTGACCTCAACCTGTTAGTCGAGCGTTGAGTTCAAGGCTGATGCTTTG CAGTGAATCTGGTTATGCAAAGGATCTACGTCCCTGCCCTAGTGAAGCTACAGGAAGAGAACTTA
--

TCAGCGATGAACACAGACCCATTGTACTCAGCTTATCACTACTCACACCCTCCTTGAGAGAG GCTTCGAGCCATTGATGGAGAAAGACAAGAACAGATTAA

A disclosed CPP polypeptide (SEQ ID NO:15) encoded by SEQ ID NO:14 has 424 amino acid residues and is presented in Table 4B using the one-letter amino acid code.

Table 4B. Encoded CPP protein sequence (SEQ ID NO:15).

MAIPFMETVVGFMIVMYVFETYLDLRQHTALKLPLPKTLVGVISQEKFKEKSRAYSLDKSHFHF VHEFVTILMDSAIILFFGILPWFWKISGGFLPMVGLDPENEILHTLSFLAGLMTWSQITDLPFSL YSTFVIESRHGFNKQTIIWMFIRDMIKGILLSVIPAPPIVAAIIVIVQKGGPYLAIYLWAFMFIL SLVMMTIYPVLIAPLFNKFPLPDGDLREKIEKLASSLKFPKKLFVVVDGSTRSSHNSAYMYGF FKNKRIVLYDTLIQQCQNENEIVAVIAHELGHWKLNHTTYSFIAVQILAFLQFGGYTLVRNSTD LFRSFGFDTQPVLIGLIIFQHTVIPLQHLVSFDLNLVSRAFEFQADAFAVNLGYAKDLRPALVK LQEENLSAMNTDPLYSAYHYSHPPLVERLRAIDGEDKKTD

5

The present invention also includes a nucleic acid sequence complimentary to the *Brassica napus* CaaX prenyl protease of SEQ ID NO:14. The disclosed complimentary sequence is shown as SEQ ID NO:16.

10 SEQ ID NO:16

TTAATCTGTCTTCTTGTCTTCTCCATCAATGGCTCGAACGCCTCTACAAGAGGAGGGT
GTGAGTAGTGATAAGCTGAGTACAATGGGTCTGTGTTCATCGCTGATAAGTTCTCTTCC
TGTAGCTTCACTAGGGCAGGACGTAGATCCTTGCATAACCAAGATTCACTGCAAAAGC
ATCAGCCTGAAACTCAAACGCTCGACTAACAAAGGTTGAGGTCAAAGCTTACTAGGTGTT
15 GAAGTGGTATTACAGTGTGCTGAAATATGATCAAACCAATGAGAACTGGTTGTATCA
AAACCAAAACTCCTGAAGAGATCAGTGGAGTTCTGACAAGAGTGTATCCTCCAAATTG
CAAGAAGGCAAGGATTGAACAGCAATGAACGAGTATGTAGTGTGATTCACTGCTTCCAGT
GTCCCAGCTCGTGTGCAATAACCGCCACAATTCTCATTCTGGCACTGCTGAATC
AATGTGTATAAAGAACAAATCCTTTGTTCTGAAGAAACCATACTGTAAGCATTACT
20 ATGGCTTGACCTTGTAGATCCATCGACAACAAACAGCTTCTCAGAGGAAACTTAGAG
AAGAAGCAAGTTCTCAATCTTCTCCGGAGGTCTCCATCAGGAAGAGGAGTGAACCTG
TTGAAAAGAGGTGCAATCAAACAGGGTATATAGTCATCATCACTAGAGACAGGATAAA
CATGAATGCCACAGATAGATGGCGAGGTAAGGACCTCCTTCTGAACTATAACAATAA
TTGCGGCAACGATAGGAGGGCAGGTATGACAGAGAGGAGTATTCCCTTGATCATGTCC
25 CTAATGAACATCCATATTGTTGTTGAACCCATGCCGAGACTCGATCACGAAAGT

TGAGTACAAAGAAAATGGCAAATCAGTGATCTGTGACCATGTCATAAGACCAGCCAAGA
 ATGAAAGAGTGTGCAGGATTCTATTCTCTGGATCGAGTCCCACCATTGGTAGAAAGCCG
 CCAGATATCTTCAAAACCAAGGCAAGATCCAAAGAACAGAACAGAATCGCAGAGTCCATAAG
 TATAGTAACAAACTCATGAACAAAGTGAAATGGCTTGTCAAGACTGTAAGCTCGAG
 5 ATTCTCAAACCTCTTGGCTAATGACTCCAACCAAAGTCTTGGAGAGTGGGAAGC
 TTGAGAGCAGTATGTTGCCTCAGATCAAATACGTCTAAAAACGTACATCACTATCAT
 AAAACCAACGACGGTTCCATGAAAGGAATGCCAT

Due to the nature of the cloning strategy the sequence presented is not full length
 10 but is missing the 5' and 3' non-translated regions. The percent identities of the *Brassica napus* nucleotide sequence and its encoded amino acid sequence to that of other CPP sequences as determined by ClustalW analysis are shown in Figure 2.
 Using the sequences disclosed herein as hybridization probes, one is able to screen and isolate full length sequences from cDNA or genomic libraries or use the rapid
 15 amplification of cDNA ends (RACE) technology or other such PCR techniques.

Glycine max CPP (GmCPP)

A disclosed nucleic acid of 1275 nucleotides (SEQ ID NO:17) and also referred to as GmCPP, is shown in Table 5.

20

Table 5A. GmCPP Nucleotide Sequence (SEQ ID NO:17).

ATGGCGTTCCCTACATGGAAGCCGTTGTCGGATTTATGATATTAATGTACATTGAAACTTA CTTGGATGTGCGACAACATAGGGCCCTCAAACCTTCTACTCTTCCAAAGACTTTAGAGGGTGT TCAGCCAAGAGAAATTGAGAAATCTAGAGCCTATAGTCTTGATAAAAGCCACTCCATTGTT CACGAGTTGTGACAATAGTGACAGACTCTACAATTGTTACTTTGGGTATTGCCCTGGTTTG GAAGAAATCAGGAGATTGACAATAGCTGGTTCAATGCTGAGAATGAAACTGCATACCC TTGCCTCTTAGCAGGGCTGATGATTGGTCACAGATAACAGATTGCCCTTTCTGTACTCA ACTTTGTGATTGAGGCCGTCATGGTTAATAAGCAAACACCAGGTTATTCTTAGGGACAT GCTTAAAGGAATTTCCTTCTGTAATAATTGGTCCACCTATTGTGGCTGCAATCATTGTAATAG TACAGAAAGGAGGTCCATACTTGGCCATCTATCTTGGTTTACGTTGGTCTTCTATTGTG ATGATGACCCATTATCCAGTACTAATAGCTCCACTCTCAATAAGTTCACTCCACTCCAGATGG TCAACTCAGGGAGAAAATCGAGAAACTGCTTCCTCCCTCAACTATCGTTAAAGAAACTATTG TTGTCGATGGATCCACAAGATCAAGTCACAGCAATGCCATTGATGGATTCTCAAGAACAG AGGATTGTCCTTATGACACATTAATTCAACAGTGAAAGACGATGAGGAAATTGTTGCTTAT TGCCCATGAGTTGGGACACTGGAAGCTCAACCATACTGTGACACATTGTTGCTATGCAGATT TTACACTCTACAATTGGAGGATATACACTAGTGCAGCTGATCTGTATCGAAGCTT

```

GGGTTTGATACGCAGCCAGTCCTCATGGGCTCATCATATTCACTGTAATCCCACCTCA
GCAATTGGTCAGCTTGGCTGAACCTAGTCAGCCGATCATTGAATTTCAGGCTGATGGCTTG
CCAAGAAGCTTGGATATGCATCTGGATTACCGCGGTTGTGAAACTACAGGAGGAGAACCTG
TCAGCTATGAATAACAGATCCTGGTACTCTGCTTATCACTATTCTCATCCTCCCTGTTGAAAG
ATTGGCCCGCTGGACGAACCGATAAGAAGGAAGACTAA

```

A disclosed CPP polypeptide (SEQ ID NO:18) encoded by SEQ ID NO:17 has 424 amino acid residues and is presented in Table 5B using the one-letter amino acid code.

Table 5B. Encoded CPP protein sequence (SEQ ID NO:18).

```

MAFPYMEAVVGFMLMYIFETYLDVRQHRAKLPLPKTLLEGVISQEKFKEKSRAYSLDKS
HFHFVHEFVTIVTDSTILYFGVLPWFWKSGDFMTIAGFNAENEILHTLAFLAGLMIWSQ
ITDLPFSLYSTFVIEARHGFNKQTPWLFFRDMLKGIFLSVIIGPPIVAIIIVIVQKGGPY
LAIYLWVFTFGLSIVMMTLYPVLIAPLFNKFPLPDGQLREKIEKLASSLNPLKKLFVV
DGSTRSSHSNAYMYGFFKNKRIVPYDTLIRQCKDDEEIVAVIAHELGHWKLNHTVYTFVA
MQILTLLQFGGYTLVRNSADLYRSFGFDTQPVLIGLIIFQHTVIPLQQLVSFGNLVSRS
FEFQADGFAKKLGYESGLRGGLVKLQEENLSAMNTDPWYSAYHYSHPPLVERLAALDEPD
KKED

```

5

The present invention also includes a nucleic acid sequence complimentary to the *Glycine max* CaaX prenyl protease of SEQ ID NO:17. The disclosed complimentary sequence is shown as SEQ ID NO:19.

10 SEQ ID NO:19

```

TTAGTCTTCCTTCTTATCCGGTTGTCCAGCGCGGCCAATCTTCAACAAGGGGAGGAT
GAGAATAGTGATAAGCAGAGTACCAAGGATCTGTATTCTAGCTGACAGATTCTCCTCC
TGTAGTTCACAAAGACCACCGCGTAATCCAGATGCATATCCAAGCTTGGCAAAGCC
ATCAGCCTGAAATTCAAATGATCGGCTGACTAGGTTCAGACCAAAGCTGACCAATTGCT
15 GAAGTGGGATTACAGTATGCTGAAATATGATGAGCCCAATGAGGACTGGCTGCGTATCA
AACCCAAAGCTTCGATAACAGATCAGCTGAATTTCGCACTAGTGTATATCCTCAAATTG
TAGAAGTGTAAAGAATCTGCATAGCAACAAATGTGTACACAGTATGGTTGAGCTTCCAGT
GTCCCAACTCATGGCAATAACAGCAACAATTCTCATCGTCTTGCACGTGTTGAATT
AATGTGTATAAGGGACAATCCTCTTGTCTTGAAGAATCCATACATATAGGCATTGCT
20 GTGACTTGATCTTGTGGATCCATCGACAACAAATAGTTCTTAACGGATAGTTGAGGG
AGGAAGCAAGTTCTGATTTCCTCCCTGAGTTGACCATCTGGAAGTGGAGTGAACCTA
TTGAAGAGTGGAGCTATTAGTACTGGATAAAGGGTCATCATCACAATAGAAAGACCAA

```

CGTAAAAACCAAAGATAGATGCCAAGTATGGACCTCCTTCTGTACTATTACAATGA
 TTGCAGCCACAATAGGTGGACCAATTATTACAGAAAGGAAAATCCTTTAACATGTCC
 CTAAGAATAACCATGGTGTGCTTATTAAAACCATGACGGGCCTCAATCACAAAAGT
 TGAGTACAGAGAAAAGGGCAAATCTGTTATCTGTGACCAAATCATCAGCCCTGCTAAGA
 5 AGGCAAGGGTATGCAGTATTTCATTCTCAGCATTGAAACCAGCTATTGTCAAAATCT
 CCTGATTCTTCCAAAACCAGGGCAATACCCAAAGTACAAAATTGTAGAGTCTGTCAC
 TATTGTCACAAACTCGTAACAAAATGGAAGTGGCTTTATCAAGACTATAGGCTCTAG
 ATTTCTCAAATTCTTGGCTGATAACACCCCTCTAAAGTCTTGGAAAGAGTAGGAAGT
 TTGAGGGCCCTATGTTGTCGCACATCCAAGTAAGTTCAAAAATGTACATTAATATCAT
 10 AAATCCGACAACGGCTCCATGTAGGGAAACGCCAT

Due to the nature of the cloning strategy the sequence presented is not full length but is missing the 5' and 3' non-translated regions. The percent identities of the *Glycine max* nucleotide sequence and its encoded amino acid sequence to that of other CPP sequences as determined by ClustalW analysis are shown in Figure 2.

Using the sequences disclosed herein as hybridization probes, one is able to screen and isolate full length sequences from cDNA or genomic libraries or use the rapid amplification of cDNA ends (RACE) technology or other such PCR techniques. The CPP nucleic acids and amino acids disclosed above have homology to other disclosed CPP sequences (GenBank ID NOs: AL161491 (AT4g01320), AF007269 and AF353722; WO 02/16625 A2). The homology between these and other sequences is shown in the ClustalW alignment analysis shown in Tables 6A-6B.

	<u>Table 6A. ClustalW Nucleic Acid Analysis of CaaX Prenyl Protease</u>	
25	1: PPI-AtCPP	SEQ ID NO:1
	2: PPI-BnCPP	SEQ ID NO:14
	3: PPI-GmCPP	SEQ ID NO:17
	4: BASF_AT1	SEQ ID NO:21
30	5: BASF_AT2	SEQ ID NO:23
	6: BASF-Corn	SEQ ID NO:25
	7: BASF-Gm	SEQ ID NO:27
	8: AFC1	SEQ ID NO:29
	9: AT4g01320	SEQ ID NO:31
35	10: AF007269	SEQ ID NO:33

CLUSTAL W (1.81) multiple sequence alignment

	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
	AF007269	-----
5	PPI-AtCPP	ATGGCGATTCCCTTCATGGAAACCGTCGTGGTAAGCTCAAAACCTTTCTGAGACAT
	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
	PPI-BnCPP	-----
10	BASF-Corn	-----
	PPI-GmCPP	-----
	BASF-Gm	-----
15	AT4g01320	-----
	AF007269	-----
	PPI-AtCPP	TTTACTATCCTGTTCACTCATCGTATTCGTTGGGTTTGCTTCTGTGTTG
	BASF_AT2	-----
	afc1	-----
20	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	-----
25	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
	AF007269	-----
	PPI-AtCPP	TGTGTGTTGAGATTCCATGACTCGTTGTTCATATACCATCGTCTGCTTCTCGTTTC
30	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	-----
35		.
	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
40	AF007269	TAAATTGTTCTTTCTAATAGTGCCTACCTTGATCTGAGGTTTATTACTCCTACTAG
	PPI-AtCPP	-----
	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
45	PPI-BnCPP	-----
	BASF-Corn	-----
50	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
	AF007269	-----
	PPI-AtCPP	TTTCTTGTCTTACTCGTGCCTTGATTGAGCTTATGTGATTTCATCATCTCTTC
55	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	-----
60		.
	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
65	AF007269	CTCGGTTTAGAATGTACGGAGCTTCTGTAAACAAAATCTAGGATTGGGAAGAAAA
	PPI-AtCPP	-----
	BASF_AT2	-----
	afc1	-----

BASF_AT1	-----
PPI-BnCPP	-----
BASF-Corn	-----
 5	
PPI-GmCPP	-----
BASF-Gm	-----
AT4g01320	-----
AF007269	GTCGGAGTCTTTTTCTCATTCCGATTGAAATTGAGAATCTTGAATTTCTT
10 PPI-AtCPP	
BASF_AT2	-----
afc1	-----
BASF_AT1	-----
PPI-BnCPP	-----
15 BASF-Corn	-----
 20	
PPI-GmCPP	-----
BASF-Gm	----- CTAATACGACTCACTATAAGGC
AT4g01320	-----
AF007269	GTTCAAGTCATACAGCTTGAGGTTGGGTTCTGTCAAGGTATTATTATGTTCGTGA
PPI-AtCPP	-----
BASF_AT2	-----
afc1	-----
25 BASF_AT1	
PPI-BnCPP	-----
BASF-Corn	-----
 30	
PPI-GmCPP	-----
BASF-Gm	AAGCAGTGGTAACAACGCAAGTACGCCGGGGAGACGCATGGTCTGAACTAATTGTTA
AT4g01320	-----
AF007269	CTGCAACTAGAGTTCTGGAGTTTGAAATGGGTTTGTGTTGTGAAACCGTATGTG
PPI-AtCPP	-----
BASF_AT2	-----
35 afc1	-----
BASF_AT1	-----
PPI-BnCPP	-----
BASF-Corn	-----
 40	
PPI-GmCPP	-----
BASF-Gm	TAAATAATACCTAAAATTGAGTTGCTCAAACATTGGGTTAACAAATCCAATCTC
AT4g01320	-----
AF007269	AATGTTGCATAAAATCTTCAGTGTCCAATGTTCCATCAGTAGTCAGCACAGAGA
45 PPI-AtCPP	
BASF_AT2	-----
afc1	-----
BASF_AT1	-----
PPI-BnCPP	-----
50 BASF-Corn	-----
 55	
PPI-GmCPP	-----
BASF-Gm	TCAATATAAAACCAATGATCTCACC--CTCACTCCGTTCTGATTCTCACTCTCGTT
AT4g01320	-----
AF007269	TCTTTTATATCTGGTTGATCAAAAAAGTAGATGATGTTATTGAATTTCAGTGATGGAG
PPI-AtCPP	-----
BASF_AT2	-----
afc1	-----
60 BASF_AT1	
PPI-BnCPP	-----
BASF-Corn	-----
 65	
PPI-GmCPP	----- ATGGCGTTCCC--TACATGGAAGCCG
BASF-Gm	TCTCGTTGGTTCATCAGCGTGTCTCAGC-CATGGCGTTCCC--TACATGGAAGCCG
AT4g01320	----- ATGGCGATTCCCT--TTCATGGAACCG
AF007269	TATCTGTTGTTGGCATTAGAGTAGATTCTGATTCTGTTTATTCTTTTC
PPI-AtCPP	----- ATGGCGATTCCCT--TTCATGGAACCG

	BASF_AT2	-----	ATGGCGATTCCCT--TTCATGGAAACCG
	afc1	-----	ATGGCGATTCCCT--TTCATGGAAACCG
	BASF_AT1	-----	ATGGCGATTCCCT--TTCATGGAAACCG
5	PPI-BnCPP	-----	ATGGCGATTCCCT--TTCATGGAAACCG
	BASF-Corn	-----	
	PPI-GmCPP	TTGTCGGATTTATGATATTAATGTACATTTGAAACTTACTTGGATGTGCGACAACATA	
	BASF-Gm	TTGTCGGATTTATGATATTAATGTACATTTGAAACTTACTTGGATGTGCGACAACATA	
10	AT4g01320	TCGTGGGTTTATGATAGTGATGTACATTTGAGACGTATTGGATCTGAGGCAACTCA	
	AF007269	TTACAGGTTTATGATAGTGATGTACATTTGAGACGTATTGGATCTGAGGCAACTCA	
	PPI-AtCPP	TCGTGGGTTTATGATAGTGATGTACATTTGAGACGTATTGGATCTGAGGCAACTCA	
	BASF_AT2	TCGTGGGTTTATGATAGTGATGTACATTTGAGACGTATTGGATCTGAGGCAACTCA	
	afc1	TCGTGGGTTTATGATAGTGATGTACATTTGAGACGTATTGGATCTGAGGCAACTCA	
15	BASF_AT1	TCGTGGGTTTATGATAGTGATGTACATTTGAGACGTATTGGATCTGAGGCAACTCA	
	PPI-BnCPP	TCGTGGGTTTATGATAGTGATGTACATTTGAGACGTATTGGATCTGAGGCAACTCA	
	BASF-Corn	TCGTGGGTTTATGATAGTGATGTACATTTGAGACGTATTGGATCTGAGGCAACTCA	
	PPI-GmCPP	GGGCCCTCAAACCTCCTACTCTTCAAAGACTTAGAGGGTGTATCAGCCAAGAGAAAAT	
	BASF-Gm	GGGCCCTCAAACCTCCTACTCTTCAAAGACTTAGAGGGTGTATCAGCCAAGAGAAAAT	
20	AT4g01320	CTGCTCTCAAGCTCCAACCTCTCCGAAACCTTGGTTGGTGTAAATTAGCCAAGAGAAGT	
	AF007269	CTGCTCTCAAGCTCCAACCTCTCCGAAACCTTGGTTGGTGTAAATTAGCCAAGAGAAGT	
	PPI-AtCPP	CTGCTCTCAAGCTCCAACCTCTCCGAAACCTTGGTTGGTGTAAATTAGCCAAGAGAAGT	
	BASF_AT2	CTGCTCTCAAGCTCCAACCTCTCCGAAACCTTGGTTGGTGTAAATTAGCCAAGAGAAGT	
	afc1	CTGCTCTCAAGCTCCAACCTCTCCGAAACCTTGGTTGGTGTAAATTAGCCAAGAGAAGT	
25	BASF_AT1	CTGCTCTCAAGCTCCAACCTCTCCGAAACCTTGGTTGGTGTAAATTAGCCAAGAGAAGT	
	PPI-BnCPP	CTGCTCTCAAGCTCCAACCTCTCCGAAACACTTGGTTGGTGTAAATTAGCCAAGAGAAGT	
	BASF-Corn	CTGCTCTCAAGCTCCAACCTCTCCGAAACACTTGGTTGGTGTAAATTAGCCAAGAGAAGT	
	PPI-GmCPP	TTGAGAAATCTAGAGCCTATAG-----	
	BASF-Gm	TTGAGAAATCTAGAGCCTATAG-----	
	AT4g01320	TTGAGAAATCACGAGCATACAG-----	
	AF007269	TTGAGAAATCACGAGCATACAG-----	
	PPI-AtCPP	TTGAGAAATCACGAGCATACAG-----	
30	BASF_AT2	TTGAGAAATCACGAGCATACAG-----	
	afc1	TTGAGAAATCACGAGCATACAG-----	
	BASF_AT1	TTGAGAAATCACGAGCATACAG-----	
	PPI-BnCPP	TTGAGAAATCTCGAGCTTACAG-----	
	BASF-Corn	-----	
40	PPI-GmCPP	-----	TCTTGATAAA---AGCCA
	BASF-Gm	-----	TCTTGATAAA---AGCCA
	AT4g01320	-----	GGATATCATCACTGAGAACCTTAATATATGCAGCTA
45	AF007269	TTTAGTTTTATAATTGCCAGGGATATCATCACTGAGAACCTTAATATATGCAGCTA	
	PPI-AtCPP	-----	TCTTGACAAA---AGCTA
	BASF_AT2	-----	TCTTGACAAA---AGCTA
	afc1	-----	TCTTGACAAA---AGCTA
	BASF_AT1	-----	TCTTGACAAA---AGCTA
50	PPI-BnCPP	-----	TCTTGACAAA---AGCTA
	BASF-Corn	-----	
	PPI-GmCPP	CTTCCATTTGTTCACGAGTTGTGACAATAGTGACAGACTCTACAATTGGTACTTGG	
	BASF-Gm	CTTCCATTTGTTCACGAGTTGTGACAATAGTGACAGACTCTACAATTGGTACTTGG	
55	AT4g01320	TTTCACTTTGTTCATGAGTTGTAACTATACTTATGGACTCTGCAATTGGTCTTGG	
	AF007269	TTTCACTTTGTTCATGAGTTGTAACTATACTTATGGACTCTGCAATTGGTCTTGG	
	PPI-AtCPP	TTTCACTTTGTTCATGAGTTGTAACTATACTTATGGACTCTGCAATTGGTCTTGG	
	BASF_AT2	TTTCACTTTGTTCATGAGTTGTAACTATACTTATGGACTCTGCAATTGGTCTTGG	
	afc1	TTTCACTTTGTTCATGAGTTGTAACTATACTTATGGACTCTGCAATTGGTCTTGG	
60	BASF_AT1	TTTCACTTTGTTCATGAGTTGTAACTATACTTATGGACTCTGCAATTGGTCTTGG	
	PPI-BnCPP	TTTCACTTTGTTCATGAGTTGTAACTATACTTATGGACTCTGCAATTGGTCTTGG	
	BASF-Corn	-----	
	PPI-GmCPP	GGTATTGCCCTGGTTTGGAAAG-----	
65	BASF-Gm	GGTATTGCCCTGGTTTGGAAAG-----	
	AT4g01320	GATCTGCCCTGGTTTGGAAAG-----	
	AF007269	GATCTGCCCTGGTTTGGAAAGGTACATATCTGGTTCGGTACAGTATCTCATTGAA	
	PPI-AtCPP	GATCTGCCCTGGTTTGGAAAG-----	

	afcl	AACAA-----	
	BASF_AT1	AACAA-----	
	PPI-BnCPP	AACAA-----	
	BASF-Corn	AACAAG-----	
5		*** * ***	
	PPI-GmCPP	-----CAAACACCAGGTTATTCTTAGGGACA	
	BASF-Gm	-----CAAACACCAGGTTATTCTTAGGGACA	
	AT4g01320	-----CAAACAATATGGATGTTCATAGGGACA	
	AF007269	-----GGATTAAATTGCTTCTAAATTGTTGACAGCAAACAATATGGATGTTCATAGGGACA	
10	PPI-AtCPP	-----CAAACAATATGGATGTTCATAGGGACA	
	BASF_AT2	-----CAAACAATATGGATGTTCATAGGGACA	
	afcl	-----CAAACAATATGGATGTTCATAGGGACA	
	BASF_AT1	-----CAAACAATATGGATGTTCATAGGGACA	
	PPI-BnCPP	-----CAAACAATATGGATGTTCATAGGGACA	
15	BASF-Corn	-----CAAACATATGGCTCTCATAGGGATA	
		***** * ***** * * * * ***** * *	
	PPI-GmCPP	TGCTTAAAGGAATTTCCTTCTGTAAATAATTGGTCCACCTATTGTGGCTGCAATCATTG	
	BASF-Gm	TGCTTAAAGGAATTTCCTTCCGTAAATAATTGGTCCACCTATTGTGGCTGCAATCATTG	
20	AT4g01320	TGATCAAAGGAACATTCCCTCTGTCTGTCAACTAGGCCACCCATTGTTGCTGCGATAATT	
	AF007269	TGATCAAAGGAACATTCCCTCTGTCTGTCAACTAGGCCACCCATTGTTGCTGCGATAATT	
	PPI-AtCPP	TGATCAAAGGAACATTCCCTCTGTCTGTCAACTAGGCCACCCATTGTTGCTGCGATAATT	
	BASF_AT2	TGATCAAAGGAACATTCCCTCTGTCTGTCAACTAGGCCACCCATTGTTGCTGCGATAATT	
	afcl	TGATCAAAGGAACATTCCCTCTGTCTGTCAACTAGGCCACCCATTGTTGCTGCGATAATT	
25	BASF_AT1	TGATCAAAGGAACATTCCCTCTGTCTGTCAACTAGGCCACCCATTGTTGCTGCGATAATT	
	PPI-BnCPP	TGATCAAAGGAACATTCCCTCTGTCTGTCAACTAGGCCACCCATTGTTGCTGCGATAATT	
	BASF-Corn	TGATCAAAGGAATTACTATCCATGATATTGGGCCACCAATGTGGCTGCTATCATCT	
		***** * ***** * * * * ***** *	
30	PPI-GmCPP	TAATAGTACAG-----	
	BASF-Gm	TAATAGTACAG-----	
	AT4g01320	TCATAGTCCAG-----	
	AF007269	TCATAGTCCAGGTTGATGATTCTGGATTCATCTTATTCTGAGTTTTCACATGGATGA	
	PPI-AtCPP	TCATAGTCCAG-----	
35	BASF_AT2	TCATAGTCCAG-----	
	afcl	TCATAGTCCAG-----	
	BASF_AT1	TCATAGTCCAG-----	
	PPI-BnCPP	TTATAGTTCAAG-----	
	BASF-Corn	ACATAGTACAG-----	
40		***** * ***	
	PPI-GmCPP	-----	
	BASF-Gm	-----	
	AT4g01320	-----	
	AF007269	-----CTATTCTCCATTGAGTGTGAGCTCAAAGTTTTAGTTCTGTGTTAAAATTAAAATT	
45	PPI-AtCPP	-----	
	BASF_AT2	-----	
	afcl	-----	
	BASF_AT1	-----	
	PPI-BnCPP	-----	
50	BASF-Corn	-----	
	PPI-GmCPP	-----AAAGGAGGTCCATACTTGGCATT	
	BASF-Gm	-----AAAGGAGGTCCATACTTGGCATT	
	AT4g01320	-----AAAGGAGGTCCATTCTTGCCATT	
55	AF007269	-----TGCTTCTCTGAGCATGAAGTTCTATTTTCCAGAAAGGAGGTCTTATCTTGCCATT	
	PPI-AtCPP	-----AAAGGAGGTCCATTCTTGCCATT	
	BASF_AT2	-----AAAGGAGGTCCATTCTTGCCATT	
	afcl	-----AAAGGAGGTCCATTCTTGCCATT	
	BASF_AT1	-----AAAGGAGGTCCATTCTTGCCATT	
60	PPI-BnCPP	-----AAAGGAGGTCCATTACCTCGCCATT	
	BASF-Corn	-----ATTGGAGGACCTTACCTGGCTATA	
		***** * * * * * * * * * * * * * *	
65	PPI-GmCPP	TATCTTGGGTTTTACGTTGGCTTCTATTGTGATGATGACCTTATCCAGTACTA	
	BASF-Gm	TATCTTGGGTTTTACGTTGGCTTCTATTGTGATGATGACCTTATCCAGTACTA	
	AT4g01320	TATCTGTGGGCATTCTATGTTATCCTGTCTAGTGTGATGACTATACCCGGCTTG	
	AF007269	TATCTGTGGGCATTCTATGTTATCCTGTCTAGTGTGATGACTATACCCGGCTTG	
	PPI-AtCPP	TATCTGTGGGCATTCTATGTTATCCTGTCTAGTGTGATGACTATACCCGGCTTG	
	BASF_AT2	TATCTGTGGGCATTCTATGTTATCCTGTCTAGTGTGATGACTATACCCGGCTTG	

	afc1	TATCTGTGGGCATTCACTGTTATCCTGCTCTAGTGATGATGACTATATACCCGGTCTTG
	BASF_AT1	TATCTGTGGGCATTCACTGTTATCCTGCTCTAGTGATGATGACTATATACCCGGTCTTG
	PPI-BnCPP	TATCTGTGGGCATTCACTGTTATCCTGCTCTAGTGATGATGACTATATACCCGGTCTTG
5	BASF-Corn	TATCTGTGGGGTATGTTGATTAGCTACTGATGATGACAATATACCCATTGTC
	*****	*****
	PPI-GmCPP	ATAGCTCCACTCTCAATAAGTTCACTCCA-----
	BASF-Gm	ATAGCTCCACTCTCAATAAGTTCACTCCA-----
10	AT4g01320	ATAGCACCCTCTCAACAAAGTTCACTCCTGTGTATTCTGTATGCCATTAAACAA
	AF007269	ATAGCACCCTCTCAACAAATTCACTCCT-----
	PPI-AtCPP	ATAGCACCCTCTCAACAAAGTTCACTCCT-----
	BASF_AT2	ATAGCACCCTCTCAACAAAGTTCACTCCT-----
	afc1	ATAGCACCCTCTCAACAAAGTTCACTCCT-----
15	BASF_AT1	ATAGCACCCTCTCAACAAAGTTCACTCCT-----
	PPI-BnCPP	ATTCACCTCTTCACAAAGTTCACTCCT-----
	BASF-Corn	ATAGCTCCTCTGTTCAACAAAGTTCACTCCT-----
	*****	*****
	PPI-GmCPP	-----
	BASF-Gm	-----
20	AT4g01320	TTCACTGTTGTTGCATATGTTTACAGACAAATATAATCTCCGTTTTATGGCT-----
	AF007269	-----
	PPI-AtCPP	-----
	BASF_AT2	-----
	afc1	-----
25	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	-----
	*****	*****
	PPI-GmCPP	-----CTTCCAGATGGTCAACTCAGGGAGAAAATCGAGAAACTTGCTTCCTCCCTCAACTA
30	BASF-Gm	-----CTTCCAGATGGTCAACTCAGGGAGAAAATCGAGAAACTTGCTTCCTCCCTCAACTA
	AT4g01320	-----CTTCCAGATGGAGACCTCCGGGAGAAGATTGAGAAACTTGCTTCTCTCAAAGTT
	AF007269	ATAGCTTCCAGATGGAGACCTCCGGGAGAAGATTGAGAAACTTGCTTCTCTCAAAGTT
	PPI-AtCPP	-----CTTCCAGATGGAGACCTCCGGGAGAAGATTGAGAAACTTGCTTCTCTCAAAGTT
	BASF_AT2	-----CTTCCAGATGGAGACCTCCGGGAGAAGATTGAGAAACTTGCTTCTCTCAAAGTT
35	afc1	-----CTTCCAGATGGAGACCTCCGGGAGAAGATTGAGAAACTTGCTTCTCTCAAAGTT
	BASF_AT1	-----CTTCCAGATGGAGACCTCCGGGAGAAGATTGAGAAACTTGCTTCTCTCAAAGTT
	PPI-BnCPP	-----CTTCCAGATGGAGACCTCCGGGAGAAGATTGAGAAACTTGCTTCTCTCAAAGTT
	BASF-Corn	-----CTTCCAGATGGAGACCTCCGGGAGAAGATTGAGAAACTTGCTTCTCTCAAAGTT
	*****	*****
40	PPI-GmCPP	TCCGTTAAAGAAACTATTGTTGTCGATGGATCCACAAGATCAAGTCACAGCAATG-----
	BASF-Gm	TCCGTTAAAGAAACTATTGTTGTCGATGGATCCACAAGATCAAGTCACAGCAATG-----
	AT4g01320	TCCTTGAGAGAACGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATG-----
	AF007269	TCCTTGAGAGAACGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATG-----
45	PPI-AtCPP	TCCTTGAGAGAACGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATG-----
	BASF_AT2	TCCTTGAGAGAACGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATG-----
	afc1	TCCTTGAGAGAACGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATG-----
	BASF_AT1	TCCTTGAGAGAACGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATG-----
	PPI-BnCPP	TCCTCTGAGAGAACGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGTAATG-----
50	BASF-Corn	TCCTTGAGAGAACGCTGTTGTCGATGGATCTACAAGGTCAAGCCACAGTAATG-----
	*****	*****
	PPI-GmCPP	-----
	BASF-Gm	-----
55	AT4g01320	AAGCTTGAGATCTCTCCTACCTACTTACTCTAGTTACCATAGAAGCTACGTATCT-----
	AF007269	-----
	PPI-AtCPP	-----
	BASF_AT2	-----
	afc1	-----
60	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	-----
	*****	*****
	PPI-GmCPP	-----CCTATATGTATGGATTCTCAAGAACAGAGGATTGTCCTTAT
	BASF-Gm	-----CCTATATGTATGGATTCTCAAGAACAGAGGATTGTCCTTAT
65	AT4g01320	-----CTTACATGTATGGTTCTTAAGAACAAAAGGATTGTTCTTAT
	AF007269	TGTTACATCATACAGGCTTACATGTATGGTTCTTAAGAACAAAAGGATTGTTCTTAT
	PPI-AtCPP	-----CTTACATGTATGGTTCTTAAGAACAAAAGGATTGTTCTTAT
	BASF_AT2	-----CTTACATGTATGGTTCTTAAGAACAAAAGGATTGTTCTTAT

	afc1	-----CTTACATGTATGGTTCTTAAGAACAAAAGGATTGTTCTTAT
	BASF_AT1	-----CTTACATGTATGGTTCTTAAGAACAAAAGGATTGTTCTTAT
	PPI-BnCPP	-----CTTACATGTATGGTTCTCAAGAACAAAAGGATTGTTCTTAT
	BASF-Corn	-----CCTACATGTATGGTTCTCAAGAACAAAGCGCATAGTACTCTAT
5		* *
	PPI-GmCPP	GACACATTAATTCAACAG-----
	BASF-Gm	GACACATTAATTCAACAG-----
	AT4g01320	GATACTGATTGATTCAGCAG-----
	AF007269	GATACTGATTGATTCAGCAGGTACTGTGACTCTGATGCTCAAACGAGCTATACTCACATT
10	PPI-AtCPP	GATACTGATTGATTCAGCAG-----
	BASF_AT2	GATACTGATTGATTCAGCAG-----
	afc1	GATACTGATTGATTCAGCAG-----
	BASF_AT1	GATACTGATTGATTCAGCAG-----
	PPI-BnCPP	GACACATTGATTCAGCAG-----
15	BASF-Corn	GACACATTGATTCAGCAG-----
		* *
	PPI-GmCPP	-----TGCAAAGACGATGAGG
	BASF-Gm	-----TGCAAAGACGATGAGG
20	AT4g01320	-----TGCAAGAACATGAGGATG
	AF007269	TCTGTTCTGGTCTGAAACATAACATAATCTCTATTGTGCAAGAACATGAGGATG
	PPI-AtCPP	-----TGCAAGAACATGAGGATG
	BASF_AT2	-----TGCAAGAACATGAGGATG
	afc1	-----TGCAAGAACATGAGGATG
25	BASF_AT1	-----TGCAAGAACATGAGGATG
	PPI-BnCPP	-----TGCCAGAACATGAGGATG
	BASF-Corn	-----TGTAGCAATGAGGATG
		* *
30	PPI-GmCPP	AAATTGTTGCTGTTATTGCCCATGAGTTGGGACACTGGAAGCTAACCAACTGTGTACA
	BASF-Gm	AAATTGTTGCTGTTATTGCCCATGAGTTGGGACACTGGAAGCTAACCAACTGTGTACA
	AT4g01320	AAATTGTTGCGGTTATTGCACACGAGCTGGACATTGAAACTGAATCACACTACACT
	AF007269	AAATTGTTGCGGTTATTGCACACGAGCTGGACATTGAAACTGAATCACACTACACT
	PPI-AtCPP	AAATTGTTGCGGTTATTGCACACGAGCTGGACATTGAAACTGAATCACACTACACT
35	BASF_AT2	AAATTGTTGCGGTTATTGCACACGAGCTGGACATTGAAACTGAATCACACTACACT
	afc1	AAATTGTTGCGGTTATTGCACACGAGCTGGACATTGAAACTGAATCACACTACACT
	BASF_AT1	AAATTGTTGCGGTTATTGCACACGAGCTGGACATTGAAACTGAATCACACTACACT
	PPI-BnCPP	AAATTGTTGCGGTTATTGCACACGAGCTGGACACTGGAAGCTGAATCACACTACACT
	BASF-Corn	AGATAGTTCTGTTATAGCACATGAACATTGGACACTGGAAGCTCAATCATACTGTCTATT
40		* *
	PPI-GmCPP	CATTTGTTGCTATGCAG-----
	BASF-Gm	CATTTGTTGCTATGCAG-----
	AT4g01320	CGTCATTGCAAGTCAAGTGAGGCTAACCGACAGTTCAAAACTTACTCACATCTACAT
	AF007269	CGTCATTGCAAGTCAAA-----
45	PPI-AtCPP	CGTCATTGCAAGTCAA-----
	BASF_AT2	CGTCATTGCAAGTCAA-----
	afc1	CGTCATTGCAAGTCAA-----
	BASF_AT1	CGTCATTGCAAGTCAA-----
	PPI-BnCPP	CGTCATTGCTGTTCAA-----
50	BASF-Corn	CCTTGTAGCTGTCCAG-----
		* *
	PPI-GmCPP	-----ATTCTTACA
	BASF-Gm	-----ATTCTTACA
	AT4g01320	-----ATCCTTGCC
55	AF007269	TTCACTTAAGAACATGTCTTATGACCCTCTCTCAATGTTGCTTGAGATCCTTGCC
	PPI-AtCPP	-----ATCCTTGCC
	BASF_AT2	-----ATCCTTGCC
	afc1	-----ATCCTTGCC
	BASF_AT1	-----ATCCTTGCC
60	PPI-BnCPP	-----ATCCTTGCC
	BASF-Corn	-----CTGCTTATG
		* * * * *
65	PPI-GmCPP	CTTCTACAATTGGAGGATACACTAGTGCAGCTGATCTGTATCGAACGCTT
	BASF-Gm	CTTCTACAATTGGAGGATACACTAGTGCAGCTGATCTGTATCGAACGCTT
	AT4g01320	TTCTTACAATTGGAGGATACACTCTGTGAGAACACTCCACTGATCTCTCAGGAGTTTC
	AF007269	TTCTTACAATTGGAGGATACACTCTGTGAGAACACTCCACTGATCTCTCAGGAGTTTC
	PPI-AtCPP	TTCTTACAATTGGAGGATACACTCTCTCAGAACACTCCACTGATCTCTCAGGAGTTTC

	BASF_AT2	TTCTTACAATTGGAGGATACACTCTTGTCAAGAAACTCCACTGATCTTCAGGAGTTTC
	afc1	
	BASF_AT1	TTCTTACAATTGGAGGATACACTCTTGTCAAGAAACTCCACTGATCTTCAGGAGTTTC
5	PPI-BnCPP	TTCTTGCATAATTGGAGGATACACTCTTGTCAAGAAACTCCACTGATCTTCAGGAGTTTC
	BASF-Corn	TTTCTCAATTGGAGGATATACTCTAGTAAGGAGCTCAAAGATCTATTGGAAGTTT
		* * ***** *
10	PPI-GmCPP	GGGTTTGATACGCAGCAGTCCTCATGGGCTCATCATATTCAG-----
	BASF-Gm	GGGTTTGATACGCAGCAGTCCTCATGGGCTCATCATATTCAG-----
	AT4g01320	GGATTTGATACACAGCTGTTCTCATGGTTGATCATATTCAG-----
	AF007269	GGATTTGATACACAGCTGTTCTCATGGTTGATCATATTCAG-----
	PPI-AtCPP	GGATTTGATACACAGCTGTTCTCATGGTTGATCATATTCAG-----
	BASF_AT2	GGATTTGATACACAGCTGTTCTCATGGTTGATCATATTCAG-----
15	afc1	GGATTTGATACACAGCTGTTCTCATGGTTGATCATATTCAG-----
	BASF_AT1	GGATTTGATACACAGCTGTTCTCATGGTTGATCATATTCAG-----
	PPI-BnCPP	GGTTTGATACACAACCAGTTCTCATGGTTGATCATATTCAG-----
	BASF-Corn	GGCTTCAGGACAGCCAGTAATAATTGGATTGATCATTTCGG-----
		* *
20	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
	AF007269	CTTTTGACACTAATCTAATGAATCAAGGATGGATTAAGAAAAAAAACCTAAACCTTTG
25	PPI-AtCPP	-----
	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	-----
30	PPI-GmCPP	----- CATACTGTAATCCCACCTTCAGCAATTGGTCAGC
	BASF-Gm	----- CATACTGTAATCCCACCTTCAGCAATTGGTCAGC
	AT4g01320	----- CACACTGTAATACCAACTGCAACATCTAGTAAGC
	AF007269	GTTATATCTCCTGTCGATTATCACAGCACACTGTAATACCAACTGCAACATCTAGTAAGC
35	PPI-AtCPP	----- CACACTGTAATACCAACTGCAACATCTAGTAAGC
	BASF_AT2	----- CACACTGTAATACCAACTGCAACATCTAGTAAGC
	afc1	----- CACACTGTAATACCAACTGCAACATCTAGTAAGC
	BASF_AT1	----- CACACTGTAATACCAACTGCAACATCCAGTAAGC
	PPI-BnCPP	----- CACACTGTAATACCAACTTCACACACCTAGTAAGC
40	BASF-Corn	----- CACACCATATAACCCATCCAACACACCTCTGAGC
		* *
45	PPI-GmCPP	TTTGGTCTGAACCTAGTCAGCGATCATTGAAATTTCAGG-----
	BASF-Gm	TTTGGTCTGAACCTAGTCAGCGATCATTGAAATTTCAGG-----
	AT4g01320	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----
	AF007269	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----
	PPI-AtCPP	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----
	BASF_AT2	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----
	afc1	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----
	BASF_AT1	TTTGGCCTCAACCTTGTAGTCGAGCGTTGAGTTTCAGG-----
50	PPI-BnCPP	TTTGACCTCAACCTTGTAGTCGAGCGTTGAGTTTCAGG-----
	BASF-Corn	TTTCGCCTGAACCTTGTCAAGCAGAGCATTGAAATTTCAGG-----
		* *
55	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
	AF007269	AGATCCAACCATAGTTCTTATTGCAATGGCAGCCTCATCTACTAATCTGAGTTAACGT
	PPI-AtCPP	-----
	BASF_AT2	-----
	afc1	-----
60	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	-----
65	PPI-GmCPP	----- CTGATGGCTTGGCCAAGAACGCTTGGATATGCATCTGGATTACCGGGTG
	BASF-Gm	----- CTGATGGCTTGGCCAAGAACGCTTGGATATGCATCTGGATTACCGGGTG
	AT4g01320	----- CTGATGCTTGTGCTGTGAAGCTTGGCTATGCAAAAGATCTTCGGTCTG
	AF007269	TCCTTTGCAGGCTGATGCTTGTGAAGCTTGGCTATGCAAAAGATCTTCGGTCTG
	PPI-AtCPP	----- CTGATGCTTGTGCTGTGAAGCTTGGACTATGCAAAAGATCTTCGGTCTG

	BASF_AT2	-----CTGATGCTTTGCTGTGAAGCTGGCTATGCAAAAGATCTCGTCCTG
	afc1	-----CTGATGCTTTGCCGTGAAGCTGGCTATGCAAAAGATCTCGTCCTG
	BASF_AT1	-----CTGATGCTTTGCTGTGAAGCTGGCTATGCAAAAGATCTCGTCCTA
5	PPI-BnCPP	-----CTGATGCTTTGCCAGTGAATCTGGTTATGCAAAGGATCTACGTCCTG
	BASF-Corn	-----CTGATGCTTTGCCAAGAACCTGGATATGCCCTCAGCTCCGAGCAG ***** * *** * *** * *** * *** * *** *
	PPI-GmCPP	GTCTTGTGAAACTACAGG-----
	BASF-Gm	GTCTTGTGAAACTACAGG-----
10	AT4g01320	CTCTAGTGAAACTACAGGTCAAGAGAATAACAACAGAACACAAACTGTTACCTCAATT
	AF007269	CTCTAGTGAAACTACAGGTCAAGAGAATAACAACAGAACACAAACTGTTACCTCAATT
	PPI-AtCPP	CTCTAGTGAAACTACAGG-----
	BASF_AT2	CTCTAGTGAAACTACAGG-----
	afc1	CTCTAGTGAAACTACAGG-----
15	BASF_AT1	CTCTAGTGAAACTACAGG-----
	PPI-BnCPP	CCCTAGTGAAGCTACAGG-----
	BASF-Corn	CCCTTGTAAACTACAGG----- *** * *** * *** * ***
	PPI-GmCPP	-----AGGAGAACCTGTCAGCTA
20	BASF-Gm	-----AGGAGAACCTGTCAGCTA
	AT4g01320	GTGTCACACACTTAAATGGATTTTGTGGGATTTGCAGGAAGAGAACCTATCAGCAA
	AF007269	GTGTCACACACTTAAATGGATTTTGTGGGATTTGCAGGAAGAGAACCTATCAGCAA
	PPI-AtCPP	-----AAGAGAACCTATCAGCAA
	BASF_AT2	-----AAGAGAACCTATCAGCAA
25	afc1	-----AAGAGAACCTATCAGCAA
	BASF_AT1	-----AAGAGAACCTATCAGCAA
	PPI-BnCPP	-----AAGAGAACCTATCAGCGA
	BASF-Corn	-----AGGAGAACCTGTCAGCTA ***** * *** * ***
30	PPI-GmCPP	TGAATACAGATCCTGGTACTCTGCTTATCACTATTCTCATCCTCCCCTGTTGAAAGAT
	BASF-Gm	TGAATACAGATCCTTGCT--CGTGCCTG-----
	AT4g01320	TGAACACTGATCCATTGTAACAGCTTATCACTACTCACATCCTCCTGTTGAAAGGC
	AF007269	TGAACACTGATCCATTGTAACAGCTTATCACTACTCACATCCTCCTGTTGAAAGGC
35	PPI-AtCPP	TGAACACTGATCCATTGTAACAGCTTATCACTACTCACATCCTCCTGTTGAAAGGC
	BASF_AT2	TGAACACTGATCCATTGTAACAGCTTATCACTACTCACATCCTCCTGTTGAAAGGC
	afc1	TGAACACTGATCCATTGTAACAGCTTATCACTACTCACATCCTCCTGTTGAAAGGC
	BASF_AT1	TGAACACTGATCCATTGTAACAGCTTATCACTACTCACATCCTCCTGTTGAAAGGC
40	PPI-BnCPP	TGAACACAGACCCATTGTAACAGCTTATCACTACTCACACCCCTCCTGTTGAAAGGC
	BASF-Corn	TGAACACCGATCCTGGTATTGGCATATCACTACTCCCACCCACACTCGTCGAGAGGC ***** * *** * ***
	PPI-GmCPP	TGGCCGCGCTGGACGA---ACCGGATAAGAAGGAAGACTAA-----
	BASF-Gm	-----
45	AT4g01320	TTCGAGCCATTGATGG---AGAAGACAAGAACAGACAGATTAA-----
	AF007269	TTCGAGCCATTGATGG---AGAAGACAAGAACAGACAGATTAA-----
	PPI-AtCPP	TTCGAGCCATTGATGG---AGAAGACAAGAACAGACAGATTAA-----
	BASF_AT2	TTCGAGCCATTGATGG---AGAAGACAAGAACAGACAGATTAA-----
	afc1	TTCGAGCCATTGATGG---AGAAGACAAGAACAGACAGATTAA-----
50	BASF_AT1	TTCGAGCCATTGATGG---AGAAGACAAGAACAGACAGATTAA-----
	PPI-BnCPP	TTCGAGCCATTGATGG---AGAAGACAAGAACAGACAGATTAA-----
	BASF-Corn	TGCAAGCTTGGAGATTACAGACAAAAAGAAGATTAGTCGATCCTGTATGAGGTT
55	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
	AF007269	-----
	PPI-AtCPP	-----
60	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	TACATATGGATTTCCCTGCCACATGCACACCGATTCACTGCTGGATGGTGAGGTTT
65	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----

	AF007269	-----
	PPI-AtCPP	-----
	BASF_AT2	-----
	afc1	-----
5	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	TGACATAGGAGTGTCAAAGCTTAGAGTGCATCTTCGGTCAGGTGCAACAGCCTT
10	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
	AF007269	-----
	PPI-AtCPP	-----
15	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	CGGTCAATTGAGACATATAAGCGAATTAGCTATTAAAAAAAACAGAACTGTTGCATCAAA
20	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
25	AF007269	-----
	PPI-AtCPP	-----
	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
30	PPI-BnCPP	-----
	BASF-Corn	AAAAAAAAAAAAAGAAACAAAAAAGAAAAAAGAAAAAAGAAAAAAGAAAAAAGAAAAA
35	PPI-GmCPP	-----
	BASF-Gm	-----
	AT4g01320	-----
	AF007269	-----
	PPI-AtCPP	-----
40	BASF_AT2	-----
	afc1	-----
	BASF_AT1	-----
	PPI-BnCPP	-----
	BASF-Corn	AAAAAGTGCTCTGCGTTGTTACCACTGCTGCCCTATAGTGATCGTATCAGA

45

Table 6B. ClustalW Amino Acid Analysis of CaaX Prenyl Protease

1:	PPI-AtCPP	SEQ ID NO:2
2:	PPI-BnCPP	SEQ ID NO:15
50	3: PPI-GmCPP	SEQ ID NO:18
	4: BASF_AT1	SEQ ID NO:22
	5: BASF_AT2	SEQ ID NO:24
	6: BASF-Corn	SEQ ID NO:26
	7: BASF-Gm	SEQ ID NO:28
55	8: AFC1	SEQ ID NO:30
	9: AT4g01320	SEQ ID NO:32
	10: AF007269	SEQ ID NO:34
60	PPI-GmCPP	MAFPYMEAVVGFIMILMYIFETYLDVRQHRAKLPTLPKTLEG-----VISQEKFESR
	BASF-Gm	MAFPYMEAVVGFIMILMYIFETYLDVRQHRAKLPTLPKTLEG-----VISQEKFESR
	AF007269	MAIPFMETVVGFMIVMYIFETYLDLRQLTALKLPTLPKTLI-----
	AT4g-AtCPP	MAIPFMETVVGFMIVMYIFETYLDLRQLTALKLPTLPKTLVGVISQEKFESRAYRDIIT

	.:.*:.*:	**:***:***:***
	<hr/>	
	PPI-GmCPP BASF-Gm AF007269 AT4g-AtCPP BASF_AT2 AFC1 BASF_AT1 PPI-AtCPP PPI-BnCPP BASF-Corn	LNLVSRSFEFQADGFAKKLGYASGLRG----- LNLVSRSFEFQADGFAKKLGYASGLRG----- LNLVSRAFEFQADAFAVKLGYAKDLR-----PALV---KLQVREDNNRTQ----- LNLVSRAFEFQADAFAVKLGYAKDLR-----PALV---KLQVREDNNRTQTVTSICV LNLVSRAFEFQADAFAVKLGYAKDLR-----PALV---KLQE----- LNLVSRAFEFQADAFAVKLGYAKDLR-----PALVKLQE----- LNLVSRAFEFQADAFAVKLGYAKDLRPTLVKLQ----- LNLVSRAFEFQADAFAVKLDYAKDLRPALVKLQ----- LNLVSRAFEFQADAFAVNLGYAKDLRP----- LNLVSRAFEFQADAFAKNLGYAPQLR----- *****:*****:*** :*** ***
5	<hr/>	
10	<hr/>	
15	<hr/>	
20	<hr/>	
25	<hr/>	

Example 4: Plant Transformation

30 *Arabidopsis* transgenic plants were made by the method of dipping flowering plants into an *Agrobacterium* culture, based on the method of Andrew Bent in, Clough SJ and Bent AF, 1998. Floral dipping: a simplified method for *Agrobacterium*-mediated transformation of *Arabidopsis thaliana*. Wild type plants were grown under standard conditions until the plant has both developing flowers and open flowers. The plant was inverted for 2 minutes into a solution of *Agrobacterium* culture carrying the appropriate gene construct. Plants were then left horizontal in a tray and kept covered for two days to maintain humidity and then righted and bagged to continue growth and seed development. Mature seed was bulk harvested.

Transformed T1 plants were selected by germination and growth on MS plates containing 50 µg/ml kanamycin. Green, kanamycin resistant (Kan^R) seedlings were identified after 2 weeks growth and transplanted to soil. Plants were bagged to ensure self fertilization and the T2 seed of each plant harvested separately. During growth of T1 plants leaf samples were harvested, DNA extracted and Southern blot and PCR analysis performed.

T2 seeds were analysed for Kan^R segregation. From those lines that showed a 3:1 resistant phenotype, surviving T2 plants were grown, bagged during seed set, and T3 seed harvested from each line. T3 seed was again used for Kan^R segregation analysis and those lines showing 100% Kan^R phenotype were selected as homozygous lines. Further 5 molecular and physiological analysis was done using T3 seedlings.

Transgenic *Brassica napus*, *Glycine max* and *Zea maize* plants were produced using *Agrobacterium* mediated transformation of cotyledon petiole tissue. Seeds were sterilized as follows. Seeds were wetted with 95% ethanol for a short period of time such as 15 seconds. Approximately 30 ml of sterilizing solution I was added (70% Javex , 10 100µl Tween20) and left for approximately 15 minutes. Solution I was removed and replaced with 30 ml of solution II (0.25% mecuric chloride, 100µl Tween20) and incubated for about 10 minutes. Seeds were rinsed with at least 500 ml double distilled sterile water and stored in a sterile dish. Seeds were germinated on plates of $\frac{1}{2}$ MS medium, pH 5.8, supplemented with 1% sucrose and 0.7% agar. Fully expanded 15 cotyledons were harvested and placed on Medium I (Murashige minimal organics (MMO), 3% sucrose, 4.5 mg/L benzyl adenine (BA), 0.7% phytoagar, pH5.8). An *Agrobacterium* culture containing the nucleic acid construct of interest was grown for 2 days in AB Minimal media. The cotyledon explants were dipped such that only the cut portion of the petiole is contacted by the *Agrobacterium* solution. The explants were then 20 embedded in Medium I and maintained for 5 days at 24°C, with 16,8 hr light dark cycles.

Explants were transferred to Medium II (Medium I, 300 mg/L timentin,) for a further 7 days and then to Medium III (Medium II, 20 mg/L kanamycin). Any root or shoot tissue which had developed at this time was dissected away. Transfer explants to fresh plates of Medium III after 14 -21 days. When regenerated shoot tissue developed 25 the regenerated tissue was transferred to Medium IV (MMO, 3% sucrose, 1.0% phytoagar, 300 mg/L timentin, 20 mg/L 20 mg/L kanamycin). Once healthy shoot tissue developed shoot tissue dissected from any callus tissue was dipped in 10X IBA and transferred to Medium V (Murashige and Skooge (MS), 3% sucrose, 0.2 mg/L indole butyric acid (IBA), 0.7% agar, 300 mg/L timentin, 20 mg/L 20 mg/L kanamycin) for 30 rooting. Healthy plantlets were transferred to soil. The above method, with or without modifications, is suitable for the transformation of numerous plant species including *Glycine max*, *Zea maize* and cotton.

Transgenic *Glycine max*, *Zea maize* and cotton can be produced using *Agrobacterium*-based methods which are known to one of skill in the art. Alternatively one can use a particle or non-particle biolistic bombardment transformation method. An example of non-particle biolistic transformation is given in U.S. Patent Application 5 20010026941. This method has been used to produce transgenic *Glycine max* and *Zea maize* plants. Viable plants are propagated and homozygous lines are generated. Plants are tested for the presence of drought tolerance, physiological and biochemical phenotypes as described elsewhere.

10 The following table identifies the constructs and the species which they have been transformed.

Table 7 Transformation List

<u>SEQ ID NO:</u>	<u>Construct</u>	<u>Species Transformed</u>
4	pBII121-AtCPP	<i>A. thaliana</i> , <i>B. napus</i>
5	pBII121-HP-AtCPP	<i>A. thaliana</i>
15	pRD29A-AtCPP	<i>A. thaliana</i> , <i>B. napus</i>
36	pRD29A-HP-AtCPP	<i>A. thaliana</i>
37	MuA-AtCPP	<i>Glycine max</i> , <i>Zea mays</i>

Non-limiting examples of vector constructs suitable for plant transformation are given in 20 SEQ ID NO: 4, 5, 35-53.

SEQ ID NO:4

```
gttacccggcaatatacctgtcaaacactgatagttaaactgaaggcgaaacga
caatctgatcatgagcggagaattaaggagtcacgttatgaccccgccatgacgcg
ggacaagccgtttacgttggactgacagaaccgcaacgttgaaggagccactcagc
25 cgcgggtttctggagttaatgagctaaggcacatacgtcagaaaccattattgcgcgtt
caaaagtgcctaaggtcactatcagcttagcaaataatttcttgcaaaaatgctccact
gacgttccataaattcccctcggtatccaatttagagtctcatattcactctcaatccaa
ataatctgcaccggatctggatcggttcgcattgttgcattgaacaagatggattgcacgcagg
ttctccggccgcttgggtggagaggctattcggtatgactggcacaacagacaatcg
30 gctgctctgatgcccggtgttccggctgtcagcgcaggggcgccgggtttttgtc
aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcgtg
gctggccacgacggcggttcctgcgcagctgtgctcgcacgttgtcactgaagcggaa
```

gggactggctgttattggcgaaagtgccggggcaggatctcctgtcatctcaccttgct
cctgcccagaaaagtatccatcatggctgtatgcataatgcggcggctgcatacgcttgatcc
ggctacacctgcccattcgaccaccaagcgaaacatcgcatcgagcgagcacgtactcgga
tggaaagccggctttgtcgatcaggatgtatggacgaagagcatcagggctcgccca
5 gccgaactgttcgcaggctcaaggcgcatgcggcgtatgtatctcgatcgac
ccatggcgatgcctgttgcgaatatcatggtaaaaatggccgctttctggattca
tcgactgtggccggctgggtgtggcggaccgctatcaggacatagcgttggctaccgat
gatattgctgaagagcttggcggcgaatggctgaccgcctcgtgcttacggtat
cgccgctcccgattcgcatcgccatcgcccttatcgcccttgcacgagttttctgag
10 cgggactctgggttcgaaatgaccgaccaagcgacgccccaacctgcacgagatt
tcgattccaccgcgccttatgaaagggtggcttcggaatcgccccggacgccc
ggctggatgatcctccagcgccggatctcatgctggagttctcgccacggatctc
tgccgaacaggcggtcgaaggtgccatattacgacagcaacggccgacaagcaca
acgcccacgatcctgagcgacaaatatgatcggcccccgtccacatcaacggcgtcggc
15 gggactgcccaggcaagaccgagatgcaccgcgatattttgtcggtcgatatttt
cgtggagttccgcacagaccggatgtatccgcgttcaaacatttggcaataaa
gtttcttaagattgaatcctgttgcggcttgcgtatgattatcatataatttctgtt
aattacgttaagcatgtataattacatgtaatgcacgttattttagatgggt
tttatgatttagagtcccgaattatacattaaacgcataaaaaacaaaatatacg
20 gcgcaaactaggataaattatcgccgcgggtgtcatctatgttactagatcgccctcc
tgtcaatgctggcggcggctctgggtggttctggcggctctgaggaggcggttccgggtggct
ctgagggtggcggtctgagggtggcggctctgaggaggcggttccgggtggct
gggtccgggtgattttgattatgaaaagatggcaaacgctaataagggggctatgaccga
aaatgccatgaaaacgcgtacagtctgacgctaaaggcaaacttgattctgtcgcta
25 ctgattacggtgctgtatcgatggttcattggtgacgttccggccttgcataatgg
aatggtgctactggtagttgtctggcttaattccaaatggctcaagtggcgtacgg
tgataattcaccttaatgaataattccgtcaatattacctccctccctcaatcg
ttgaatgtcgccctttgtcttggccaatacgcaaaccgcctctcccgcggtgg
ccgattcattaatgcagctggcacgacaggttccgactggaaagcggcagtgagcg
30 caacgcataatgtgagttagctcactcattaggcacccaggcttacactttatgc
ttccggctcgatgttgttggaaattgtgagcgataacaattcacacaggaaacagc
tatgaccatgattacgccaagcttgcacgcccacagatggtagagaggctt
acgcagcaggctcatcaagacatctaccgagcaataatctccagggaaatcaaatac
cttcccaagaaggtaaagatgcagtcataaaagattcaggactaactgcataagaacac

agagaaaagatataatttctcaagatcagaagtactattccagtagtgacgattcaaggct
tgcttcacaaaccaggcaagtaatagagattggagttctctaaaaaggtagttcccact
gaatcaaaggccatggagtcaaagattcaaatagaggacctaacagaactcgccgtaaa
gactggcgaacagttcatacagagtcttacgactcaatgacaagaagaaaatcttcg
5 tcaacatggtgagcacgacacacttgtctactccaaaaatatacaagatacagtctca
gaagaccaaaggcaattgagactttcaacaaaggtaatatccggaaacctcctcgg
attccattgccagctatctgtcactttattgtgaagatagtggaaaaggaaggtagggct
cctacaaatgccatcattgcataaaggaaaggccatcgtaagatgcctctggcag
agtggtcccaaagatggacccccacccacgaggagcatcgtaaaaaagaagacgttcc
10 aaccacgtcttcaaagcaagtggattgtatgtatctccactgacgtaagggatgacg
cacaatcccactatccttcgcaagacccttccttatataaggaagttcattcatttgc
gagagaacacggggactctagaggatccatggcgattccttcatggaaaccgtcg
ggtttatgatagtgtatgtacatttttagacgtattggatctgaggcaactcactgc
tctcaagcttccaactctccgaaaaccttgggtggtaattagccaagagaagtttgc
15 agaaaatcacgagcatacagtcttgacaaaagctatttcactttgtcatgagttgt
actatacttatggactctgcaattttgttcttggatcttcgttgggtttggaaagat
gtctggagcttttaccgagggttggccttgcacggatctggaaatactgcataactc
tttcattttggctgggttatgacatggtcacagatcactgatggccatttttttgc
tactcaacttcgtatcgagtctcgcatgggttcaacaaacaaacaaatatggatgtt
20 cattagggacatgatcaaaggaacattcctctgtcataacttaggcccacccattgttgc
ctgcgataatttcatagtccagaaaggaggccttatcttgcacatctatctgtgggca
ttcatgttatcctgtcttagtgtatgactatataccggcttgcatacgaccgct
cttcaacaaattcactccttccagatggagacctcgggagaagattgagaaacttg
cttccctaaagttcatttgcataaggctgtttgtcgatggatctacaaggatca
25 agccatagcaatgcttacatgtatggttcttaagaacaaaaggattgttcttgc
tacgttgattcagcagtgcagaatgaggatgaaattgtggcggttattgcacacgagc
ttggacattggaaactgaatcacactacatactcgatggccatttgcagttcaacatc
tttacaattggaggatacactcttcagaaactccactgtatcttgcaggatgttgc
cgatggatcacacacgcctgttgcattgggttgcataatttcagcacactgt
30 cactgcaacatctagtaagcttggcctgaacctcgtagtcgagcggttgcagtt
gctgatgctttgtgaagcttgcactatgcacaaaagatcttcgtcctgtcttagt
actacaggaagagaacttatcaacaatgaacactgatccattgtactcagcttact
actcacatcctccttgcattggaaaggctcgagccactgtatggagaagacaaga
gatataaccctcgatcgtaacattggcaataaagttcttaagat

tgaatcctgttgcggcttgcgtattatcatataatttctgttgaattacgttaag
catgttaataattaacatgtaatgcgttatggatgggttttatgatttag
agtcccgaattatacattaaatacgcgttagaaaacaaaatatagcgcgcaactagg
ataaaattatcgcgccgggtgtcatctatgttactagatcgggaattcactggccgtcg
5 tttacaacgtcgtgactggaaaaaccctggcgtaacccaaacttaatcgcccttcgcagcac
atcccccttcgcccagctggcgtaatagcgaagaggccgcaccgatcgcccttccaa
cagttgcgcagcctgaatggcgccgctccttcgcttctcccttcgttccgc
cgttcgccggcttccccgtcaagctctaattcggggctccctttagggttccgattt
agtgcattacggcacctcgacccaaaaacttgattgggtatggttacgttagtgg
10 gccatcgccctgatagacggtttcgcccattgacgttggagtccacgttcttaata
gtggactttgttccaaactggaaacaacactcaaccstatctcggttattctttgat
ttataagggatttgcgatttcggaaccaccatcaaacaggatttcgcctgctggg
caaaccagcgtggaccgcttgcactctcaactctcagggccaggcggtgaagggcaatca
gctgttgcggcttcactggtaaaaagaaaaaccaccccaactacattaaaaacgtccgc
15 aatgtgttattaaatgttcaagcgtcaattgttacaccacaatatacctgcca

SEQ ID NO:4 is the nucleic acid sequence of pBI121-AtCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the 35S promoter and bolded sequence is the AtCPP sense sequence.

tcgattccaccgcgccttatgaaagggtggctcggaatcgccccggacgcc
 ggctggatgatcctccagcgcgggatctcatgctggagttctcgcccacggatctc
 tgccgaacaggcggtcgaagggtgccgatatacgcacagcaacggccgacaagcaca
 acgcccacgatcctgagcacaatatgatcggcccggtccacatcaacggcgtcggc
 5 ggcgactgcccaggcaagaccgagatgcaccgcgatatacttgctgcgttggatattt
 cgtggagttcccgccacagacccggatgatccccgatcgtaaacaacattggcaataaa
 gtttcttaagattgaatcctgttgcggcttgcgtatgattatcatataattctgtt
 aattacgttaagcatgtaataaattacatgtaatgcacgttattttagagatgggt
 tttatgattagatgtccgcattatacattataacgcataaaaaaaaatatacgc
 10 gcgcaaactaggataaattatcgccgcgggtcatctatgttactagatcgccctcc
 tgtcaatgctggcggcggctctgggtgggtctgggtggcgctctgagggtggct
 ctgagggtggcggtctgaggggtggcggtctgagggaggcggttcgggtggct
 ggtccgggtatggattatgaaaagatggcaaacgctaataaggggctatgaccga
 aaatgccatgaaaacgcgtacagtctgacgctaaaggcaaacttgattctgcgcta
 15 ctgattacgggtctgctatcgatggttcattggtgacgttccggcctgctaatttgt
 aatggtgctactggtagttgtggctctaattccaaatggctcaagtgggtgacgg
 tgataattcaccttaatgaataattccgtcaatattacccctccctcaatcg
 ttgaatgtccccctttgtcttggccaatacgc当地accgcctctcccgccgttgg
 ccgattcatatgcagctggcacaggttcccactggaaagcgggcaactggcg
 20 caacgc当地ataatgtgagtttagctactcattaggcacccaggcttacacttatgc
 ttccggctcgtatgtgtggaaattgtgagggataacaatttcacacaggaaacagc
 tatgaccatgattacgccaagcttgcattgcctgcagcccacagatggtagagaggctt
 acgc当地gggtctcatcaagacgttacccgagaataatctccaggaaatcaaatac
 cttccaaagaaggtaaagatgcagttaaagattcaggactaactgc当地aaacac
 25 agagaaagatatttc当地aaatcagatcagacttccactatggacgattcaaggct
 tgcttc当地aaaccaggcaactatggacttctctaaaaggtagttccact
 gaatcaaaggccatggagtcaaagattcaatagaggacctaacaactcgccgtaaa
 gactggc当地acagttc当地atcagacttcttgcactcaatgacaagaagaaaatctcg
 tcaatggtgagcacacacttttactccaaaatataaggatacagttca
 30 gaagaccaaaaggcaatttgagactttcaacaaaggtaatatccggaaacctcctcg
 attccattgcccactatctgtcactttattgtgaagatagtggaaaaggtaggtggct
 cctacaaatgccc当地tattgc当地ataaggccatcgtaagatgcctctggccac
 agtggtcccaaagatggaccccccaccacgaggagcatcgtaaaaaagaagacgttcc
 aaccacgttcaagcaagtggattgtgatatccactgc当地aggatgacg
 35 cacaatcccactatcctcgcaagacccttcttatataaggaaatgttcaatttgc
 gagagaacacggggactctaggatcctccaaatgtccaaatcgctgtgc当地aaacc
 gccacaatttcatcctcatttgcactgc当地atcaacgtatcataaagaacaatcct
 tttgttcttaagaaaccatatacatgttaagcattgc当地atggcttgc当地atccat
 cgacaacaacagtttcttcaaaggaaacttttagggagaaggaaatgttcaatcttc
 40 tcccgaggtctccatctggaaagaggagtgaatttgc当地atggacttgc当地atcaagac
 cgggtatatagtcatcatcactagagacaggataaaatgc当地atggccacagatagatgg
 caagataaggacctccttctggactatgaaaattatgc当地aggaaatgggtggccct
 agtgc当地aggaggaaatgttgc当地atggacttgc当地atggacttgc当地atgg
 tttgttgc当地accatgc当地aggactcgatc当地aggaaatgttgc当地atgg
 45 cagtgc当地atgtgaccatgtc当地acaccaggccaaaggaaatgtc当地atgttca
 ttctccggatcaaggccaaacctcgtaaaaggatccccATCTACCCGCTTCGCGTC
 GGCATCCGGTCAGTGGCAGTGAAGGGCGAACAGTTCTGATTAACCACAAACCGTTCTA
 CTTACTGGCTTGGCTCATGAAGATGCGGACTTGCCTGGCAAAGGATTGATAACG
 TGCTGATGGTGACGACCACGCATTAATGGACTGGATTGGGGCAACTCCTACCGTACC
 50 TCGCATTACCCTTACGCTGAAGAGATGCTCGACTGGCAGATGAACATGGCATCGTGGT
 GATTGATGAAACTGCTGCTGCGCTTTCGCTCTTAGGCATTGGTTCGAAGCGG

GCAACAAGCCGAAAGAACTGTACAGCGAAGAGGCAGTCACGGGAAACTCAGCAAGCG
 CACTTACAGGCAGTAAAGAGCTGATAGCGCGTACAAAAACCACCCAAGCGTGGTGA
 GTGGAGTATTGCCAACGAACCGGATAACCGTCCGCAAGGTGCACGGGAATATTCGCG
 CACTGGCGGAAGCAACCGTAAACTCGACCCGACCGTCCGATCACCTCGTCAATGTA
 5 ATGTTCTGCGACGCTCACACCGATAACCATCAGCGATCTTTGATGTGCTGTGCCTGAA
 CGTTATTACGGATGGTATGCCAACAGCGGATTGAAACGGCAGAGAAGGTACTGG
 AAAAAGAACTCTGGCCTGGCAGGAGAAACTGTACACCGACATGTGGAGTGAAGAGTAT
 CAGTGTGCATGGCTGGATATGTATCACCGCTTGTGATCGCGTCAGGCCGTCGG
 TGAACAGGTATGGAATTGCCGATTGCGACCTCGCAAGGCATATTGCGCGTGGCG
 10 GTAACAAGAAAGGGATCTTCACTCGCGACCGCAAACCGAAGTCGGCGGCTTCTGCTG
 CAAAAACGCTGGACTGGCATGAACCTCGGTGAAAACCGCAGCAGGGAGGCAAACAATG
 AATCAACAACTCTCTGGCGCACATCGTGGCTACAGCCTGGGAATTGCTACCGAGC
 TCttttaccgaggttgggccttgatccggagaatgaaatactgcataactcttcattct
tggctggtgttatgacatggcacagatcactgattgccatttttttactcaact
 15 ttcgatcgagtcgcgatgggttcaacaaacaaatatggatgttcattaggga
catgatcaaaggaacattccctctgtcatactaggcccaccattgttgctgcataaa
tttcatagtccagaaaggaggcccttatcttgccatctatctgtggcattcatgttt
atcctgtctctagtatgactataccggcttgcatacgaccgcttcaacaa
attcaactccctttccagatggagaccccccggagaagattgagaaacttgcttccc
 20 taaagttcccttgaagaagctgtttgtcgatggatctacaaggtaagccatagc
aatgcttacatgtatggtttttaagaacaaaaggattgttctttagatacgttgat
tcagcagtgcagaatgaggatgaaattgtggcggttattgcacacqaqcttgacatt
ggagctcgaaattccccgatcgtaaacacattggcaataaagtttcttaagattgaa
tccgttgcggcgtttgcgtatgattatcatataattctgttgaattacgttaagcatg
 25 taataattaacatgtaatgcgtacgttattatgagatgggttttatgattagatc
ccgcaattatacattaatacgcgatagaaaacaaaatagcgcgcacactagatgaa
attatcgccgcgggtcatctatgttactagatcggaattcactggcgtcggttta
caacgtcgactggaaaaccctggcgttacccaacttaatcgcttgcagcacatcc
cccttcgcccagctggcgtaatagcgaagagggccgcaccatcgcccttccaaacagt
 30 tgcgcagctgaatggcgccgctcccttcgttctcccttcgttgcacgcgtt
cggcgttcccgctcaagcttaatcggggtccctttaggttccgatttagtg
cttacggcacctcgacccaaaaacttgattgggtatggttacgttagtggcca
tcgcccctgatagacggttttcgcccttgcgttgcacgttgaggatccacgtttaatagtgg
 35 actttgttccaaactggaacaacactcaaccstatctcggttattttttagttat
aagggatttgcgatttcggaaccaccatcaaacaggatttcgctgtggggaaa
ccagcgtggaccgcttgcactctcagggccaggcgggtgaagggaatcagctg
ttgcccgtctcactggtaaaaagaaaaaccaccccagtagacattaaaacgtccgcaatg
tgttattaaagttgtctaagcgtcaattgtttacaccacaatatacctgcca

40 SEQ ID NO:5 is the nucleic acid sequence of pBI121-HP-AtCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the 35S promoter and bolded sequence is the AtCPP anti-sense sequence. Sequence in upper case is the truncated GUS fragment. Sequence in bold and underlined is the AtCPP sense sequence.

SEQ ID NO:35

45 gtttacccgccaatatacctgtcaaacactgtatgtttaaactgaaggcggaaacga
caatctgatcatgagcggagaattaaggagtcacgttatgaccccccgcgtacgcg
ggacaagccgtttacgtttggactgacagaaccgcaacgttgaaggagccactcagc

cgcgggtttctggagttaatgagctaaggcacatacgtcagaaaccattattgcgcgtt
caaaagtgcgctaaggtcaactatcagtagcaaatatcttgcgtaaaaatgctccact
gacgttccataaattcccctcggtatccaatttagagtctcatattcactctcaatccaa
ataatctgcaccggatctggatcggtcgcatgattgaacaagatggattgcacgcagg
5 ttctccggccgcttgggtggagaggctattcggtatgactggcacaacagacaatcg
gctgctctgatgccgcgtgttccggctgtcagcgcagggcgcccggtttttgc
aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcg
gctggccacgacggcggtcctgcgcagctgtgcacgttgtcactgaagcggaa
gggactggctgtattggcgaagtgccggggcaggatctcctgtcatctcacctgct
10 cctgcccggaaaatccatcatggctgatgcaatgcggcggctgcatacgcttgc
ggctacctgcccattcgaccaccaagcggaaacatcgcatcgagcgcacgtactcg
tggaaagccggcttgcgatcaggatgttgcacgaagagcatcagggctcgccca
gccgaactgttcgcaggctcaaggcgcgcattgcgcacggcgatgtctcgctgac
ccatggcgatgcctgcattgcgaatatcatggtgaaaatggccgtttctggattca
15 tcgactgtggccggctgggtgtggcggaccgctatcaggacatagcgttgctacc
gatattgcgatcggatgttgcgatcggatgttgcgcgttgcgtgtttacggat
cgccgctcccgattcgcagcgcattgcgcattgcgcattgcgcattgcgcatt
cgggactctgggttcgaaatgaccgaccaagcgcacgcacccacctgcac
tcgattccaccgcgccttatgaaagggtggcttcggaatcggtttccggacgcc
20 ggctggatgatcctccagcgcgggatctcatgctggagttttcgccccacggatctc
tgcggAACAGGCGGTcgaagggtgcgcattacgcacagcaacggccgacaagcaca
acgcacgcattgcgacaaatatgatcggccggcgtccacatcaacggcgtcggc
ggcgcattgcgcaggcaagaccgagatgcaccgcgcattgcgttgcgttgcgc
cgtggagttccgcacagacccggatgcattttgcattttgcgttcaacat
25 gtttcttaagattgaatcctgttgcggcttgcgtatgattatcatataattctgttgc
aattacgttaagcatgtataattacatgtatgcgtacgttattatgagatgggt
ttttatgatttagagtccgcattatcatatgcgtatgcgtatgcgtatgcgt
gcgcacactaggataattatgcgcgcggcgtcatctatgttactagatcgggc
tgtcaatgcgtggcggcggctctgggtggttctggcggctctgggtggct
30 ctgagggtggcggttctgagggtggcggctctgaggaggcgggtccgggtggct
gggtccgggtgattttgattatgaaaagatggcaacgcataataaggggctatgacc
aaatgcccgtgaaaacgcgcctacagtctgcgtacgcataaggcaacttgc
ctgattacggtgctgcattgcgttgcgttgcgttgcgttgcgttgcgt
aatggtgctactggtgattttgcgtggctctaattccaaatggctcaagtc
ggcgtacgg

tgataattcaccttaatgaataattccgtcaatattacctccctccctcaatcg
 ttgaatgtgcgcctttgtcttgcccaatacgcaaaccgcctctccccgcgcgtgg
 ccgattcattaatgcagctggcacgcacaggttcccactggaaagcgggcagtgagcg
 caacgcattaatgtgagtagtcactcattaggcaccccaaggcttacactttatgc
 5 ttccggctcgatgtgtgaaattgtgagcggataacaattcacacaggaaacagc
 tatgaccatgattacgccaagcttgcattgcgcgcagcccacagatggtagagaggctt
acgcagcaggctcatcaagacgatctaccgagcaataatctccaggaaatcaaatac
cttccaagaaggtaaagatgcagtcaaaagattcaggactaactgcataagaacac
agagaaagatataattctcaagatcagaagtactattccagtagtgacgattcaaggct
 10 tgcttcacaaaccaaggcaagtaatagagattggagtctctaaaaggtagttcccact
gaatcaaaggccatggagtcaaagatcaaataagaggacctaacagaactcgccgtaaa
gactggcgaacagttcatacagagtcttacgactcaatgacaagaagaaaatctcg
tcaacatggtgagcacgacacacttgtctactccaaaaatataagatacagtctca
gaagaccaaaggcaattgagactttcaacaaaggtaatatccggaaacctcctcgg
 15 attccattgcccagctatctgtcactttattgtgaagatagtggaaaaggaagggtggct
cctacaaatgccatcattgcgataaaggaaaggccatcggtgaagatgcctctgcccac
agtggtcccaaagatggacccccaccacgaggagcatcggtggaaaaagaagacgttcc
aaccacgtctcaaagcaagtggattgtgatatctccactgacgtaagggatgacg
cacaatcccactatccttcgaagacccttccttatataaggaagttcatttcatttg
 20 gagagaacacggggactctagaggatccTTAATCTGCTTCTGTCTTCCATCAGT
GGCTCGAACGCCTTCACAAAGAGGGAGTGTGAGTAGTGATAAGCTGAGTACAATGGAT
CAGTGTTCATTGTTGATAAGTTCTCTCCTGTAGTTCACTAGAGCAGGACGAAGATCT
TTTGCATAGTCAGCTCACAGCAAAGCATCAGCCTGAAACTCAAACGCTCGACTAAC
GAGGTTCAGGCCAAAGCTTACTAGATGTTGCAGTGGTATTACAGTGTGCTGAAATATGA
 25 TCAAACCAATGAGAACAGGCTGTATCAAATCCGAAACTCCTGAAAGAGATCAGTGGAG
TTTCTGAGAAGAGTGTATCCTCCAAATTGTAAGAAGGCAAGGAGTTGAAGTCAATGAA
CGAGTATGTAGTGTGATTCAAGCTGCTGAATCAACGTATCATAAAGAACAAATCCTTTGTC
TTAAAGAAACCATACTGTAAGCATTGCTATGGCTTGACCTTAGATCCATCGACAAC
 30 AAACAGCTTCTCAAAGGAAACTTAGGGAAGAAGCAAGTTCTCAATCTCCCAGGAA
GGTCTCCATCTGGAAGAGGAGTGAATTGTTGAAGAGCGGTGCTATCAAGACCGGGTAT
ATAGTCATCATCACTAGAGACAGGATAAACATGAATGCCACAGATAGATGGCAAGATA
AGGACCTCCTTCTGGACTATGAAAATTATCGCAGCAACAATGGTGGCCTAGTATGA
CAGAGAGGAATGTTCTTGATCATGTCCTAATGAACATCCATATTGTTGTTGTC

AACCCATGCCGAGACTCGATCACGAAAGTTGAGTACAAAGAAAATGGCAAATCAGTGAT
 CTGTGACCATGTCATAAACACCAGCCAAGAATGAAAGAGTATGCAGTATTCTATTCTCCG
 GATCAAGGCCAACCTCGGTAAAACAGCTCCAGACATCTCCAAAACCAAGGCAAGATC
 CCAAAGAACAAAATTGCAGAGTCCATAAGTATAAGTACAAACTCATGAACAAAGTGAAA
 5 ATAGTTTGTCAAGACTGTATGCTCGTATTCTCAAACCTCTTGGCTAATTACAC
 CAACCAAGGTTTCGGGAGAGTTGGAAGCTTGAGAGCAGTGAGTTGCCTCAGATCCAAA
 TACGTCTAAAAATGTACATCACTATCATAAAACCCACGACGGTTCCATGAAAGGAAT
 CGCCATcccctcgatccccgatcgtaacattggcaataagttcttaagat
 tgaatcctgttgcggcttgcgtattcatataattctgttgaattacgtttaag
 10 catgtataattaaatgtatgcattgtacgttatttatgagatgggtttatgattag
 agtcccgcaattatacatttataacgcgttagaaaaacaaaatagcgcgcaaactagg
 ataaaattatcgcgccgggtgtcatctatgttactagatcggaattcactggccgtcgt
 ttacaacgtcgtactggaaaaccctggcgtaatcgaaactggccgcaccgatcgcccttccaa
 15 cagttgcgcagcctgaatggcgcccgtccttcgtttctcccttcgttccca
 cgttcgccggcttcccgtaagctctaaatcggggctccctttagggttcgatt
 agtgcttacggcacctcgacccaaaaacttgattgggtgatggttacgttagtgg
 gccatcgccctgatagacggtttcgccccttgcgttgcgttgcacgtttaata
 gtggactcttgcgttccaaacttggaaacaacactcaaccatatctggctatttttgc
 20 ttataagggattttgcgatttcggaaccaccatcaaacaggatttcgcctgctggg
 caaaccagcgtggaccgcgttgctgcaactctctcaggccaggcggtgaagggaatca
 gctgttgcggctcactggtaaaagaaaaaccacccagttacattaaaacgtccgc
 aatgttttataagttgtctaagcgtaattttacaccacaatatacctgcca

SEQ ID NO:35 is the nucleic acid sequence of pBI121-antisense-AtCPP.

25 **Italicized sequences are the right and left border repeats. Underlined sequence is the 35S promoter. Sequence in upper case is the AtCPP anti-sense sequence.**

SEQ ID NO:36

gttacccccaatatacctgtcaaacactgtatgtttaactgaaggcggaaacga
caatctgatcatgagcggagaattaaggagtcacgttatgaccccccgcgtgacgcg
 30 *ggacaagccgtttacgttggactgacagaaccgcacgttgaaggagccactcagc*
cgcgggtttctggagttaatgagctaagcacatacgtcagaaaccattattgcgcgtt
caaaagtgccttaaggtcactatcagctagcaaataatttctgtcaaaaatgctccact
gacgttccataaattccctcggtatccaatttagagtctcatattcactctcaatccaa
ataatctgcaccggatctggatcgatcgatgttgcacgcagg

ttctccggccgcttgggtggagaggctattcggtatgactggcacaacagacaatcg
 gctgctctgatgccgcgtttccggctgtcagcgcagggggccccgggtctttgtc
 aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgccatcg
 gctggccacgacggcggttcctgcgcagctgtgctcgcacgttgtcaactgaagggaa
 5 gggactggctgtattggcgaagtgccggggcaggatctcctgtcatctcaccttgct
 cctgccgagaaagtatccatcatggctgtcaatgcggcggctgcatacgcttgatcc
 ggctacctgcccattcgaccaccaagcgaaacatcgcatcgagcgacgtactcgga
 tggaaagccggcttgcgtcaggatctggacgaagagcatcagggctcgccca
 gccgaactgttcgcaggctcaaggcgcatgcggcgcacggcatgtctcgctgac
 10 ccatggcgatgcctgcttgcgaatatcatggaaaatggccgtttctggattca
 tcgactgtggccggctgggtgtggcggaccgctatcaggacatagcgttgcgtaccgt
 gatattgctgaagagcttggcggcgaatggctgaccgcattcgtgcgttgcgttat
 cgccgctccgattcgcatgcgcattctatgcgcatttgcgtacgagttcttgcgt
 cggactctggggttcgaaatgaccgaccaagcgacgccccacctgcacgagatt
 15 tcgattccaccgcgccttatgaaagggttggcttcggaatcggtttccggacgcc
 ggctggatgatcctccagcgccggatctcatgctggagttctcgccacggatctc
 tgcggAACAGGCGGTcgaagggtgccatattacgacagcaacggccgacaagcaca
 acgcccacgatcctgagcgacaatatgatcggccggcgtccacatcaacggcgctgg
 ggcactgcccaggcaagaccgagatgcaccgcgatatttgctgcgttgcgtatattt
 20 cgtggagttccgcacagacccggatgatccccgatcgtaaacatattggcaataaa
 gtttcttaagattgaatcctgttgcggcttgcgtatgattatcatataattctgttg
 aattacgttaagcatgtataattacatgtaatgcgtacgttattatgagatgggt
 tttatgatttagagtccgcattatcatattacgcgtatggaaacaaaatatacg
 gccaactaggataattatgcgcgcgggtgtcatctatgttactagatcggccctcc
 25 tgtcaatgctggcgccggctctgggtggttctggcggctctggggaggcgggtccgggtggct
 ctgagggtggcggtctgagggtggcggctctgaggaggcgggtccgggtggct
 ggttccgggtgatttgattatgaaaagatggcaaacgctaataaggggctatgaccga
 aaatgccatgaaaacgcgtacagtctgacgctaaaggcaaacttgattctgtcgcta
 ctgattacggtgctgtatcgatggttcattggtgacgttccggccttgcataatgg
 30 aatggtgctactggtgatttgctggctctaattccaaatggctcaagtgggtgacgg
 tgataattcaccttaatgaataattccgtcaatattacctccctccctcaatcg
 ttgaatgtcgccctttgtcttggccaaatcgcaaaccgcctctcccgccgttgg
 ccgattcattaatgcagctggcacgacaggttcccactggaaagcgggagtgacg
 caacgcaattaatgtgagtttagtcactcattaggcacccaggcttacactttatgc

ttccggctcgatgttgtgaaattgtgagcggataacaattcacacaggaaacagc
tatgaccatgattacgccaagcttgcattgcctgcagggagccatagatgcattcaatc
aaactgaaatttctgcaagaatctcaaacacggagatctcaagttgaaagaaaattt
atttcttcgactcaaaacaaacttacgaaatttaggtagaacttatacattatattg
5 taatttttgtaacaaaatgttttattattatagaattttactggtaaattaa
aatgaatagaaaaggtaattaaaggagagaggaggtaaacatttcttcatttt
catatttcaggataaattattgtaaaagttacaagattccatttgacttagtgtaaa
tgaggaatattctctagtaagatcatttcatctacttctttatcttctaccagta
gaggaataaacaatatttagtccttgtaaatacaaattaatttccttcttgacatc
10 attcaatttaatttacgtataaaaataaaagatcatacctattagaacgattaaggag
aaatacaattcgaatgagaaggatgtgccgttgtataataacagccacacgacgta
aacgtaaaatgaccacatgatggccaatagacatggaccgactactaataatagaag
ttacattttaggatgaaataatcataccgacatcagtttggaaagaaaaggaaaa
aaagaaaaaaaataaaaagatatactaccgacatgagttccaaaaagcaaaaaaaaaag
15 atcaagccgacacagacacgcgtagagagcaaaatgacttgcgtcacaccacgaaaa
cagacgcttcatacgtgtcccttatctctcagtctctataaacttagtgagacc
ctcctctgtttactcacaatatgcaaactagaaaacaatcatcaggaataaagggtt
tgattactctattggaaaggactctagaggatccatggcgattccttcatggaaacc
gtcgtgggtttatgatagtgtacattttgagacgtattggatctgaggcaact
20 cactgctctcaagcttccaactctccgaaaaccttgggtgttaattgccaagaga
agtttggaaaatcacgagcatacagtcgtacaaaagctatttcacttgcattgag
tttgcattacttatggactctgcatttttttttttttttttttttttttttttt
gaagatgtctggagctgtttaccggaggttggccttgcggagaatgaaataactgc
atactcttcatttttgcatttgcatttgcatttgcatttgcatttgcatttgcattt
25 tctttgtactcaacttcgtatcgagtctcgcatgggtcaacaaacaaacaatatg
gatgttcatttagggacatgatcaaaggaacatccctctgtcataactaggcccaccca
ttgttgcgtgcataatttcatagtccagaaaaggaggtccttgcatttgcatttgc
tgggcattcatgtttaccgtcttagtgcatttgcatttgcatttgcatttgcattt
acggctttcaacaaattcactccattccagatggagacccggagaagattgaga
30 aacttgcttccctaaagttccatttgcatttgcatttgcatttgcatttgcatttgc
aggtaagccatagcaatgcttacatgtatggtttttgcatttgcatttgcatttgc
ttatgatacgttgcattcagcagtgcatttgcatttgcatttgcatttgcatttgc
acgagctggacattggaaactgaatcacactacactcgatttgcatttgcatttgc
cttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgcatttgc

gagtttcggatttatacacacacgcgttctcattggttgatcatattcagcacactg
taataccactgcaacatcttagtaagcttggcctgaacctcgtagtcgagcgtttag
ttcaggctgatgctttgctgtgaagcttgactatgcaaaagatcttcgtcctgctct
agtgaaaactacaggaagagaacttatcaacaatgaacactgatccattgtactcagctt
5 **atcactactcacatcctcctttgttgaagaggcttcgagccactgatggagaagacaag**
aagacagat~~taa~~cccccgcgaattccccgatcgtaaacacattggcaataaagttct
taagattgaatcctgttgcggcttgcgtatgattatcatataattctgttgaattac
gttaagcatgtataattacatgtaatgcgttattatgagatgggttttat
gattagagtcccgcaattatacatttataacatcgcatagaaaaacaaaatagcgcgcaa
10 actaggataaattatcgcgccgggtgtcatctatgttactagatcgggaattcactggc
cgtcgtttacaacgtcgactggaaaaccctggcgtaacccacttaatcgcttg
cagcacatcccccttcgccagctggcgtaatagcgaagaggccgcaccgatcgccct
tcccaacagttgcgcagcctgtaatggcgccgccttcgcctttcccttc
tcgccacgttcgcggcttccccgtcaagctctaaatggggctccctttagggttc
15 cgatttagtgcacggcacctcgacccaaaaacttgattgggtatggtcacg
tagtggccatcgccctgatagacggtttcgccttgacgttgagtcacgttct
ttaatagtggactttgttccaaactggaaacaacactcaaccstatctcggctattct
tttgcattataagggatttgcgatttcgaaaccaccatcaaacaggatttcgcctg
ctggggcaaaccagcgtggaccgctgtcaactctctcaggccaggcggtaaggg
20 caatcagctgttgcgcgtctcactggtaaaagaaaaaccacccaggatattc
gtccgcaatgtgttattaagggtcttaaggcgtcaatttgccttacaccacaatatacct
gcca

SEQ ID NO:36 is the nucleic acid sequence of RD29A-AtCPP. Italicized
25 sequences are the right and left border repeats. Underlined sequence is the RD29A
promoter. Sequence in bold is the AtCPP sense sequence.

SEQ ID NO:37

gtttacccgccaatatacctgtcaaacactgatagttaaactgaaggcggaaacga
30 *caatctgatcatgagcggagaattaaggagtcacgttatgaccccccgcgtacgcg*
ggacaagccgtttacgtttggaaactgacagaaccgcaacgttgaaggagccactcagc
cggggttctggagttaatgagctaaggcacatacgtcagaaaccattattgcgcgtt
caaaagtgcctaaggcactatcagctagcaaatttctgtcaaaaatgctccact
gacgttccataaattccctcggtatccaatttagtctcatattcactctcaatccaa

caacgcaattaatgtgagttagctactcattaggcacccaggcttacacttatgc
 ttccggctcgatgtgtggaattgtgagcggataacaattcacacaggaaacagc
 tatgaccatgattacgccaagcttgcattgcctgcaggagccatagatgcaattcaatc
 aaactgaaatttctgcaagaatctcaaacacggagatctcaaagttgaaagaaaattt
 5 atttctcgactcaaaacaacttacgaaatttaggtagaacttatatacattatattg
 taattttgttaacaaaatgttttattattatagaattttactggtaaattaaa
 aatgaatagaaaaggtgaattaagaggagagaggtaaacatttcttatttt
 catatttcaggataaatttgcatttttttttttttttttttttttttttttttttt
 tgaggaatattctctagtaagatcatttcatctacttctttatcttctaccagta
 10 gagaataaacaatatttagtccttgtaaatacaaaattaatttccttgcacatc
 attcaatttaatttacgtataaaataaaagatcatacctattagaacgattaaggag
 aaatacaattcgaatgagaaggatgtgccgttgtataataaacagccacacgacgta
 aacgtaaaatgaccacatgatggccaatagacatggaccgactactaataatagtaag
 ttacattttaggatgaaataatcataccgacatcagtttgcatttttttttttttt
 15 aaagaaaaaaaataaaaatataactaccgacatgagttccaaaaagcaaaaaaaaaaag
 atcaagccgacacagacacgcgttagagagcaaaatgactttgacgtcacaccacgaaaa
 cagacgcttcatacgtgtcccttatctctcagtctctataaaacttagtgagacc
 ctctctgtttactcacaatatgcaactagaaaacaatcatcaggaataaagggtt
 tgattacttctattgaaaggactctagaggatcctcccaatgtccaaagctcgatgtgca
 20 ataaccgccacaatttcatcctcatttgcactgctgaatcaacgtatcataaagaac
 aatcctttgttcttaagaaaccatacatgtaaagcattgctatggcttgcacctttag
 atccatcgacaacaaacagcttcttcaaaggaaactttaggaaagaagcaagttctca
 atcttctccggaggctccatctgaaagaggagtgaatttgcatttttttttttttttt
 caagaccgggtatatagtcatcatcactagagacaggataaacatgaatgcccacagat
 25 agatggcaagataaggacctccttctggactatgaaaattatcgccagcaacaatgggt
 gggcttagtatgacagagaggaatgttgcatttgcatttgcatttgcatttgcatttgcatt
 ttt
 gcaaatcagtgtatgtgaccatgtcataacaccagccaagaatgaaagagtatgcagt
 atttcatttccggatcaaggccaaacctcgtaaaagaggatccccATCTACCCGCTT
 30 CGCGTCGGCATCCGGTCAGTGGCAGTGAAGGGCGAACAGTCCTGATTAACCACAAACC
 GTTCTACTTTACTGGCTTGGTCGTCACTGAAGATGCAGACTTGCGTGGCAAAGGATTG
 ATAACGTGCTGATGGTCACGACCACGCATTAATGGACTGGATTGGGGCAACTCCTAC
 CGTACCTCGCATTACCCTACGCTGAAGAGATGCTCGACTGGCAGATGAACATGGCAT
 CGTGGTGATTGATGAAACTGCTGCTGGCTTTCGCTCTTTAGGCATTGGTTCG

AAGCGGGCAACAAGCCGAAAGAACGTACAGCGAAGAGGGAGTCAACGGGGAAACTCAG
CAAGCGCACTTACAGGCATTAAAGAGCTGATAGCGCGTGACAAAAACCACCCAAGCGT
GGTATGTGGAGTATTGCCAACGAACCGGATACCGTCCGCAAGGTGCACGGGAATATT
TCGCGCCACTGGCGGAAGCAACCGTAAACTCGACCCGACCGTCCGATCACCTGCGTC
5 AATGTAATGTTCTCGCAGCCTCACACCGATACCATCAGCGATCTTTGATGTGCTGTG
CCTGAACCGTTATTACGGATGGTATGTCAAAGCGCGATTGGAAACGGCAGAGAAGG
TACTGGAAAAGAACCTCTGGCTGGCAGGAGAAACTGTACACCGACATGTGGAGTGAA
GAGTATCAGTGTGCATGGCTGGATATGTATCACCGCGTCTTGATCGCGTCAGCGCCGT
CGTCGGTGAACAGGTATGGAATTTCGCCGATTTGCGACCTCGCAAGGCATATTGCGCG
10 TTGGCGGTAACAAGAAAGGGATCTTCACTCGCAGCGAACCGAAGTCGGCGGCTTT
CTGCTGCAAAACGCTGGACTGGCATGAACCTCGGTAAAAACCGCAGCAGGGAGGCAA
ACAATGAATCAACAACACTCCCTGGCGACCACATCGTGGCTACAGCCTGGGATTGCTA
CCGAGCTC ttttaccgaggttggc ttgatccggagaatgaaatactgcatactttt
cattcttgctggttatgacatggcacagatcaactgatttgccat
ttttgtac tcaactttcgatcgagtcggcatgggtcaacaaacaatatggatgttcat
tagggacatgatcaaaggAACATTCCCTCTGTcatacttaggcccacccattttgtcgt
cgataatttcatagtcagaaaggaggtccttatcttgcacatctatgtggcattc
atgtttatcctgtcttagtgcatactgactatataccggcttgatagcaccgcttt
caacaaattcactcctccagatggagacacctccggagaagattgagaaacttgctt
20 cttccctaaagtcccttgaagaagctgttgcgtatggatctacaaggtaacgc
catagcaatgcttacatgtatggtttttaagaacaaaaggattgttcttgcata
gttgcattcagcagtgcagaatgaggatgaaattgtggcggttattgcacacgagcttgc
gacattggag ctcgaattccccgatcgtaaaccattggcaataaagtttcttaag
attgaatcctgttgcggcttgcgtatgattatcatataattctgttgaattacgtta
25 agcatgtataattaacatgtaatgcgtacgttattatgagatgggtttatgatt
agagtcccgcaattatacatttatgcgtatggaaaacaaaatatacgccgcaacta
ggataaattatcgccgggtgtcatctatgttactatgcgtatggaaattcactggccgtc
gttttacaacgtcgtgactggaaaaccctggcgtaacccaacttaatcgccctgcagc
acatcccccttcgcccagctggcgtaatgcgtatggggccgcaccgatcgcccttccc
30 aacagttgcgcagcctgaaatggcgccgctccttcgtttcccttccttcgc
cacgttgcggcttccccgtcaagctctaattcggtttcccttgcgtatgggttccgat
ttagtgcttacggcacctcgacccaaaaacttgattgggtatggttcacgtat
ggccatcgccctgatagacggtttcccttgcgttgcgttggagttccacgttcttaa
tagtggactcttgttccaaactgaaacaactcaaccctatctcggttgcgttcttgc

atttataagggattttgccgatttcggaaccaccatcaaacaggattttcgccctgctgg
ggcaaaccagcgtggaccgcgttgctgcaactctctcagggccaggcggtgaaggcaat
cagctgttgcgcgtctcactggtaaaaagaaaaaccaccccagtacattaaaaacgtcc
gcaatgtgttattaagttgtctaagcgtcaatttgtttacaccacaatatacctgcca

5

SEQ ID NO:37 is the nucleic acid sequence of RD29A-HP-AtCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the RD29A promoter. Sequence in bold is the AtCPP anti-sense sequence. Upper case sequence represents the truncated GUS fragment. Bold and underlined sequence represents the *A. thaliana* CaaX prenyl protease sense fragment.

10 *thaliana* CaaX prenyl protease sense fragment.

SEQ ID NO:38

gtttaccgc₁caatatac₂c₃gtcaa₄acactgat₅ttaaactgaaggcggaaacga
caatctgatcatgagcggagaattaagggagt₆cac₇ttatgacccccg₈ccatgacgcg
15 ggacaaggc₉gtttac₁₀gtt₁₁g₁₂aactgacagaacc₁₃cg₁₄caac₁₅gttgaaggagccactc₁₆c₁₇agc₁₈
cg₁₉cg₂₀gg₂₁tttctgg₂₂agtt₂₃aatgag₂₄ctaagcacatac₂₅gtc₂₆agaaaccattattgcgc₂₇gtt
caaaagtcgc₂₈ctaagg₂₉t₃₀c₃₁actatc₃₂agct₃₃tagcaa₃₄atatttctgt₃₅caaaaatgctcc₃₆act
gac₃₇gttccataa₃₈attccc₃₉ctcg₄₀gtatcca₄₁att₄₂ag₄₃gtct₄₄catattc₄₅actct₄₆caatccaa
ataatctgc₄₇acc₄₈ggatctggat₄₉cg₅₀ttcg₅₁catgatt₅₂gaacaagatggattgcacgc₅₃agg
20 ttctccggccg₅₄cttgg₅₅gttgg₅₆agagg₅₇ctattcg₅₈gctatgact₅₉gggcacaac₆₀agacaatcg₆₁
gctgctctgat₆₂gccg₆₃ccgt₆₄gttccgg₆₅ctgt₆₆cagcgc₆₇agg₆₈ggc₆₉cccgg₇₀ttctttgtc
aagaccg₇₁ac₇₂ctgtcc₇₃gg₇₄gtcc₇₅ctgaat₇₆gaactgc₇₇agg₇₈ac₇₉gagg₈₀cagcgc₈₁gg₈₂ctatcg₈₃tg₈₄
gctggcc₈₅ac₈₆cg₈₇ac₈₈gg₈₉gc₉₀ttc₉₁cttgc₉₂gc₉₃ag₉₄ctgt₉₅gtc₉₆actgaagc₉₇gg₉₈aa
gg₉₉actgg₁₀₀ctg₁₀₁tattgg₁₀₂gc₁₀₃aa₁₀₄gtgc₁₀₅cc₁₀₆gg₁₀₇gc₁₀₈agg₁₀₉atctc₁₁₀c₁₁₁ac₁₁₂cttgc₁₁₃t₁₁₄
25 cctgcc₁₁₅gaga₁₁₆aa₁₁₇gttatcc₁₁₈catcatgg₁₁₉ctgat₁₂₀gcaatgc₁₂₁ggc₁₂₂gg₁₂₃ctgc₁₂₄at₁₂₅ac₁₂₆gtt₁₂₇gatcc₁₂₈
ggctac₁₂₉ctgccc₁₃₀attc₁₃₁gacc₁₃₂aca₁₃₃agc₁₃₄gaa₁₃₅ac₁₃₆atcg₁₃₇catcg₁₃₈gagc₁₃₉gac₁₄₀gtactcg₁₄₁ga
t₁₄₂ggaagcc₁₄₃gg₁₄₄t₁₄₅ttgt₁₄₆cgat₁₄₇c₁₄₈agg₁₄₉at₁₅₀ctgg₁₅₁ac₁₅₂ga₁₅₃ag₁₅₄gat₁₅₅c₁₅₆agg₁₅₇gg₁₅₈ctcg₁₅₉gc₁₆₀cca
gcc₁₆₁ga₁₆₂ct₁₆₃gttgc₁₆₄cc₁₆₅agg₁₆₆ctcaaggc₁₆₇gc₁₆₈gc₁₆₉at₁₇₀gccc₁₇₁gac₁₇₂gg₁₇₃gc₁₇₄at₁₇₅gtatctcg₁₇₆tc₁₇₇gt₁₇₈gac₁₇₉
ccatgg₁₈₀cgatgc₁₈₁c₁₈₂ctg₁₈₃cttgc₁₈₄gaat₁₈₅atcat₁₈₆ggt₁₈₇ggaaa₁₈₈atggcc₁₈₉g₁₉₀tttctgg₁₉₁attca
30 tcgactgtggccgg₁₉₂ctgg₁₉₃gt₁₉₄ggc₁₉₅gacc₁₉₆gc₁₉₇tatc₁₉₈agg₁₉₉aca₂₀₀tagc₂₀₁gttgg₂₀₂ctacc₂₀₃gt
gatatt₂₀₄gt₂₀₅ga₂₀₆ag₂₀₇ag₂₀₈cttgg₂₀₉cg₂₁₀ga₂₁₁atgg₂₁₂ctgacc₂₁₃gc₂₁₄ttc₂₁₅ctcg₂₁₆gt₂₁₇ttac₂₁₈gg₂₁₉ta₂₂₀
cgcc₂₂₁gc₂₂₂tc₂₂₃cc₂₂₄gattc₂₂₅gc₂₂₆agc₂₂₇gc₂₂₈at₂₂₉cg₂₃₀c₂₃₁tt₂₃₂tatc₂₃₃gc₂₃₄tt₂₃₅cttgc₂₃₆gac₂₃₇gag₂₃₈tt₂₃₉ct₂₄₀tg₂₄₁ag₂₄₂
cg₂₄₃gg₂₄₄act₂₄₅ct₂₄₆gggg₂₄₇ttcg₂₄₈aa₂₄₉atgacc₂₅₀gac₂₅₁cca₂₅₂ac₂₅₃c₂₅₄ctg₂₅₅ccat₂₅₆cac₂₅₇gag₂₅₈att₂₅₉
tcgattccacc₂₆₀ggcc₂₆₁cc₂₆₂tt₂₆₃ctat₂₆₄gaa₂₆₅agg₂₆₆ttgg₂₆₇ctcg₂₆₈gaat₂₆₉cg₂₇₀tttcc₂₇₁gg₂₇₂ac₂₇₃gc₂₇₄

ggctggatgatcctccagcgccccatctcatgctggagttctcgcccacggatctc
tgccgaacaggcggtcgaaagggtgccatatcattacgacagcaacggccacaagcaca
acgccacgatcctgagcgacaatatgatcggccggcgtccacatcaacggcgtcggc
ggcactgcccaggcaagaccgagatgcaccgcatacttgcgtcgatatttt
5 cgtggagttcccggccacagacccggatgtccccgatcgtaaacatggcaataaa
gttcttaagattgaatcctgttgcggcttgcgtgattatcatataattctgttg
aattacgttaagcatgtataataacatgtaatgcacgttatggatgggt
tttatgatttagagtcccgaattatacattaatacgcataaaaaaaaatagc
gccaactaggataaattatcgccgcgggtgtcatctatgttactagatcgccctcc
10 tgtcaatgctggccggctctgggtgggtctgggtggccgtctgagggtggct
ctgagggtggccggttctgagggtggccgtctgagggaggccgggtccgggtggct
gggtccgggtgatttgattatgaaaagatggcaaacgctaataaggggtatgaccga
aaatgccatgaaaacgcgctacagtctgacgctaaaggcaacttgattctgtcgcta
ctgattacggtgctgctatcgatggttcattggtacggttcggccctgctaattggt
15 aatggtgctactgggattttgtggctctaattccaaatggctcaagtccgtgacgg
tgataattcaccttaatgaataattccgtcaatattacccctccctcaatcggt
ttgaatgtcgccctttgtcttggcccaatacgcaaaccgcctctcccgccgttgg
ccgattcattaatgcagctggcacgacaggttccgactggaaagcggcagtgagcg
caacgcaattaatgtgagttagctcactcattaggcacccaggcttacactttatgc
20 ttccggctcgatgttgttggatttgtgagccgataacaattcacacaggaaacagc
tatgaccatgattacgccaagcttgcattgcctgcaggagccatagatgcaattcaatc
aaactgaaattctgcaagaatctcaaacacggagatctcaaagttgaaagaaaaatt
atttctcgactcaaaacaaactacgaaatttaggtagaacttatatacattatattg
taattttttaacaaaatgttttattattataagaattttactggttaaattaaa
25 aatgaatagaaaaggtaatagaggagagaggaggtaaacatttcttattttt
catattttcaggataaattttagttaaaagtttacaagattccatttgcattttt
tgaggaatttcttagtaagatcatttcatctacttctttatcttaccagta
gaggaataaacaatatttagtccttgcataacaaattaatttccatttgcattt
attcaatttaatttacgtataaaataaaagatcatacctattagaacgattaaggag
30 aaatacaattcgaatgagaaggatgtgccgttgttataataaaacagccacacgacgta
aacgtaaaatgaccacatgatggccaatagacatggaccgactactaataatagtaag
ttacatttttaggatggataataatcataccgacatcagtttgcatttttttttt
aaagaaaaataaataaaagatataactaccgacatgagttccaaaaagcaaaaaaaaaaag
atcaagccgacacacacacgcgttagagagcaaaatgactttgcgtcacaccacgaaaa

cagacgcttcatacgtgtcccttatctctctcagtctctctataaacttagtgagacc
ctcctctgtttactcacaaatatgcaaactagaaaacaatcatcaggaataaagggtt
tgattacttctattggaaaggactctagaggatccTTAATCTGTCTTGTCTTCTCC
ATCAGTGGCTCGAACGCCTTCAACAAAGAGGAGGATGTGAGTAGTGATAAGCTGAGTACA
5 ATGGATCAGTGGTCAATTGTTGATAAGTTCTCTCCTGTAGTTCACTAGAGCAGGACGA
AGATCTTTGCATAGTCAGCTCACAGCAAAAGCATCAGCCTGAAACTCAAACGCTCG
ACTAACGAGGTTCAGGCCAAAGCTTACTAGATGTTGCAGTGGTATTACAGTGTGCTGAA
ATATGATCAAACCAATGAGAACAGGCTGTATCAAATCCGAAACTCCTGAAGAGATCA
GTGGAGTTCTGAGAACAGAGTGTATCCTCAAATTGTAAGAAGGCAAGGAGTTGAACGTG
10 AATGAACGAGTATGTAGTGTGATTCACTAGTTCCAATGTCCAAGCTCGTGTGCAATAACCG
CCACAATTTCATCCTCATTCTGCAGCTGAATCAACGTATCATAAAGAACAAATCCTT
TTGTTCTAAAGAAACCACATGTAAGCATTGCTATGGCTTGACCTTGTAGATCCATC
GACAACAAACAGCTTCTCAAAGGAAACTTAGGGAAAGAAGCAAGTTCTCAATCTTCT
CCCAGGGTCTCCATCTGGAAAGAGGAGTGAATTGTTGAAGAGCGGTGCTATCAAGACC
15 GGGTATATAGTCATCATCACTAGAGACAGGATAAACATGAATGCCCACAGATAGATGGC
AAGATAAGGACCTCCTTCTGGACTATGAAAATTATCGCAGCAACAATGGGTGGCCTA
GTATGACAGAGAGGAATGTTCTTGATCATGTCCCTAATGAACATCCATATTGTTGT
TTGTTGAACCCATGCCGAGACTCGATCACGAAAGTTGAGTACAAAGAAAATGGCAAATC
AGTGATCTGTGACCATGTCATAACACCAGCCAAGAACATGAAAGAGTATGCAGTATTTCAT
20 TCTCCGGATCAAGGCCAACCTCGTAAACAGCTCCAGACATCTCCAAAACCAAGGC
AAGATCCAAAGAACAAAATTGCAGAGTCATAAGTATAGTTACAAACTCATGAACAAA
GTGAAAATAGCTTGTCAAGACTGTATGCTCGTGTGATTCTCAAACCTCTGGCTAA
TTACACCAACCAAGGTTTGGAGAGTTGGAAGCTTGAGAGCAGTGAGTTGCCTCAGA
TCCAAATACGTCTAAAAATGTACATCACTATCATAAAACCCACGACGGTTCCATGAA
25 AGGAATGCCATccccctcgaaattccccgatcgtaaacacattggcaataaagttct
taagattgaatcctgttgcggcttgcgtatgattatcatataattctgttgaattac
gttaagcatgtaataattaacatgtaatgcgtacgttattatgagatgggtttat
gattagagtcccgaattatacattaatacgcgtatggaaacaaaatatacgcgcaaa
actaggataaattatcgccgcgggtgtcatctatgttactagatcgaaattcactggc
30 cgtcgaaaaacaacgtcgactggaaaacctggcgatccacttaatcgcccttg
cagcacatcccccttcgcgcgtatggcgtaatagcgaagaggcccgcaccgatcgccct
tcccaacagttgcgcgcgtatggcgccgcgtatggcgatccacttaatcgcccttg
tcggccacgttgcgcggcttccccgtcaagctctaatacgggggctccctttagggttc
cgatttagtgcttacggcacctcgacccaaaaacttgattgggtgatggttcacg

tagtggccatgc~~ccc~~ttgatagacgg~~ttt~~gc~~cc~~tttgcgtggagtccacgttct
 ttaatagtggactctt~~gtt~~caaact~~gga~~acaacactcaacc~~t~~atctcggctattct
 tttgattataaggattt~~g~~ccgatttc~~g~~gaaccaccatcaa~~a~~caggatttgc~~c~~ctg
 ctggggcaaacc~~a~~cagg~~tg~~ggaccg~~c~~ttg~~c~~taactctc~~a~~ggccaggcg~~g~~gtgaagggg
 5 caatc~~a~~gctgttgc~~cc~~gtct~~c~~act~~gg~~t~~g~~aaaagaaaaaccacccc~~c~~agtacatta~~aa~~ac
 gtccg~~ca~~atgtgttattaagg~~t~~gtcta~~g~~c~~t~~caattt~~g~~tttacaccacaatata~~t~~c~~c~~
 gcca

SEQ ID NO:38 is the nucleic acid sequence of RD29A-antisense-AtCPP.

- 10 Italicized sequences are the right and left border repeats. Underlined sequence is the RD29A promoter. Sequence in upper case sequence is the AtCPP anti-sense sequence.

SEQ ID NO:39

*gttacccgccaatatacctgtcaa*acactgatagttaaactgaaggcggaaacga
 15 caatctgatcatgagcggagaattaaggagtcacgttatgacccccg~~cc~~gatgacg~~cg~~
 ggacaaggcgtttacgtt~~g~~gaactgacagaaccg~~ca~~acgttgaaggagccactc~~ag~~
 c~~cg~~gggttctggagtt~~a~~t~~g~~ag~~c~~ta~~a~~g~~c~~acatac~~g~~t~~c~~aga~~a~~accattattgc~~gc~~gtt
 caaaagtc~~gc~~ctaagg~~t~~cactatc~~ag~~ct~~a~~g~~c~~aaatattt~~c~~ttgt~~c~~aaaat~~g~~ctccact
 gacg~~tt~~ccataaattccc~~c~~tc~~g~~gtatcc~~a~~tttag~~g~~at~~t~~ct~~c~~atattc~~a~~c~~t~~tc~~a~~atccaa
 20 ataatctgcac~~cc~~gatctggatcg~~tt~~cgat~~g~~attgaacaagatggattgc~~ac~~gcagg
 ttctccggccg~~ct~~gggtggagagg~~c~~tattc~~gg~~ctatgact~~gg~~gcacaacagacaat~~cg~~
 gctg~~c~~t~~g~~tatgc~~cc~~cg~~gt~~ttcc~~gg~~ctgtc~~ag~~cg~~cc~~agg~~gg~~cg~~ag~~cg~~cc~~gtatc~~gt~~
 gctggccac~~g~~ac~~gg~~gc~~tt~~c~~tt~~gc~~g~~c~~ag~~ct~~gt~~g~~c~~t~~g~~ac~~gt~~tt~~gt~~act~~g~~a~~g~~cg~~gg~~aa
 25 gggactgg~~ct~~g~~c~~tattgg~~g~~cg~~a~~gtg~~cc~~ggggc~~agg~~at~~ct~~c~~ct~~gt~~c~~at~~ct~~c~~ac~~c~~tt~~g~~c~~t
 c~~ct~~g~~cc~~g~~a~~gaaa~~a~~gtatcc~~c~~at~~g~~g~~c~~t~~g~~at~~g~~ca~~a~~at~~g~~cc~~gg~~g~~g~~ct~~g~~cata~~g~~c~~tt~~g~~at~~cc
 gg~~c~~t~~ac~~c~~tc~~g~~cc~~att~~tc~~g~~ac~~cca~~g~~cg~~aa~~ac~~at~~cg~~cat~~cg~~ag~~cg~~ac~~gt~~ac~~t~~tc~~cg~~ga~~
 t~~g~~ga~~a~~g~~cc~~gg~~t~~tt~~gt~~cg~~at~~c~~agg~~at~~g~~tg~~g~~ac~~g~~ga~~a~~g~~g~~cat~~c~~agg~~gg~~ct~~cg~~gc~~cc~~
 g~~cc~~ga~~ac~~t~~gt~~tc~~g~~cc~~ag~~g~~c~~t~~ca~~agg~~gc~~cg~~cat~~g~~cc~~g~~ac~~gg~~cg~~at~~g~~at~~tc~~gt~~cg~~tg~~ac~~
 30 c~~ca~~atgg~~cg~~at~~gc~~c~~tc~~g~~ct~~g~~cc~~g~~a~~at~~at~~cat~~g~~gt~~gg~~aaa~~a~~at~~g~~g~~cc~~g~~ct~~tt~~ct~~gg~~att~~ca
 tc~~g~~act~~gt~~gg~~cc~~gg~~ct~~gg~~gt~~tg~~g~~cg~~gg~~acc~~g~~c~~t~~at~~c~~agg~~ac~~at~~g~~cg~~tt~~gg~~ct~~acc~~cg~~t
 gat~~at~~tg~~c~~t~~g~~aa~~g~~ag~~g~~c~~tt~~gg~~cc~~g~~g~~ca~~at~~gg~~g~~ct~~g~~acc~~g~~c~~tt~~c~~ct~~gt~~g~~ct~~tt~~ac~~g~~gt
 cg~~cc~~g~~ct~~cc~~cg~~att~~tc~~g~~cg~~ag~~cg~~cat~~g~~c~~ct~~t~~at~~cg~~cc~~tt~~ct~~gt~~g~~ac~~g~~ag~~tt~~ct~~t~~ct~~g~~ag
 cg~~gg~~act~~ct~~gg~~gg~~tt~~cg~~aa~~a~~at~~g~~acc~~g~~ac~~ca~~ag~~cg~~ac~~g~~g~~cc~~aa~~c~~c~~t~~g~~cc~~at~~c~~ac~~g~~ag~~at~~

tcgattccaccgcgccttatgaaagggtggcttcggaatcgccccggacgcc
 ggctggatgatcctccagcgcgggatctcatgctggagttctcgcccacggatctc
 tgcggAACAGGCGGTcgaagggtgccatATCATTACGACAGCAACGGCCACAAGCACA
 acGCCACGATCCTGAGCGACAATATGATCGGGCCCCGGCGTCCACATCAACGGCGTCGGC
 5 ggcgactgcccaggcaagaccgagatgcaccgcgatATCTTGCTCGTTGGATATTT
 CGTGGAGTTCCCACAGACAGCCGGATGATCCCCGATCGTTCAAACATTGGCAATAAA
 GTTCTTAAGATTGAATCCTGTTGCCGGTCTTGCATGATTATCATATAATTCTGTTG
 AATTACGTTAAGCATGTAATAATTACATGTAATGCATGACGTTATTATGAGATGGGT
 TTTATGATTAGAGTCCCACATTACATTAAACGCGATAGAAAACAAAATATAGC
 10 GCGCAAACCTAGGATAAATTATCGCGCGCGGTGTCATCTATGTTACTAGATCGGGCTCC
 TGTCAATGCTGGCGCGCTCTGGTGGTGGTCTGGTGGCGGCTCTGGAGGGTGGCT
 CTGAGGGTGGCGGTCTGAGGGTGGCGGCTCTGAGGGAGGCAGGTTCCGGTGGCTCT
 GGTCCGGTGGTGGCT
 15 CTGATTACGGTGTGCTATCGATGGTTATTGGTACGTTCCGGCCTGCTAATGGT
 AATGGTGCTACTGGTATTGCTGGCTTAATTCCAAATGGCTAAGTCGGTACCG
 TGATAATTACCTTAATGAATAATTCCGTCAATATTACCTCCCTCCCTCAATCGG
 TTGAATGTCGCCCTTGTCTTGGCCAATACGCAAACCGCCTCTCCCGCGCTGG
 CCATTGATTACGCTGGCACGACAGGTTCCGACTGGAAAGCGGGAGTGAGCG
 20 CAACGCAATTATGTGAGTTAGCTACTCATTAGGCACCCAGGCTTACACTTATGC
 TTCCGGCTCGTATGGTGTGGAATTGTGAGCGGATAACAATTACACAGGAAACAGC
 TATGACCATGATTACGCCAAGCTGGAAATTTCGCCAGTTCTAAATATCCGGAAACC
 TCTTGGGATGCCATTGCCATCTATCTGTAATTATTGACGAAATAGACGAAAAGGAAG
 GTGGCTCCTATAAAGCACATCATTGCGATAACAGAAAGGCCATTGTTGAAGATAACCTCT
 25 GCTGACATTGGTCCCCAAGTGGAAAGCACCACCCATGAGGAGCACCCTGGAGTAAGAAG
 ACGTTGAGGCCACGTCGAAAAAGCAAGTGTGTTGATGTAGTATCTCATTGACGTAAGG
 GATGACGCACAATCCAATCCATCGCAAGACCATTGCTCTATATAAGAAAGTTAATA
 TCATTCGAGTGGCACGCTGAGGGGGATCCatggcgattccttcatggaaacccgtcg
 tgggtttatgatagtgtacattttgagacgtattggatctgaggcaactcact
 30 gctctcaagcttccaactctccgaaaaccttggttgttaattagccaagagaagtt
 tgagaaatcacgagcatacagtcttgcacaaaagctatttcacttgcattgatgg
 taactatacttatggactctgcacattttgttggatcttgccttggtttggaaag
 atgtctggagctgtttaccgaggttggccttgcggagaatgaaatactgcatac
 tcttcattttggctggatgcacatggcacagatcactgatttgcatttctt

tgtaactcaactttcgatcgagtctcgcatgggttcaacaacaaacaatatggatg
 ttcattagggacatgatcaaaggaacattcctctgtcatactaggcccacccattgt
 tgctgcataatttcatagtccagaaaggaggtccttatcttgcacatctatctgtgg
 cattcatgtttatcctgtcttagtcatgtactatataccggcttgatagcacccg
 5 ctcttcaacaaattcactcctcttccagatggagaccccccggagaagattgagaaact
 tgcttctccctaaagttccttgaagaagctgtttgtcgatggatctacaaggt
 caagccatagcaatgtttacatgtatggttttaagaacaaaaggattgttcttata
 gatacgttgcattcagcagtgcagaatgaggatgaaattgtggcggttattgcacacga
 gcttggacattggaaactgaatcacactacactcgttcattgcagttcaaatccttg
 10 cttcttacaatttggaggatacactcttcagaaactccactgtatcttcaggagt
 ttccgatttgcatacacagcctgttctcattggttgtatcatatttcagcacactgtat
 accactgcaacatcttagtaagcttggcctgaacctcgtagtcgagcgttgcagtt
 aggctgatgccttgcgtgaagcttgactatgcaaaagatctcgtctgctctgt
 aaactacaggaagagaacttatcaacaatgaacactgtatccattgtactcagcttatca
 15 ctactcacatcctcctttgaaaggcttcgagccactgtatggagaagacaagaaga
 cagattaaccctcgaatttccccgatcgtaaacaatggcaataaagttcttaag
 attgaatcctgttgcggcttgcgtatgattatcatataattctgttgcattacgtt
 agcatgtataattacatgtatgcgtacgttattatgagatgggtttatgatt
 agagtcccgcaattatacattaatacgcgatagaaaacaaaatagcgcgcaacta
 20 ggataaattatcgcgcgccgtgtcatctatgttacttagatcggaattcactggccgtc
 gtttacaacgtcgactggaaaaccctggcgtaacccacttaatcgcccttgcagc
 acatcccccttcgccagctggcgtaatagcgaagaggccgcaccgatcgcccttccc
 aacagttgcgcagcctgaatggcgccgctccttcgcttctcccttccttcgc
 cacgttgcggcgttcccgtaagctctaattcggggctccctttaggggtccat
 25 ttatgtcttacggcacctcgacccaaaaacttgattgggtgtatggttcacgtat
 gggccatcgccctgatagacgggtttcgcccttgcgttgcgttgcgttgcacgtt
 tagtggactcttgcgttccaaactggaacaacactcaaccctatctcggttattctt
 attataaggatttgcgatttgcgaaccaccatcaaacaggatttcgctgtgg
 ggcaaacccagcgtggaccgcttgcgtcaactcttcagggccaggcggtgaaggcaat
 30 cagctgttgcggctcactggtaaaaagaaaaaccacccaggatcataaaaacgtcc
 gcaatgtgttattaaagttgtctaagcgtaattgttacaccacaatatacctgcca

SEQ ID NO:39 is the nucleic acid sequence of MuA-AtCPP. Italicized sequences are the right and left border repeats. Sequence in upper case is the MuA promoter. The *A. thaliana* CaaX prenyl protease sense sequence is in bold.

5 SEQ ID NO:40

gtttacccgccaatatacctgtcaaaca*cactgatagttaaactgaaggcggaaacga*
 caatctgatcatgagcggagaattaaggagtcacgttatgaccccccgcgatgacgcg
 ggacaagccgtttacgtttggaactgacagaaccgcaacgttgaaggagccactcagc
 cgccggtttctggagttaatgagctaagcacatacgtcagaaaccattattgcgcgtt
 10 caaaagtgcctaaggtcactatcagctagcaaataatttcttgcataaaaaatgctccact
 gacgttccataaattccctcggtatccaatttagagtctcatattcactctcaatccaa
 ataatctgcaccggatctggatcggtcgcatgattgaacaagatggattgcacgcagg
 ttctccggccgcttgggtggagaggctattcggtatgactgggcacaacagacaatcg
 gctgctctgatgccgcgtgttccggctgtcagcgcaggggcggccggttctttgtc
 15 aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcgtg
 gctggccacgcggcggtcccttgccgcagctgtgcgtcgcacgttgtcactgaagcggaa
 gggactggctgctattggcgaagtgcggggcaggatctcctgtcatctcaccttgct
 cctgccgagaaagtatccatcatggctgatgcaatgcggcggctgcatacgcttgc
 ggctacctgcccattcgaccaccaagcgaaacatcgcatcgagcgacgtactcgga
 20 tggaaagccggcttgcgatcaggatgatctggacgaagagcatcaggggctcgccca
 gccgaactgttcgcccaggctcaaggcgcatgcccacggcgatgtctcgatcgac
 ccatggcgatgcctgctgccaatatcatggtaatggggatggccgctttctggattca
 tcgactgtggccggctgggtgtggcgaccgctatcaggacatagcgttggctaccgt
 gatattgctgaagagcttggcggaatgggctgaccgcttcctcgatgtttacggat
 25 cggccgtcccgattcgcagcgcatgccttcatgccttcttgcgtcgcacgttcttgag
 cggactctgggttcgaaatgaccgaccaagcgacgccccacctgcacatcagagatt
 tcgattccaccgcccgccttcatgaaaggtggcttcggatcgatgtttccggacgccc
 ggctggatgatcctccagcgcgggatctcatgctggatcttgcggccacggatctc
 tgcggAACAGGCGGTcgaagggtgccgatattacgacagcaacggccgacaagcaca
 30 acgccacgatcctgagcgacaatatgatcggccggcgatccacatcaacggcgatcg
 ggcgactgcccaggcaagaccgagatgcaccgcgatcttgcgtcgatgttt
 cgtggagttccgcacagacccggatgtcccgatcgatgttcaaacatattggcaataaa
 gtttcttaagattgaatcctgttgcggcttgcgtatgattatcatataattctgttg
 aattacgttaagcatgtaataattaacatgtaatgcgtacgttatttatgagatgggt

tttatgattagagtccgcattatacattaaatcgatagaaaacaaaatata
gcgcaactaggataaattatcgccgcgggtcatctatgttactagatcgccctcc
tgtcaatgctggcggcggctctggtggtggctggcggctctgagggtggct
ctgagggtggcggttctgagggtggcggctctgagggaggcggccgggtggct
5 ggttccgggtgatttgattatgaaaagatggcaaacgctaataaggggctatgaccga
aaatgccatgaaaacgcgtacagtctgacgctaaaggcaaacttgattctgtcgcta
ctgattacggtgctgctatcgatggttcattggtgacgttccggccttgctaattgg
aatggtgctactggtgatttgctggctctaattccaaatggctcaagtcggtgacgg
tgataattcaccttaatgaataattccgtcaatattacccctccctcaatcg
10 ttgaatgtccccctttgtcttggccaaatcgcaaaccgcctctcccgccgttgg
ccgattcattaatgcagctggcacgacagggttcccactggaaagcggcagtgagcg
caacgcaattaatgtgagtagtcactcattaggcaccccaggcttacactttatgc
ttccggctcgatgtgtgtggatttgagcggataacaattcacacaggaaacagc
tatgaccatgattacgccaagctGGGAAATTTTCGCCAGTTCTAAATATCCGGAAACC
15 TCTTGGGATGCCATTGCCATCTATCTGAATTATTGACGAAATAGACGAAAAGGAAG
GTGGCTCCTATAAAGCACATCATTGCGATAACAGAAAGGCCATTGTTGAAGATAACCTCT
GCTGACATTGGTCCCCAAGTGGAAAGCACCACCCATGAGGAGCACCCTGGAGTAAGAAG
ACGTTGAGGCCACGTCGAAAAAGCAAGTGTGTTGATGTAGTATCTCCATTGACGTAAGG
GATGACGCACAATCCAATCCATCGCAAGACCATTGCTCTATATAAGAAAGTTAATA
20 TCATTCGAGTGGCACGCTGAGGGGATCGGGAATGGCGTTCCCTACATGGAAGCCGT
TGTCGGATTATGATATTAATGTACATTTGAAACTACTTGGATGTGCGACAACATA
GGGCCCTCAAACCTCCTACTCTTCCAAAGACTTAGAGGGTGTATCAGCCAAGAGAAA
TTTGAGAAATCTAGAGCCTATAGTCTTGTATAAAAGCCACTTCCATTGTTACGGAGTT
TGTGACAATAGTGACAGACTCTACAAATTGTACTTTGGGTATTGCCCTGGTTTGG
25 AGAAATCAGGAGATTTATGACAATAGCTGGTTCAATGCTGAGAATGAAATACTGCAT
ACCCTTGCCTTCTTAGCAGGGCTGATGATTGGTCACAGATAACAGATTGCCCTTTC
TCTGTACTCAACTTTGTGATTGAGGCCGTATGGTTAATAAGCAAACACCAGGT
TATTCTTAGGGACATGCTAAAGGAATTTCCTTCTGTAATAATTGGTCCACCTATT
GTGGCTGCAATCATTGTAATAGTACAGAAAGGAGGTCCACTTGGCCATCTATTTG
30 GGTTTTACGTTGGCTTCTATTGTGATGATGACCCATTCCAGTACTAATAGCTC
CACTCTCAATAAGTCACTCCACTTCCAGATGGTCAACTCAGGGAGAAAATCGAGAAA
CTTGCTTCCCTCAACTATCGTTAAAGAAACTATTGTTGTCGATGGATCCACAAG
ATCAAGTCACAGCAATGCCTATATGTATGGATTCTCAAGAACAAAGAGGATTGCCCTT
ATGACACATTAATTCAACAGTGCAGACGATGAGGAAATTGTTGCTGTTATTGCCCAT

GAGTTGGGACACTGGAAGCTCAACCATACTGTGTACACATTGTTGCTATGCAGATTCT
TACACTTCTACAATTGGAGGATATACACTAGTGCAGAAATTCACTGATCTGTATCGAA
GCTTTGGGTTTGATACGCAGCCAGTCCTCATTGGGCTCATCATATTTCAGCATACTGTA
ATCCCACTTCAGCAATTGGTCAGCTTGGTCTGAACCTAGTCAGCCGATCATTGAATT
5 TCAGGCTGATGGCTTGCCAAGAAGCTTGGATATGCATCTGGATTACGCCGGTGGCTTG
TGAAAACTACAGGAGGAGAATCTGTCACTATGAATAACAGATCCTGGTACTCTGCTTAT
CACTATTCTCATCCTCCCCCTGTTGAAAGATTGGCCGCGTGGACGAACCGGATAAGAA
GGAAGACTAAagagctcgaatttccccgatcgtaaaacattggcaataaagtttctta
agattgaatcctgttgcggcttgcgatgattatcatataattctgttgaattacgt
10 taagcatgtaataattaacatgtaatgcatgacgttatttatgagatgggttttatga
tttagagtcccgcaattatacattaatacgcgatagaaaacaaaatatacgccgcaaac
taggataaattatcgccgcgggtgtcatctatgttactagatcgggattactggccg
tcgtttacaacgtcgtgactggaaaaccctggcgtaacccaacttaatcgcccttgca
gcacatcccccttcgcgcagctggcgtaatagcgaagaggcccgcaccgatcgcccttc
15 ccaacagttgcgcagcctgaatggcgcccgtccttcgctttcccttccttc
gccacgttcgcggcttccccgtcaagctctaaatcgaaaaacttgattgggtatggttccg
attttagtgctttacggcacctcgacccccaaaaacttgattgggtatggttcactgta
gtggccatcgccctgatagacggtttcgccttgacggtggagtcacggttcc
aatagtggactcttgttccaaactggaacaacactcaaccctatctcggttattctt
20 tgatttataagggatttgccgattcggaaccaccatcaaacaggatttcgcctgct
gggcaaaaccagcgtggaccgcttgctgcaactctctcaggccaggcggtgaaggca
atcagctgttgcctgtctactggtaaaaagaaaaaccacccagtagtacattaaaaacgt
ccgcaatgtgttattaaagggttctaaaggcgtcaatttgttacaccacaatatacctgc
ca

25

SEQ ID NO:40 is the nucleic acid sequence of MuA-GmCPP. Italicized sequences are the right and left border repeats. Sequence in upper case is the MuA promoter. The *G. max* CaaX prenyl protease sense sequence is in upper case and bold.

30

SEQ ID NO:41

gtttacccgccaatatacctgtcaaacactgatagttaaactgaaggcggaaacga
caatctgatcatgagcggagaattaagggagtacgttatgacccccgcccgtacgcg
ggacaagccgtttacgtttggaaactgacacaaccgcaacgttgaaggagccactcaqc

cgcggtttctggagttaatgagctaaggcacatacgtcagaaaccattattgcgcgtt
caaaagtgcctaaggtaactatcagtagcaaataatttcttgcaaaaatgctccact
gacgttccataaattcccctcggtatccaatttagactctcatattcaactcaatccaa
ataatctgcaccggatctggatcggtcgcatgattgaacaagatggattgcacgcagg
5 ttctccggccgcttgggtggagaggctattcggtatgactggcacaacacagacaatcg
gctgctctgatgccgcgtgtccggctgtcagcgcagggggccccgggtctttgtc
aagaccgacctgtccggtgcctgaatgaactgcaggacgaggcagcgcggctatcg
gctggccacgacggcggtcctgcgcagctgtgcacgttgtactgaagcggaa
gggactggctgtattggcgaagtgccggggcaggatctcctgtcatctcacctgct
10 cctgccgagaaagtatccatcatggctgtgcataatgcggcggctgcatacgcttgc
ggctacctgcccattcgaccaccaagcgaacatcgcatcgagcgcacgtactcgga
tggaaaggccgtcttgcgtcaggatgtggacgaagagcatcagggctcgccca
gccgaactgttcgcaggctcaaggcgcgcatgcccacggcgatgtctcgctgac
ccatggcgatgcctgcgttgcgaatatcatggtgaaaatggccgtttctggattca
15 tcgactgtggccggctgggtgtggcgaccgtatcaggacatagcgttggctaccg
gtatattgcgtgaagagcttggcggcgaatggctgaccgcgttgcgtgttacggtat
cgccgctcccgattcgcagcgcattgcgcattgcgcattgcgcattgcgcattgc
cggactctgggttcgaaatgaccgaccaagcgcacgcccacctgcacgagatt
tcgattccaccgcgccttatgaaagggtggcttcggaatcggtttccggacgcc
20 ggctggatgtcctccagcgcgggatctcatgctggagttttcgccacggatctc
tgccgaacaggcggcgaaggtgccgatattacgcacagcaacggccgacaagcaca
acgccacgatcctgagcgcataatgatcggcccggtccacatcaacggcgtcggc
ggcgcactgcccaggcaagaccgagatgcaccgcgatatttgcgtcgatatttt
cgtggagttccgcacagaccggatgtatccgcattttgcgttcaaacattggcaataaa
25 gtttcttaagattgaatcctgttgcggcttgcgtatgattatcatataattctgttgc
aattacgttaagcatgtataataattacatgtatgcgtacgttattatgagatgggt
tttatgattagagtccgcattatacatattacgcgtatggaaaacaaaatata
gcgcgaaactaggataaattatcgccgcgggtgtcatctatgttactagatcg
tgtcaatgcgtggcggcggctctgggtggttctggcggctctgagggtgggt
30 ctgagggtggcggtctgagggtggcggtctgaggaggcgggtccgggtggct
gggtccgggtgatttgattatgaaaagatggcaaacgctaataaggggctatgaccga
aaatgcgcgtgaaaacgcgcgtacagtctgcgtacgctaaaggcaaacttgc
ctgattacggtgctgtatcgatggttcattgggtgacgtttccggcattgc
aatggtgctactggtgatttgctggctctaattccaaatggctcaagtcgg
gtacgg

tgataattcaccttaatgaataattccgtcaatattacacctccctccctaatcg
ttgaatgtcgccctttgtcttggccaatacgcacaccgcctctccccgcgcgttgg
ccgattcattaatgcagctggcacgcacaggttcccactggaaagcggcagtgagcg
caacgcacatataatgtgagtttagtcactcattaggcacccaggcttacacttatgc
5 ttccggctcgtatgttgttggaaattgtgagcggataacaattcacacaggaaacagc
tatgaccatgattacgccaagcttgcattgcctgcagcccacagatggtagagaggct
acgcagcaggtctcatcaagacgatctacccgagcaataatctccagggaaatcaaatac
cttcccaagaaggtaaagatgcagtcataagattcaggactaactgcattcaagaacac
agagaaagatataattctcaagatcagaagtactattccagttatggacgattcaaggct
10 tgcttcacaaaccaaggcaagtaatagagattggagtctctaaaaaggtagttccact
gaatcaaaaggccatggagtcaaagattcaaatagaggacctaacagaactcgccgtaaa
gactggcgaacagttcatacagactcttacgactcaatgacaagaagaaaatcttcg
tcaacatggtgagcacgacacacttgtctactccaaaaatataagatacagttca
gaagaccaaaaggcaattgagactttcaacaaaggtaatatccggaaacctcctcgg
15 atccattgcccagctatctgtcactttattgtgaagatagtggaaaaggtaggtggct
cctacaaatgccatcattgcgataaaggaaaggccatcgtaagatgcctctggc
agtggccaaagatggaccccccacccacgaggagcatcgtaaaaaagaagacgttcc
aaccacgtttcaagcaagtggattgtgatatctccactgacgtaagggatgacg
cacaatcccactatcctcgcaagacccttccttatataaggaagttcatttcatttg
20 gagagaacacggggactctagaggatccccgggatggcgtttccctacatggaaagccg
ttgtcggtttatgatattaatgtacattttgaaacttacttggatgtgcgacaacat
agggccctcaaacttcctactttccaaagacttttagagggttattcagccaaagagaa
atttgagaaatctagccatatgtttgataaaagccacttcatttttcacgagt
tttgacaaatgtgacagactctacaatttttactttgggtattccctgggtttgg
25 aagaaatcaggagattttatgacaatagctggttcaatgctgagaatgaaatactgca
tacccttgccctttagcaggcgtgatgattggtcacagataacagatttgcctttt
ctctgtactcaactttgtgattgaggccgtatggttataaagcaaacaccatgg
ttattctttagggacatgcttaaggaatttccttctgtataattggccacat
tgtggctgcaatcattgtaatagtagcagaaaggaggtccatactggccatctatctt
30 gggttttacgtttggcttttattgtgatgatgacccttattccagttactaataagct
ccactcttcaataagttcactccacttccagatggcaactcaggagaaaatcgagaa
acttgcttcctccctcaactatccgttaagaaactattgttgcgatggatccacaa
gatcaagtacagcaatgcctatatgtatggattttcaagaacaagaggattgcct
tatgacacatataattcaacagtgcacaaagacgatgagggaaattttgttattgc
ccat

tgagttgggacactggaagctcaaccatactgttacacatttgttatgcagattc
 ttacacttacaatttgaggatataacttagtgcgaaattcagctgttatcgaa
 agctttgggttatacgccagtcctcattggctcatcatatccagcatactgt
 aatcccacttcagcaattggcagctttggctgaaccttagtcagccatattgaat
 5 ttcaggctgtggcttgcagaagaagcttggatatgcattgttgcgtggctt
 gtgaaactacaggaggagaatctgtcagctatgaatacagatcctggactctgctta
 tcactatttcatcctcccttggaaagattggccgcgtggacgaaccggataaga
 aggaagacta**a**gagctcgaaattccccgatcgtaaacatattggcaataaagtttctt
 aagattgaatcctgttgcgcgtttgcgtattatcatataattctgttgaattacg
 10 ttaagcatgtaataattaacatgtaatgcattgttgcgttattttatgagatgggtttatg
 attagagtccgcattatacattaatacgcataaaaaacaaaatagcgcgcaaa
 ctaggataaattatcgccgcgggtgtcatctatgttactagatcggaaattcactggcc
 gtcgtttacaacgtcgtgactggaaaaccctggcgttacccaacttaatgccttgc
 agcacatcccccttcgcagctggcgtaatagcgaagaggcccgcaccgatgcctt
 15 cccaaacagtgcgcgcctgaatggccgcgcgcgcgcgcgcgcgcgcgcgcgcgc
 cgccacgttgc
 gatttagtgtttacggcacctcgacccaaaaacttgattgggtatggttacgt
 agtggccatgcgcctgatagacgggtttgcgcgcgcgcgcgcgcgcgcgcgc
 taatagtggactcttgcgttccaaactggaacaacactcaaccctatctcggctattctt
 20 ttgatttataaggatttgccgatttcggaaaccaccatcaaacaggatttgcctgc
 tggggcaaccagcgtggaccgcttgcgtcaactctctcaggggccaggcggtaagggc
 aatcagctgttgcgcgtctactggtaaaaagaaaaaccacccagtcattaaaaacg
 tccgcaatgtgttattaaagtgtctaagcgtcaattgtttacaccacaatatacctg
 cca

25

SEQ ID NO:41 is the nucleic acid sequence of pBI121-GmCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the 35S promoter. The *G. max* CaaX prenyl protease sense sequence is in bold.

30 SEQ ID NO:42

gttacccgccaatatacctgtcaaacactgtatgtttaaactgaaggcggaaacga
 caatctgatcatgagcggagaattaaggagtcacgttatgaccccccgcgtacgcgc
 ggacaaggccgtttacgtttggactgacagaaccgcaacgttgaaggagccactcagc
 cgcgggttctggagttaatgagctaagcacatacgtcagaaaccattattgcgcgtt

caaaagtgcctaaggtaactatcagtagcaaataattcttgcaaaaatgctccact
gacgttccataaattcccctcggtatccaatttagagtcatattcactctcaatccaa
ataatctgcaccggatctggatcggttcgcattgaacaagatggattgcacgcagg
ttctccggccgttgggtggagaggctattcggtatgactggcacaacagacaatcg
5 gctgctctgatgccgcgtgttccggctgtcagcgcaggggcgcggcgtttttgtc
aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcgtg
gctggccacgacggcggtcctgcgcagctgtgctcagcgttgtcactgaagcggaa
ggactggctgctattggcgaagtgccgggcaggatctcctgtcatctcacctgct
cctgccgagaaagtatccatcatggctgatgcaatgcggcggctgcatacgcttgc
10 ggctacctgcccattcgaccaccaagcgaaacatcgcacgcgacgtactcgga
tggaagccggcttgcgatcaggatgatctggacgaagagcatcaggggctcgccca
gccgaactgttcgcccaggctcaaggcgcatgcggcgtatctcgctgac
ccatggcgatgcctgcttgcgaatatcatggtaaaaatggccgctttctggattca
tcgactgtggccggctgggtgtggcggaccgctatcaggacatagcgttggctaccg
15 gatattgctgaagagcttggcggcgaatgggctgaccgccttcctcgtgttacggat
cgccgctcccgattcgcatcgcatgccttctatgcgccttgcgatcgagttcttgc
cgggactctggggttcgaaatgaccgaccaagcgacgcggccaaacctgcac
tcgattccaccgcgccttatgaaagggtggcttcggaatcgccccggac
ggctggatgatcctccagcgcgggatctcatgctggagtttcgcacggatctc
20 tgccgaacaggcggtcgaagggtgcgatcatcattacgacagcaacggccgaca
acgcacgatcctgagcgacaatatgatcggccggcgtccacatcaacggcgtc
ggcactgcccaggcaagaccgagatgcaccgcgatcttgcgtcgatattt
cgtggagttccgcacagacccggatgatccccgatcgttcaaacattggcaata
gtttcttaagattgaatcctgttgcggcttgcgatgattatcatataattctgtt
25 aattacgttaagcatgtaataattaacatgtaatgcacgttatattatgagatgg
tttatgatttagatgtccgcattatacatattatgcgatagaaaacaaaatata
gcgcgaaactaggataaattatgcgcgcgggtcatctatgttactagatcg
tgtcaatgcggcggcggctctgggtggttctggcggctctgagggtggct
ctgagggtggcgggtctgagggtggcggctctgaggaggcgggtccgggtggct
30 ggttccggtgatttgattatgaaaagatggcaaacgctaataaggggctatgacc
aaatgcgcgatgaaaacgcgcgtacagtctgacgctaaaggcaaacttgattct
ctgattacggtgctgctatcgatggttcattggtgacgttccggccttgcta
aatggtgctactggtgatttgctggctctaattcccaaattggctcaagtc
tgataattcaccttaatgaataattccgtcaatattacctccctcccta
atcgg

ttgaatgtcgccctttgtcttgccaaatacgcaaaccgcctctccccgcgcgttgg
ccgattcataatgcagctggcacgacaggttcccactggaaagcggcagtgagcg
caacgcaattaatgtgagtagtcactcattaggcaccccaggcttacacttatgc
ttccggctcgatgtgtgttggaaattgtgagcggataacaattcacacaggaaacagc
5 tatgaccatgattacgccaagcttgcattgcctgcagcccacagatggtagagaggctt
acgcagcaggtctcatcaagacgatctacccgagcaataatctccagggaaatcaaatac
cttcccaagaaggtaaagatgcagtcaaaaagattcaggactaactgcataagaacac
agagaaagataatttctcaagatcagaagtactattccagtagtgacgattcaaggct
tgcttcacaaaccaaggcaagtaatagagattggagtctctaaaaaggtagttcccact
10 aatcaaaaggccatggagtcaaagatcaaataagaggacctaacagaactcgccgtaaa
gactggcgaacagttcatacagagtcttacgactcaatgacaagaagaaaatcttcg
tcaacatggggagcacgacacacttgtctactccaaaaatatcaaagatacagtctca
gaagaccaaaaggcaattgagactttcaacaaaggtaatatccggaaacctcctcgg
attccattgccagctatctgtcactttattgtgaagatagtggaaaaggaaggtaggtggct
15 cctacaaaatgccatattgcgataaaggaaaggccatcgtaagatgcctctggc
agtggtccaaagatggaccccccaccacgaggagcatcgtaaaaaagaagacgttcc
aaccacgtcttcaaagcaagtggattgtgatatctccactgacgtaaggatgacg
cacaatcccactatcctcgcaagacccttccttatataaggaagttcatttcatttg
gagagaacacggggactctaga**ccggttcgccagcgcccaatcttcaacaaggg**
20 **gaggatgagaatagtgataa**gcagagtaccaaggatctgtattcatagctgacagattc
tcctcctgttagttcacaagaccaccgcgtaatccagatgcata~~cc~~atccagcttcttggc
aaagccatcagcctgaaattcaa~~at~~gtatcggtacttagttcagaccaagctgacca
attgctgaagtggattacagtatgtgaaatatgtgagccaaatgaggactggctgc
gtatcaaacccaaagcttcgatacagatcagctgaaattcgcactagtgtatatcctcc
25 aaattgtagaagtgtaaatctgcata~~cc~~atccatcgatcttgcactgttgcactgt
tccagtgtcccaactcatggcaataacagcaacaatttcctcatcgatcttgcactgt
tgaattatgtgtcataaggacaatcctcttgcactgttgcactgttgcactgt
attgctgtgacttgatcttgcgttgcactgttgcactgttgcactgttgcactgt
GTGGCAGTGAAGGGCGAACAGTTCTGATTAACCACAAACCGTTCTACTTACTGGCTT
30 TGGTCGTCAAGAGATGCGGACTTGCGTGGCAAAGGATTGATAACGTGCTGATGGTGC
ACGACCACGCATTAATGGACTGGATTGGGGCCAACCTCCTACCGTACCTCGCATTACCCCT
TACGCTGAAGAGATGCTCGACTGGCAGATGAACATGGCATCGTGGTATTGATGAAAC
TGCTGCTGTCGGCTTTCGCTCTTTAGGCATTGGTTCGAAGCAGGGCAACAAGCCGA
AAGAACTGTACAGCGAAGAGGCAGTCAACGGGGAAACTCAGCAAGCGCACTTACAGGCG

ggtaaaaagaaaaaccacccagtagtacattaaaacgtccgcaatgtgttattaagtgt
 ctaagcgtcaattt~~gttt~~acaccacaata~~tat~~cctgcca

SEQ ID NO:42 is the nucleic acid sequence of pBI121-HP-GmCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the 35S promoter. Bold sequence is the antisense prenyl protease fragment of *G. max*. Bold and underlined sequence is the *G. max* sense prenyl protease fragment and sequence in upper case is the truncated GUS fragment.

10 SEQ ID NO:43

*gtttacccgccaatatacctgtcaa*actgtatggacttactgaaggcggaaacga
 caatctgatcatgagcggagaattaaggagtcacgttatgacccccgccc~~at~~gacgcg
 ggacaagccgtttacgtt~~gg~~actgacagaaccgcaacgttgaaggagccactcagc
 cgcggtttctggagttaatgagctaagcacatacgtcagaaaccattattgcgcgtt
 15 caaaaagtgcctaaggtcactatcagctagcaaataatttctgtcaaaaatgctccact
 gacgttccataaattcccctcggtatccaatttagagtctcatattcactctcaatccaa
 ataatctgcaccggatctggatcg~~tt~~cgatgattgaacaagatggattgcacgcagg
 ttctccggccgcttgggtggagaggctattcggtatgactggcacaacagacaatcg
 gctgctctgatgccgcgtgttccggctgtcagcgcagggcgccgg~~tt~~ctttgtc
 20 aagaccgac~~ctgtccgg~~gtccctgaatgaactgcaggacgaggcagcgcggctatcg~~t~~
 gctggccacgcggcg~~tt~~c~~tt~~gcgcagctgtgc~~tc~~acgttgcactgaagcggaa
 gggactggctgctattggcgaa~~gt~~tgccggcaggatctc~~ct~~gtcatctcac~~tt~~gct
 cctg~~cc~~gagaa~~gt~~atccatcatggctgatgcaatgcggcggctgcatacgctt~~at~~cc
 ggctac~~ct~~gcccattcgaccaccaagcgaa~~ac~~atcgcatcgagc~~g~~acgtactcg~~ga~~
 25 tggaa~~gg~~gttgc~~at~~caggatgatctggacgaagagcatcagg~~gg~~ctcg~~cc~~cca
 g~~cc~~gaactgtcgccaggctcaaggcg~~cg~~catg~~cc~~gacggc~~at~~gatctcg~~tc~~gtgac
 ccatggcgatgcctg~~ctt~~gcgaat~~at~~catgg~~tt~~ggaaaatggccg~~ttt~~ctggattca
 tcgactgtggccggctgggtgtggcggaccgctatcaggacatagcgttggctacc~~gt~~
 gatattgctgaagagcttggcggc~~ga~~atgg~~gg~~ctgaccgctt~~cc~~ctcg~~tg~~cttac~~gg~~at
 30 cgccgctcccattcg~~ac~~gcgc~~at~~cg~~cc~~ttctatcg~~cc~~tttgc~~ac~~gagtt~~tt~~ctcg~~ag~~
 cggactctgggttcgaa~~at~~gaccgaccaagcgacgc~~cc~~aaac~~ct~~g~~cc~~atc~~ac~~gagatt
 tcgattccaccgcgc~~cc~~ttctatgaa~~agg~~ttggctt~~cc~~g~~aa~~atcg~~ttt~~ccggacg~~cc~~
 ggctggatgatcctccagcg~~cc~~gggatctcatg~~ct~~ggagtt~~cc~~g~~cc~~acggatctc
 tgc~~gg~~aa~~ac~~aggc~~gg~~tcg~~a~~agg~~gt~~g~~cc~~g~~at~~atcattacgacagcaacggccgaca~~ag~~caca

acgccacgatcctgagcgacaatatgatcggcccccgtccacatcaacggcgtcggc
ggcgactgcccaggcaagaccgagatgcaccgcgatatcttgctgcgttggatatttt
cgtggagttccccccacagacccggatgatccccgatcgtaaaacattggcaataaa
gtttcttaagattgaatcctgttgcggcttgcgtgattatcatataattctgttg
5 aattacgttaagcatgtataattacatgtaatgcgtacgttattatgagatgggt
tttatgatttagagtcccgcaattatacattaaatacgcgatagaaaacaaaatatacg
gcgcaaactaggataaattatcgccgcgggtcatctatgttactagatcgggcctcc
tgtcaatgctggcggcggctctggtggtggctcgaggaggcggtccggtggtggct
ctgagggtggcggtctgagggtggcgctctgaggaggcggtccggtggtggct
10 ggttccggtgatttgattatgaaaagatggcaaacgctaataaggggctatgaccga
aaatgccatgaaaacgcgctacagtctgacgctaaaggcaaacttgcgtatgc
ctgattacggtgctgctatcgatggttcattggtgacggttcggccttgctaattgg
aatggtgctactggtgatttgctggctctaattccaaatggctcaagtgggtgacgg
tgataattcaccttaatgaataattccgtcaatattacacctccctccctcaatcgg
15 ttgaatgtccccctttgtcttgcccaatacgc当地accgcctctccccggcggtgg
ccgattcattaatgcagctggcacgacaggttcccactggaaaagcggcagtgg
caacgcaattaatgtgagtttagctcactcattaggcacccaggcttacacttatgc
ttccggctcgatgtgtggattgtgagcggataacaattcacacaggaaacagc
tatgaccatgattacgccaagcttgcattgcctgcagcccacagatggtagagaggct
20 acgcagcaggtctcatcaagacgatctaccgagcaataatctccaggaaatcaaatac
cttcccaagaaggtaaagatgcagtcaaaagattcaggactaactgcataagaacac
agagaaagatatttctcaagatcagaagtactattccagttatggacgattcaaggct
tgcttcacaaaccaaggcaagtaatagagattggagttctctaaaaaggtagttccact
gaatcaaaggccatggagtcaaagattcaaatagaggacctaacagaactcgccgtaaa
25 gactggcgaacagttcatacagagtctttacacttgcactcaatgacaagaagaaaatcttcg
tcaacatggtgagcacgacacacttgcacttccaaaaatataagatacagtctca
gaagaccaaaggcaattgagactttcaacaaaggtaatatccggaaacctcctcgg
attccattgccatctgtcactttattgtgaagatagtggaaaaggtaggtggct
cctacaaatgccatcattgcataaaggaaaggccatcgtaagatgcctctgcccac
30 agtggtcccaaagatggaccccccaccacgaggagcatcgatggaaaaagaagacgttcc
aaccacgttccaaagcaagtggattgtgatatctccactgcgtggaaaaagaagacgttcc
cacaatcccactatccttcgcaagacccttccttatataaggaaagttcatttgc
gagagaacacggggactctagaggatccccgggttagtcttccttatccgggttcg
tccagcgccaaatcttcaacaaggaggatgagaatagtgataagcagagttacca

agatctgtattcatagctgacagatttcctcctgttagttcacaagaccaccgcgta
 atccagatgcataatccaagcttctggcaaagccatcagcctgaaattcaaatgatcg
 ctgacttaggttcagaccaaagctgaccaattgctgaagtgggattacagtatgctgaaa
 tatgtatgagccaatgaggactggctgcgtatcaaacccaaagcttcgatacagatcag
 5 ctgaatttcgcactagtgtatccctccaaattgttagaagtgtagaatctgcatacagca
 acaaattgttacacagtatggttagtccagtgtcccaactcatggcaataacagc
 aacaatttcctcatcgctttgcactgttgaattaatgtgtcataaggacaatcctct
 tgttcttgaagaatccatacatataggcattgtgtacttgatcttgtggatccatcg
 acaacaaatagttcttaacggatagttgagggaggaaggcaagttctcgatttc
 10 cctgagttgaccatctggaagtggagtgaacttattgaagagtggagctattgtactg
 gataaagggtcatcatcacaatagaaagaccaaacgtaaaaacccaaagatagatggcc
 aagtatggaccccttctgtactattacaatgattgcagccacaatagggtggaccaat
 tattacagaaaggaaaattccttaagcatgtccctaaagaataaccatgggtttgct
 tattaaaaccatgacgggcctcaatcacaaaagttgagttacagagaaaagggcaaatct
 15 gttatctgtgaccaaatcatcagccctgctaagaaggcaagggtatgcagtatttcatt
 ctcagcattgaaaccagctattgtcataaaatctcctgatttctccaaaaccaggca
 ataccccaaagtacaaaattgttagagtctgtcactattgtcacaactcgtgaacaaaa
 tggaaagtggctttatcaagactataggcttagattctcaatttcttgtgtgat
 aacaccctctaaagtcttggaaagagtaggaagttgagggccatgttgcacat
 20 ccaagtaagttcaaaaatgtacattaatcataatccgacaacggcttcatgtag
 ggaaacgcccattagctcgaattccccgatcgttcaaacacattggcaataaagttctt
 aagattgaatcctgttgcggcttgcgtatttatcatataattctgttgaattacg
 tttagcatgtaataattaacatgtaatgcacgttattatgagatgggtttatg
 attagagtccgcattatacatttataacgcgtatggatggggatccatgtgcgc
 25 ctaggataaattatcgcgccgggtgtcatctatgttacttagatcgggaattcactggcc
 gtcgttttacaacgtcgtgactggaaaaccctggcgttacccaacttaatcgccct
 agcacatcccccttgcgcagctggcgtaatagcgaagaggccgcaccgatcgccct
 cccacagttgcgcagcctgaaatggccccgcttcgcattttcccttc
 cgccacgttgcgcggcttccccgtcaagcttaatcggggctccctttagggtcc
 30 gattttagtgcttacggcacctcgacccaaaaacttgattgggtatggttcacgt
 agtgggcattcgccctgatagacggttttcgcccttgacgttggagttccacgtt
 taatagtggactcttgccttcaaactggaacaacactcaaccctatctcggctattctt
 ttgattataaggatttgccgatttcggaaaccaccatcaaacaggatttcgc
 tggggcaaaccagcgtggaccgcttgcaactctcagggccaggcggtgaaggc

aatcagctgttgcggcttcactggtgaaaagaaaaaccacccagttacataaaaacg
tccgcaatgtgttattaaaggcttaagcgtaatggttacaccacaatatacctg
cca

SEQ ID NO:43 is the nucleic acid sequence of pBI121-antisense-GmCPP.

Italics sequences are the right and left border repeats. **Underline** sequence is the 35S promoter. **Bold** sequence is the GmCPP anti-sense sequence.

SEQ ID NO:44

5 ggtccgatttagtgcttacggcacctcgacccaaaaactgattgggtatgg
 tcacgtatggccatgcctgataagacggtttcgccttgacgtggagtcac
 gttcttaatagtggactctgttccaaactggaacaacactcaaccstatctcggc
 attctttgattataaggatttgcgattcggaccaccaaaacaggatttc
 10 gcctgctgggcaaaccagcgtggaccgcttgcaactcttcagggccaggcg
 aaggcaatcagctgtgccgtctcactggaaaagaaaaaccacccagtcatta
 aaaacgtccgcaatgtttattaagttgtctaagcgtcaattgtttacaccacaat
 atcctgcca

10 SEQ ID NO:44 is the nucleic acid sequence of pRD29A-GmCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the RD29A promoter. Sequence in bold is the GmCPP sense sequence.

SEQ ID NO:45

15 *gttacccgccaatatacctgtcaaacactgatagttaaactgaaggcggaaacga*
 caatctgatcatgagcggagaattaaggagtacgttatgaccccgccatgacgcg
 ggacaagccgtttacgtttgaactgacagaaccgcaacgttgaaggagccactcagc
 cgccggtttctggagttaatgagctaacatcgtcagaaaccattattgcgcgtt
 caaaagtgcctaaggtcactatcagtagcaaattttctgtaaaaatgctccact
 20 gacgttccataaattccccctcggtatccaatttagagtctcatattcactctcaatccaa
 ataatctgcaccggatctggatcggttcgcatttgcatttgcacgcagg
 ttctccggccgttgggtggagaggctattcggtatgactggcacaacagacaatcg
 gctgctctgatgcgcgtgttccggctgtcagcgcaggggcgcccggtttttgc
 aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcgt
 25 gctggccacgacggcggttccgcagctgtgcgtacgttgtcactgaagcggaa
 gggactggctgttggcgaaatgcggggcaggatctcgtcatctcaccttgc
 cctggcgagaaatccatcatggctgtatgcattgcggcggtgcatacgcttgc
 ggctacctgcccattcgaccaccaaggcatacgcatcgagcgcagcacgtactcg
 tggaaagccggcttgcgtatcaggatgttgcgtatggacgaaagacatcagg
 30 gccgaactgttgcgcaggctcaaggcgcgcattgcgcgtatgcggcgatgtatcgt
 ccatggcgatgcctgcgttgcgaatcatggggaaatggccgtttctggattca
 tcgactgtggccggctgggtgtggcgaccgctatcaggacatgcgttgcgtacc
 gatattgtgaagagcttggcgcaatggctgaccgttcctcgtgcgttacggat
 35 cggactctgggttcgaaatgaccgaccaaggcgcgcacgcggccatcagc
 tcgattccaccgcgccttatgaaagggtggcttcggaaatcggttccggacgc
 ggctggatgtatcctccagcgcgggatctcatgtggagtttcgcgcacggatctc

tgccgaacaggcggtcgaagggtgccgatattacgcacagcaacggccgacaagcaca
acgccacgatcctgagcgacaatatgatcgggcccggcgtccacatcaacggcgtcggc
ggcactgcccaggcaagaccgagatgcaccgcgatattcgctgcgttcggatattt
cgtggagttcccgcacagacccggatgatccccgatcgtaaaacattggcaataaaa
5 gtttcttaagattgaatcctgttgcggcttgcgtattatcatataattctgttg
aattacgttaagcatgtataattaaatcatgtatgcacgttatttatgagatgggt
tttatgatttagagtccgcattatacattaaatacgcataaaaaacaaaatatacg
gcgcaaactaggataaattatcgccgcgggtcatctatgttactagatcggcctcc
tgtcaatgctggcggcggctctggtggtggcttgcggctctgagggtggct
10 ctgagggtggcggttctgagggtggcggctctgaggaggcggttccggtggtggct
ggttccggtgatttgattatgaaaagatggcaaacgctaataaggggctatgaccga
aaatgccatgaaaacgcgctacagtctgacgctaaaggcaaacttgattctgtcgcta
ctgattacggtgctgctatcgatggttcattggtgacgttccggccttgcataatggt
aatggtgctactggtagttgctggctctaattccaaatggctcaagtcggtgacgg
15 tgataattcaccttaatgaataattccgtcaatattacctccctccctcaatcg
ttgaatgtcgccctttgtcttggcccaatacgcaaaccgcctctcccgccgttgg
ccgattcattaatgcagctggcacgacaggttcccactggaaagcggcagtgagcg
caacgcaattaatgtgagtttagctcactcattaggcacccaggcttacactttatgc
ttccggctcgatgtgtggatttgagcggataacaattcacacagggaaacagc
20 tatgaccatgattacgccaagcttgcattgcgcggcatagatgcaattcaatc
aaactgaaattctgcaagaatctcaaacacggagatctcaaagttgaaagaaaattt
atttctcgactcaaaacaaactacgaaatttaggtagaacttatatacattatattt
taattttttaacaaaatgttttattattatagaattttactggttaattaaa
aatgaatagaaaaggtgaattaagaggagagaggagtaaacatttcttattttt
25 catatttcaggataaatttgcataagtttacaagattccatttgcattttactgt
tgaggaatttctcttagtaagatcatttcatctacttctttatcttctaccagta
gaggaataaacaatatttagctccttgcataacatttgcatttttgcatttgcatt
attcaatttaatttacgtataaaataaaagatcatacatttgcatttgcatttgcatt
aaatacaattcgaatgagaaggatgtgcgttgcatttgcatttgcatttgcatttgcatt
30 aacgtaaaatgaccacatgatggccaatagacatggaccgactactaataatagtaag
ttacatttttaggatgaaataatcataccgacatcagtttgcatttgcatttgcatt
aaagaaaaataaataaaagatatactaccgacatgatgcatttgcatttgcatttgcatt
atcaagccgacacagacacgcgttagagagcaaaatgactttgcatttgcatttgcatt
cagacgcttcatacgtgtcccttattctctcagtcatttgcatttgcatttgcatttgcatt

ttcaggctgatggcttccaagaagcttgatatgcacatctggattacgcgggtgtctt
gtgaaaactacaggaggagaatctgtcagctatgaatacagatccctggtaactctgctta
tcactatttcatcctcccccttggaaagattggccgcgtggacgaaccggagctc
 gaatttccccgatcgttcaaacaatttggcaataaagtttcttaagattgaatccctgttg
 5 ccggcttgcgtattatcatataatttctgttgaattacgttaagcatgtataatt
 aacatgtaatgcacgttatttatgagatgggttttatgatttagagtcccgcatt
 atacatttaatacgcatagaaaaacaaaatagcgccaaactaggataaattatcgc
 gcgcgggtgtcatctatgttactagatcgggaattcactggccgtcgtaacaacgtcg
 tgactggaaaaccctggcgttacccaacttaatgccttgcagcacatcccccttcg
 10 ccagctggcgtaatagcgaagaggcccgcaccgatgcgccttccaaacagttgcgcagc
 ctgaatggcgcccgtccttcgtcttcccttcgtccacgttcgcggct
 ttccccgtcaagctctaaatcggggctccctttagggtccgatttagtgcttacgg
 cacctcgaccccaaaaaacttgattgggtgatggttcacgttagtggccatgcgcctg
 atagacggttttcgcccttgacgttgagttccacgttcttaatagtggactctgt
 15 tccaaactggaaacaacactcaaccatatctcggttattctttgatttataaggatt
 ttgccgatttcggaaccaccatcaaacaggatttcgcctgctgggcaaacaccagcgtg
 gaccgcttgcactctcagggccaggcggtgaaggcaatcagctgttgcggct
 ctcactggtaaaagaaaaaccacccagttacattaaacgtccgcaatgtgttatta
 agtgtctaagcgtcaatttgcgttacaccacaatatacctgcca

20

SEQ ID NO:45 is the nucleic acid sequence of pRD29A-HP-GmCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the RD29A promoter. Sequence in bold is the GmCPP antisense sequence, bold and underlined sequence is the GmCPP sense sequence.

25

SEQ ID NO:46

gtttacccgcataatataatccgtcaaacactgtatgtttaaactgaaggcgggaaacgcacaatctgtatgagcggaga
attaagggagtcacgttatgaccccccgcgtacgcgggacaagccgtttacgttggactgcagaaccgcacg
ttgaaggagccactcagecgccggttctggagttaatgagctaagcacatacgtcagaaaccattattgcgegttcaa
 30 *aagtcgcctaaggtcaactatcagtcagtcagaaatattttgtcaaaaatgtccactgcgttccataaattccccctcggtat*
ccaaatttagagtcataatcactctcaatccaaataatctgcacccggatctggatgttcgcattgttgcataacaagatgga
ttgcacgcagggtctccggccgttgggtggagaggcttgcgtatgcgttgcacaaacagacaatcggtctgtca
tgcgcgggtgttcggctgtcagcgcaggggcgcgggtttttgtcaagaccgcacgttgcgttgcgttgcacatgttgcgtatgtca
 35 *gcaggacgaggcagcgcggctatgtggctggcacaacgcgggttgcgttgcgttgcacatgttgcgtatgtca*
gccccggaaaggactggcttattggcgaagtggccggggcaggatctctgtatcgttgcacatgttgcgttgcgttgcacatgttgcgtatgtca
tatccatgttgcgtatgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgtatgtca
cgcacgttgcgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgtatgtca
gccccggacactgttgcgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgttgcacatgttgcgtatgtca

5
10
15
20
25
30
35
40
45
50

gettgcgaatatcatggaaaaatggcgctttctggattcatcgactgtggccggctgggtgtggccgaccget
aggacatacggttgcgetacccgtatgtgaagagctggggagaatggcgtgaccgttcctctgtgttaeggt
atcgccgtccccatcgccatcgccctatcgcccttgcagcgttcttgcagcggactctgggttcgaaat
gaccgaccaagcgacgccccaaacctgcacatcagcggatttcgattccacccgegeetctatgaaagggtggcttgcgaaat
atcggttccggacgcccgtggatgtatctccagcggggatctcatcggttgcggatcttgcacccgtctcg
gaacagggcgatcgaaagggtccgatattacgacagcaacggccgacaagcacaacgcaacgatctgagc
atatgatecgcccccgggtccacatcaacgggtcgccggactgcggcaggcaagcggatgcacccgtatct
tgcgttgtccgatatttgcggatcccgccacagacccggatgcgttcaaaacatttgcgaaataagtt
cttaagattgaatctgtgcggcttgcgtatgattcatataatttgcgttgcattacgttaagcatgtataatttacat
gtaatgcgtacgttattatgagatgggtttatgatttagttagtcccgcaattatacattaaacg
aatatacgccgaaactaggataaattatcgccgcgggtgtcatctatgttacttagatcgccctctgtcaatgtgg
ggggcggcttgcgggtggatctgggtccggatgtttgattatgaaaagatggcaaaacgctaataaggggctatg
acggaaaatcccgatgaaaacggegetacagtgcgttcaatttgcgttgcgtactgttgcgttgcgt
ctatcgatggttcattggtgcgttccggcttgcgtatgttgcgtactgttgcgttgcgttgcgt
atggcgtcaagtccggtgcgttgcgtataattcacattatgaaataatttgcgttcaatatttacccctcccaatcggt
atgtgcgcctttgtcttggccaatacgcaaaacgcctcccgccgttgcgttgcgttgcgt
cagggttcccgactggaaagcgggcagtgcggcgttgcgttgcgttgcgttgcgttgcgt
ttacactttatgttccggctgtatgttgcgttgcgttgcgttgcgttgcgttgcgt
gattacgccaagcttgcgttgcgttgcgttgcgttgcgttgcgttgcgttgcgttgcgt
acggagatctcaagttgaaagaaaatttttgcactcaaaacaaacttgcgttgcgttgcgttgcgt
ttatattgtattttgcgttgcgttgcgttgcgttgcgttgcgttgcgttgcgt
aattaagaggagagaggaggttgcgttgcgttgcgttgcgttgcgttgcgt
ccatttgactgtgtaaatgaggaaatttgcgttgcgttgcgttgcgttgcgt
aaacaatatttgcgttgcgttgcgttgcgttgcgttgcgttgcgt
atcatacatttgcgttgcgttgcgttgcgttgcgttgcgttgcgt
cgacgttacgttgcgttgcgttgcgttgcgttgcgttgcgt
tgaataatcataccgcacatgcgttgcgttgcgttgcgttgcgt
acatgatgttccaaaagcaaaaaagatcaagccgacacagacacg
caccacgaaaacagacgttcatatgcgttgcgttgcgttgcgt
ctcacaatatgcgttgcgttgcgttgcgttgcgttgcgt
cccggttagtctcttgcgttgcgttgcgttgcgttgcgt
gtaccaaggatctgttgcgttgcgttgcgttgcgt
cttgcgttgcgttgcgttgcgttgcgttgcgt
gtatgttgcgttgcgttgcgttgcgttgcgttgcgt
gttatatcctccaaatttgcgttgcgttgcgttgcgt
catggcgttgcgttgcgttgcgttgcgt
ccatcatataggcattgtgttgcgttgcgttgcgt
agtttgcgttgcgttgcgttgcgt
tcatcacaatagaaagaccaacgtaaaaacccaaagat
gccacaataggfggaccaatttacagaaaggaaaatttgcgttgcgt
ccatgacggccctcaatcacaaaatgttgcgttgcgt
aaggcaagggtatgcgttgcgttgcgt
ccccaaagtacaaaatttgcgttgcgt
tagatttgcgttgcgttgcgt
aagtttgcgttgcgt
atcggttcaacacatttgcgttgcgt
attacgttgcgt
tacatttgcgttgcgt
agatcggttgcgttgcgt

5 **gcacatecccccttgcgcagctggcgtaatagcgaagaggcccgcaccgatgcgcctcccaacagtgcgcagecgtga
atggcgcccgcttcgtttcccttccttcgtccacgttcgcgggtttcccgtaagctataatcggggctc
ccttagggtccgatttagtgettaacggcactcgaccccaaaaacttgatgggtatggtcacgttaggggcat
gcgcctgtatagacgggtttgcgccttgacgttggacttcacgtttaatagtgactttgtccaaactggaaacaac
acteaaccctatcggcgtattttgtatttataagggatttgcgcatttgcgaaccaccatcaaacaggatttgcgc
gttgggcaaaaccagcgtggaccgtgcactctcaggccaggcggtaaggcgaatcagctgtgcgc
tcactggtaaaagaaaaaccacccactacattaaaaacgtccgcaatgttgttataagttgtctaagcgtcaatttgtt
tacaccacaatatcctgcca**

10 SEQ ID NO:46 is the nucleic acid sequence of pRD29A-antisense-GmCPP.

Italicized sequences are the right and left border repeats. Underlined sequence is the RD29A promoter. Sequence in bold is the GmCPP antisense sequence.

SEQ ID NO:47

15 *gttacccgccaatatacctgtcaaacactgatagttaaactgaaggcggaaacga
caatctgatcatgagcggagaattaaggagtcacgttatgaccccccgcgtacgcgc
ggacaagccgtttacgtttggactgacagaaccgcaacgttgaaggagccactcagc
cgccggtttctggagttaatgagctaagcacatacgtcagaaaccattattgcgcgtt
caaaagtgcctaaggtcactatcagtagcaaataatttcttgcataaaatgctccact
20 gacgttccataaattccctcggtatccaatttagtctcatattcactctcaatccaa
ataatctgcaccggatctggatcggttcgcatttgcatttgcacgcagg
ttctccggccgcttgggtggagaggctattcggtatgcactgggcacaacagacaatcg
gctgctctgatgccgcgtgttccggctgtcagcgcaggggcgcgggttctttgtc
aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcg
25 gctggccacgacggcggtcctgcgcagctgtgcgcacgttgcactgaagcggaa
gggactggctgtattggcgaagtgcggggcaggatctcctgtcatctcaccttgct
cctgcgcagaaagtatccatcatggctgtcaatgcggcggctgcatacgcttgc
ggctacctgcccattcgaccaccaagcgaacatcgcatcgagcgcacgtactcg
tggaaagccggcttgcgtcattcaggatgtctggacgaagagcatcaggggctcgcc
30 gccgaactgttcgcaggctcaaggcgcatgcggcgtatgcgcgc
ccatggcgatgcctgcattgcgaatatcatggggaaatggccgtttctggattca
tcgactgtggccggctgggtgtggcgaccgctatcaggacatagcgttgcgttacccgt
gatattgcgtgaagagcttggcgccgaatgggtgcaccgcgttgcgttgcgttacccgt
cgccgcgtcccgattcgcagcgcattgcgcattgcgcattgcgcattgcgc
35 cgggactctgggttcgaaatgaccgaccaagcgcacgcggccacactgcgc
tcgattccaccgcgcattctatgaaagggtggcttcggaaatcggttccggacgc
ggctggatgatcctccagcgcgggatctcatgcgttgcgcgcacggatctc*

tgcggAACAGGCGGTcgaagggtgccatATCATTACGACAGCAACGGCCGACAAGCACA
acGCCACGATCCTGAGCGACAATATGATCGGGCCCAGGTCCACATCAACGGCGTCGGC
GGCGACTGCCAGGAAGACCGAGATGCACCGCGATATCTTGCTGCGTTGGATATTT
CGTGGAGTTCCCAGCACAGACCCGGATGATCCCCGATCGTTCAAACATTGGCAATAAA
5 gtttcttaagattgaatcctgttgcggcttgcgtattatcatataatttcgttg
aattacgttaagcatgtataatacatgtaatgcacgttatTTTgagatgggt
tttatgatttagagtcccgaattatacatTTAATACGCGATAAGAAAACAAAATATAGC
GCGCAAACTAGGATAAATTATCGCGCGGTGTCATCTATGTTACTAGATCGGGCCTCC
tgtcaatgctggcgccggctctggtggtggctcggtggcggctctgagggtggct
10 ctgagggtggcggtctgagggtggcggtctgagggaggcggtccgggtggct
ggttccggtgattttgattatgaaaAGATGGCAAACGCTAATAAGGGGGCTATGACC
aaatGCCATGAAAACGCGCTACAGTCTGACGCTAAAGGCAAACTTGATTCTGTCGCTA
CTGATTACGGTGCTGCTATCGATGGTTCTGGTGACGTTCCGGCCTGCTAATGGT
AAATGGTGCTACTGGTGTGATTGCTGGCTCTAATTCCAAATGGCTCAAGTCGGTACGG
15 TGATAATTACCTTAATGAATAATTCCGTCATATTACCTCCCTCCCTCAATCGG
TTGAATGTCGCCCTTGTCTTGGCCAATACGCAAACGCCCTCTCCCGCGCGTGG
CCGATTCTTAATGCAGCTGGCACGACAGGTTCCGACTGGAAAGCGGGCAGTGAGCG
CAACGCAATTATGTGAGTTAGCTCACTCATTAGGCACCCAGGCTTACACTTATGC
TTCCGGCTCGTATGTTGTGGAATTGTGAGCGGATAACAATTACACAGGAAACAGC
20 TATGACCATGATTACGCCAAGCTTGCATGCCCTGCAGCCACAGATGGTTAGAGAGGCTT
ACGAGCAGGTCTCATCAAGACGATCTACCGAGCAATACTCCAGGAAATCAAACAC
CTTCCCAAGAAGGTTAAAGATGCAAGTCAGGACTAATGCAAGAACACAG
AGAGAAAGATATATTCTCAAGATCAGAAGTACTATTCCAGTATGGACGATTCAAGGCT
TGCTTCACAAACCAAGGCAAGTAATAGAGATTGGAGTCTCTAAAAGGTTAGTCCCAC
25 GAATCAAAGGCCATGGAGTCAGGAAAGATTCAAATAGAGGACCTAACAGAACACTCGCCGTA
AAAGACTGGCGAACAGTCATCACAGAGTCCTTACGACTCAATGACAAGAAAATCTCG
TCAACATGGTGGAGCACGACACACTTGTCTACTCCAAAATATCAAAGATACTCG
GAAGACCAAAAGGGCAATTGAGACTTTCAACAAAGGGTAATATCCGAAACCTCCTCGG
ATTCCATTGCCAGCTATCTGTCACCTTATTGTGAAGATAGTGGAAAAGGAAGGTTGGCT
30 CCTACAAATGCCATCATTGCGATAAAGGAAAGGCCATCGTTGAAGATGCCCTGCGAC
AGTGGTCCCAAAGATGGACCCCCACCCACGAGGAGCATCGTGGAAAAAGAACGTTCC
AACCACGTCCTCAAGCAAGTGGATTGATGTGATATCTCCACTGACGTAAGGGATGACG
CACAATCCCACATCCTCGCAAGACCCCTCTATATAAGGAAGTTCAATTGATTG
GAGAGAACACGGGGACTCTAGAGGATCCATGGCGATTCCATTGAAACCGTCGTT

ggtttatgatagtatgtacgtttgagacgtattggatctgaggcaacatactgc
tctcaagctccactctccaaagactttgggtggactcattagccaagagaagtttg
agaaaatctcgagcttacagtcttgacaaaagccatttcacattttcatgagttgtt
actatacttatggactctgcgattctgttcttggatctgcctgggtttggaaagat
5 atctggcggcttctaccaatggggactcgatccagagaatgaaatcctgcacactc
tttcattcttggctggcttatgacatggcacagatcaactgatttgcattttcttg
tactcaacttcgtatcgagtctcgcatgggtcaacaaacaatatggatgtt
cattaggacatgatcaaaggaaatactcctctgtcatacctgcccctctatcggt
ccgcaattattgttagttcagaaaggaggccttacctcgccatctatctgtggca
10 ttcatgttatacctgtcttagttagttagtactataccctgtttgattgcacactc
tttcaacaagttcactcctcttgcgtggagaccccccggagaagattgagaaacttg
cttcttcctaaagttcctctgaagaagctgtttgtcgatggatctacaaggta
agccatagtaatgcttacatgtatggtttcaagaacaaaaggattgttctttaga
cacattgattcagcagtgccagaatgagaatgaaattgtggcggtattgcacacgagc
15 tggacactggaagctgaatcacactacatactcggttcaatccttgc
ttcttgcattttggaggatacactcttgcgtggaaactccactgatcttcaggagtt
tggtttgcatacacaaccagttctcattgggttgcattttcagcacactgttaatac
cacttcaacacccatgtttgaccaacccattgtactcagcttgc
gctgatgcatttgcagtgaatcttggttatgcacggatctacgtccctgccttagtga
20 gctacaggaagagaacttatcagcgatgaacacagacccattgtactcagcttact
actcacacccctccttgcattttgcattttggaggcttcgagccattgtactcagcttact
gattaaacccctcgatccatgttttttttttttttttttttttttttttttttttttttt
tgaatcctgttgcggcttgcgtatcgatgattatcatataatttctgttgcatttgc
catgtataataatttgcatt
25 agtcccccaattatacatgtttatgcgtatggaaaacaaaatatacgccgcacactgg
ataaaattatcgccgcgggtcatctatgttacttagatcggttttttttttttttttt
tttacaacgtcgactggaaaacccctggcgtaatcccaactttaatcgcccttgc
atcccccttcgcagctggcgtaatagcgaagaggccgcacccatgttgc
cagttgcgcagccctgatggcgccgcctttcgcttttttttttttttttttttttt
30 cgttcgccggcttcccgtaagcttaatcgccgcgggttttttttttttttttttttt
agtgcatttacggcacctcgacccaaaaacttgatgggtgatggttcacgttagtgg
gccatcgccctgatagacggttttcgcccttgcgtggagtcacgttccacgttca
gtggactcttgcatttccaaactggaaacaacactcaaccctatctcggttatttttt
tataagggttttgcgatccggaaaccaccatcaaacaggatttgcctgtgg

caaaccagcgtggaccgcttgctgcaactctctcagggccaggcggtgaagggaatca
gctgttgcggctctcactggtaaaaagaaaaaccacccaggatactaaaaacgtccgc
aatgtgttattaagtgtctaagcgtcaattgtttacaccacaatatacctgcca

5 SEQ ID NO:47 is the nucleic acid sequence of pBI121-BnCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the 35S promoter. Sequence in bold is the BnCPP antisense sequence.

SEQ ID NO:48

10 *gtttacccgccaatatacctgtcaa*acactgatagttaaactgaaggcggaaacga
caatctgatcatgagcggagaattaaggagtcacgttatgaccccccggatgacgca
ggacaagccgtttacgttggaaactgacagaaccgcaacgttgaaggagccactcagc
cgccggtttctggagttaatgagctaagcacatacgtcagaaaccattattgcgcgtt
caaaagtgcctaaggtcactatcagctagcaaataatttcttgtcaaaaatgctccact
15 gacgttccataaattccctcggtatccaatttagagtctcatattcactctcaatccaa
ataatctgcaccggatctggatcggttcgcattgttgcattttgcacgcagg
ttctccggccgcttgggtggagaggctattcggtatgactgggcacaacagacaatcg
gctgctctgatgccgcgtgttccggctgtcagcgcagggcgccccggttctttgtc
aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcgtg
20 gctggccacgacggcggtcccttgcgcagctgtgcacgttgcactgaagcggaa
gggactggctgtattggcgaaatgcgcggcaggatctcctgtcatctcaccttgct
cctgccgagaaagtatccatcatggctgtcaatgcggcggctgcatacgttgc
ggctacctgcccattcgaccaccaagcgaacatcgcatcgagcgcacgtactcgga
tggaaagccggcttgcgtcaggatgtctggacgaagagcatcagggctcgccca
25 gccgaactgttcgcaggctcaaggcgcgcattgcggcgtatcggatgtatcgt
ccatggcgatgcctgcttgcgaatatcatgttggaaaatggccgctttctggattca
tcgactgtggccggctgggtgtggcggaccgcattcaggacatagcgttgcgttacggat
gatattgtgaagagacttggcggcgaatgggctgaccgcattcctcgtgttacggat
cgccgctccgattcgcagcgcattgccttgcattgcgttgcgttgcgttgc
30 cgggactctgggttcgaaatgaccgaccaagcgcacgcggccaaacctgc
tcgattccaccgcgccttctatgaaagggtggcttcggatcgtttccggacgc
ggctggatgtatcctccagcgcgggatctcatgtggagtttcgcggccacggatctc
tgcggAACAGGCGGTCAAGGTGCCGATATCATTACGACAGCAACGGCCGACAAGCACA
ACGCCACGATCCTGAGCGACAAATGATCGGGCCCAGTCCACATCAACGGCGTCGGC

ggcgactgcccaggcaagaccgagatgcaccgcgatatcttgcgtcgatatttt
cgtggagttcccgccacagacccggatgatccccgatcgtaaacattggcaataaa
gttcttaagattgaatcctgtgccggcttgcgatgattatcatataattctgttg
aattacgttaagcatgtataataacatgtaatgcacgttatattatgagatgggt
5 ttttatgatttagagtcccgcaattatacattaataacgcgatagaaaacaatatacg
gcgcaaactaggataaattatcgccgcggctgtcatctatgttactagatcgggcctcc
tgtcaatgctggcggcggctctgggggtgggtctggcggctctgaggggtggct
ctgaggggtggcggttctgaggggtggcggctctgagggaggcgggtccggtggtggct
ggttccggtgattttgattatgaaaagatggcaaacgctaataaggggctatgaccga
10 aaatgccatgaaaacgcgctacagtctgacgctaaaggcaaacttgattctgtcgcta
ctgattacggtgctgctatcgatggttcattggtgacgttccggcctgctaattgg
aatggtgctactgggatttgctggctctaattccaaatggctcaagtgggtgacgg
tgataattcaccttaatgaataattccgtcaatattacctccctccctcaatcg
ttgaatgtcgccctttgtcttggccaatacgcaaaccgcctctcccgccgttgg
15 ccgattcattaatgcagctggcacgacaggttcccactggaaagcggcagtgagcg
caacgcataatgtgagttagctcactcattggcacccaggcttacactttatgc
ttccggctcgtatgtgtggatttgtgagcggataacaattcacacaggaaacagc
tatgaccatgattacgccaagcttgcattgcctgcagccacagatggtagagaggct
acgcagcaggtctcatcaagacgatctaccgagcaataatctccagggaaatcaaatac
20 cttcccaagaaggtaaagatgcagtc当地actgc当地caagaacac
agagaaagatataattctcaagatcagaagtagactattccagtagtgacgattcaaggct
tgcttcacaaaccaaggcaagtaatagagattggagttctctaaaaggtagttccact
gaatcaaaggccatggagtcaaagattcaaatagaggacctaacagaactcgccgtaaa
gactggcgaacagttcatacagagtctcttacgactcaatgacaagaagaaaatcttcg
25 tcaacatggggagcacgacacacttgc当地actccaaaatataagatacagtc当地
gaagaccaaaggcaatttgagactttcaacaaaggtaatatccggaaacctcctcg
attccattgcccagctatctgtcactttattgtgaagatagtggaaaaggaaggtggct
cctacaaaatgccatcattgcgataaaggaaaggccatcggtgaagatgcctctggc
actgggtcccaaagatggacccccaccacgaggagcatcggtggaaaaagaagacgttcc
30 aaccacgtcttcaaagcaagtggattgtgatgtgatctccactgc当地aggatgc当地
cacaatcccactatccttcgcaagacccttc当地tatataaggaagttcatttcatttg
gagagaacacggggactctagaccagtgccagctcggtgc当地accgc当地
ttcattctcattctggcactgc当地aatcaatgtgtcataaagaacaatcc
tttgc当地
tgaagaaaccatacatgtaagcattactatggcttgc当地acccttgc当地tagatccatcgacaaca

aacagcttcttcagaggaaacttagagaagaagaagcaagttctcaatcttccggag
 gtctccatcaggaagaggagtgaacttgtgaaaagaggtgcaatcaaaacagggtata
 tagtcatcatcaactagagacaggataaacatgaatgcccacagatagatggcgaggtaa
 ggacctccttctgaactataacaataattgcggcaacgataggagggcaggtatgac
 5 agagaggagtattccttgatcatgtccctaattgaacatccatattgttgcattttttga
 acccatgccgagactcgatcacgaaagttgagtacaagaaaatggcaaattcagtgatc
 tgtgaccatgtcataagaccagccaagaatgaaagagtgtcaggatttcattctctgg
 atcgagtcccaccattggtagaaggatccccATCTACCCGTTCGCGTCGGCATCCGGT
 CAGTGGCAGTGAAGGGCGAACAGTTCTGATTAAACCACAAACCGTTCTACTTTACTGGC
 10 TTTGGTCGTCAAGATGCGGACTTGCCTGGCAAAGGATTGATAACGTGCTGATGGT
 GCACGACCACGCATTAATGGACTGGATTGGGCCACTCCTACCGTACCTCGCATTACC
 CTTACGCTGAAGAGATGCTCGACTGGCAGATGAACATGGCATCGTGGTATTGATGAA
 ACTGCTGCTGTCGGCTTTCGCTCTCTTAGGCATTGGTTGAAGCGGGCAACAAGCC
 GAAAGAACTGTACAGCGAAGAGGCAGTCAACGGGAAACTCAGCAAGCGCACTTACAGG
 15 CGATTAAAGAGCTGATAGCGCGTGACAAAAACCACCCAAGCGTGGTATGTGGAGTATT
 GCCAACGAACCGGATACCGTCCGCAAGGTGCACGGGAATATTGCGGCCACTGGCGGA
 AGCAACCGTAAACTCGACCCGACCGTCCGATCACCTCGTCAATGTAATGTTCTGCG
 ACGCTCACACCGATACCATCAGCGATCTCTTGATGTGCTGTGCCTGAACCGTTATTAC
 GGATGGTATGTCAAAGCGCGATTGGAAACGGCAGAGAAGGTACTGGAAAAAGAACT
 20 TCTGGCCTGGCAGGAGAAACTGTACACCGACATGTGGAGTGAAGAGTATCAGTGTGCAT
 GGCTGGATATGTATCACCGCGTCTTGATCGCGTCAGCGCCGTCGTCGGTGAACAGGTA
 TGGAAATTGCGCGATTTGCGACCTCGCAAGGCATATTGCGCGTTGGCGGTAAACAAGAA
 AGGGATCTTCACTCGCGACCGCAAACCGAAGTCGGCGGCTTCTGCTGCAAAACGCT
 GGACTGGCATGAACTTCGGTAAAAACCGCAGCAGGGAGGCAAACAATGAatcaacaac
 25 tctcctggcgaccatcgtcggtacagcctcggttatgcattaccgagctttctacca
atqgtggactcgatccagaaatcctgcacactcttcattttgttatgcgtatcg
atcgacatggtcacagatcactgattgcattttcttgcattcaactttcgatcg
agtctcggtatgggtcaacaaacaatatggatgttcatagggacatgtcaaa
ggaataactcctctgtcatacctgcccctctatcggtgcgcattattgttatgt
 30 tcagaaaggaggccttacctcgccatctatctgtggcattcatgtttatcctgtctc
tagtgcgtatgactatataccctgtttgattgcacctctttcaacaagttcactcct
cttcctgtatggagacctccggagaagattgagaaacttgcattctctaaagttcc
tctgaagaagctgtttgtcgatggatctacaaggtaagccatagtaatgcttaca
tgtatggttttcaagaacaaaaggattgttcttatgacacattgattcagcagtgc

cagaatgagaatgaaattgtggcggttattgcacacgagctgggacactgg gagctga
 attccccgatcgtaaacacattggcaataaagtttcttaagattgaatcctgtgcc
 ggtcttgcgtatcatataattctgttgaattacgttaagcatgtataattaa
 catgtaatgcgttatggatgggtttatgatttagatgggtttttatgatttagatgggttttt
 5 acatttaatacgcgatagaaaaacaaaatacgccgcaactaggataaattatcgcc
 gcgggtgtcatctatgttactagatcggttactggccgtcgatccacacgtcg
 actggaaaaccctggcgtaaccacttaatcgcccttgacgttcacatcccccttcgccc
 agctggcgtaatagcgaagaggcccgcaccgtccatccacatgggttttttt
 10 ccccgtaagctctaaatcggtttccctttaggttccgattttagtgccttacggca
 cctcgacccaaaaacttgatttgggtatgggtcacgttagtggccatcgccctgat
 agacggttttcgcccttgacgttggagtccacgtttaatagtggactctgttc
 caaactggacaacaactcaaccctatctcggttattttgatttataaggatttt
 gccgatttcggaaccaccatcaaacaggatttcgcctgctggggcaaaccagcgtgga
 15 ccgcttgcactctcagggccaggcggtgaagggaatcagctgtgcccgtct
 cactggtaaaaagaaaaaccaccccagtacataaaaacgtccgcaatgtgttattaag
 ttgtctaagcgtcaatttgcgttacaccacaatatacctgc
 SEQ ID NO:48 is the nucleic acid sequence of pBI121-HP-BnCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the 35S promoter. Sequence in bold is the BnCPP antisense sequence, bold and underlined sequence is the BnCPP sense fragment and upper case indicates the truncated GUS fragment.

SEQ ID NO:49

25 *gttacccgccaatatacctgtcaaacactgatagttaaactgaaggcggaaacga*
 caatctgatcatgagcggagaattaaggagtcacgttatgaccccccgcgtacgc
 ggacaagccgtttacgttggactgacagaaccgcaacgttgaaggagccactcagc
 cgccggtttctggagttaatgagctaagcacatacgtcagaaaccattattgcgcgtt
 caaaaagtgcctaaggtcactatcagtagcaaatttcttgcgtaaaaatgctccact
 30 gacgttccataaattccctcggtatccaatttagtctcatattcactctcaatccaa
 ataatctgcaccggatctggatcggttcgcattgttgcacatggattgcacgcagg
 ttctccggccgttgggtggagaggctattcggtatgactggcacaacagacaatcg
 gctgctctgtatgcgcgtgttccggctgtcagcgcaggggcgcccggttctttgtc
 aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcg

gctggccacgacgggcgttccttgcgcagctgtgctcgacgttgtcactgaagcggaa
gggactggctgtattggcgaaagtgcgcggggcaggatctcctgtcatctcacctgct
cctgccgagaaagtatccatcatggctgtatgcatacgcttgcacgttatcc
ggctacctgcccattcgaccaccaagcgaaacatcgcatcgagcgagcacgtactcgga
5 tggaaagccggctttgtcgatcaggatgtatggacgaagagcatcaggggctcgccca
gccgaactgttcgcaggctcaaggcgcatgcccacggcgatgtatctcgatgc
ccatggcgatgcctgttgcgaatatcatggtaaaaatggccgctttctggattca
tcgactgtggccggctgggtgtggcgaccgctatcaggacatagcgttgctaccgt
gatattgtgaagagacttggcgcaatggctgaccgcttcgtgctttacggat
10 cgccgctcccgattcgacgcacgcattatcgcccttcgtgacgagttttctgag
cgggactctgggttcgaaatgaccgaccaagcgacgccccaacctgcacgcgatt
tcgattccaccgcgccttctatgaaagggtggcttcggaatcgccccggacgcc
ggctggatgtatccctcagcgccccatctcatgctggagttttcgccccacggatctc
tgcggAACAGGCGGTcgaagggtgccatattacgacagcaacggccacaaggaca
15 acgccacgatcctgagcgacaaatatgtatcggcccccgtccacatcaacggcgcc
ggcgactgcccaggcaagacccgagatgcaccgcgatatcttgcgtcgatcggatattt
cgtggagttcccgccacagacccggatgtatccccgatcgttcaaacattggcaataaa
gttcttaagattgaatcctgttgcggcttgcgtatgattatcatataattctgtt
aattacgttaagcatgtataattacatgtaatgcacgttattatgagatgggt
20 tttatgatttagagtccgcattatacattaaatacgcgatagaaaacaaaatatacg
gcgcaactaggataattatcgcgccggcttgcgtatctatgttacttagatcgccctcc
tgtcaatgctggcgccggcttgcgtatctgggtggatcggtggcgctctgagggtggct
ctgagggtggcggtctgagggtggcgctctgagggaggcggtccgggtggctct
ggttccggtgatttgattatgaaaagatggcaaacgctaataagggggctatgaccga
25 aaatgccatgaaaacgcgtacagtctgacgctaaaggcaaacttgcgtatcgct
ctgattacggtgctgtatcgatggttcattggtgacgcttcggccttgcataatgg
aatggtgctactggtgatttgctggcttaattccaaatggctcaagtgggtacgg
tgataattcaccttaatgaataattccgtcaatattacctccctccctcaatcg
ttgaatgtcgccctttgtcttggccaatacgcaaaccgcctctcccgccgttgg
30 ccgattcattaatgcagctggcacgacaggtttccgactggaaagcggcagtgagcg
caacgcattaaatgtgagtttagctactcattaggcacccaggcttacacttgc
ttccggctcgatgttgcgttggaaattgtgagcggtataacaatttcacacaggaaacagc
tatgaccatgattacgccaagcttgcgtgcagcccacagatggtagagaggctt
acgcagcaggtctcatcaagacgtacccgagcaataatctccagggaaatcaaatac

cttcccaagaaggtaaagatgcagtcaaaagattcaggactaactgcatcaagaacac
 agagaaaagatataatttctcaagatcagaagtactattccagtatggacgattcaaggct
 tgcttcacaaaccaaggcaagtaatagagattggagttctctaaaaaggtagttcccact
 gaatcaaaggccatggagttcaagattcaaatacgaggacctaacaagaactcgccgtaaa
 5 gactggcgaacagttcatacagacttcttacgactcaatgacaagaagaaaatctcg
 tcaacatggtgagcacgacacacttgtctactccaaaaatataagatcaagatacgactca
 gaagaccaaaggcaattgagactttcaacaaaggtaatatccggaaacctctcg
 attccattgccagctatctgtcactttattgtgaagatagtggaaaaggaaggtagtggct
 cctacaaatgccatcattgcataaaggaaaggccatcgtaagatgcctctgcccac
 10 agtggtcccaaagatggacccccaccacgaggagcatcgtaagggatgac
 aaccacgtcttcaagcaagtggattgtgatgttatctccactgacgtaaggatgac
 cacaatcccactatcctcgcaagacccttccttatataaggaaagttcatttgcatttgc
 gagagaacacggggactctagaggatcc
 15 ttaatctgtcttgcatttcattcaatggctcgaaagcctcttacaagaggagggtgtgagtagtgcataatgggt
 ctgtgttcatcgctgataagttcttctgttagcttcaactaggcagacgtagatcc
 tttgcataaccaagattcactgcaaaagcatcagcctgaaactcaaacgctcgactaac
 aagggttaggtcaaagcttacttaggtgttatcactgtgtgaaatataaggaaatgt
 tcaaaccaatgagaactgggtgttatcaaaacaaaactcctgaagagatcagtgag
 tttctgacaagagtgtatcctccaatttgcataaaggcaaggatttgcataatgaa
 20 cgagtatgttagtgcattcagttccagtgcccagctcgatgtgcaataaccgcacaa
 tttcatttcattctggcactgctgatcaatgtgtcataaagaacaatcctttgttc
 ttgaagaaaccatacatgttaagcattactatggcttgaccttgcatttgcataac
 aaacagcttccatcaggaagaggagtgaacttgcatttgcataatcctcccgaa
 ggtctccatcaggaagaggagtgaacttgcatttgcataatcctcccgaa
 25 atagtcatcatcacttagagacaggataaacatgatgtgcggccacagatgatggc
 aggacccatccttctgactataacaataattgcggcaacgataggaggggcaggat
 cagagaggagtattccttgcatttgcataatgcggccatattgtttgttgcatttgc
 aacccatgcccagactcgatcagaaatggcatttgcatttgcataatcctcccgaa
 ctgtgaccatgtcataagaccagccaaatggcatttgcatttgcataatcctcccgaa
 30 gatcgagttcccaccattggtagaaagccgcacatgttccatattgtttgcatttgc
 ccaaagaacagaatcgccatcggatgttgcatttgcataatcctcccgaaatggc
 atggctttgtcaagactgttgcatttgcataatcctcccgaaatggcatttgcatttgc
 caaccaaaatgtttggagagttggagttgcatttgcataatcctcccgaaatggc
 tacgtctcaaaaacgtacatcactatcataaaaccaacgcacgggtttccatgaaaggaat

cgccatccctgcgaattccccgatcgttcaaacattggcaataaagtttcttaagat
tgaatcctgttgcggcttgcgttatcatataatttctgttgaattacgttaag
catgtataattaaacatgtaatgcgttattatgagatgggtttatgattag
agtcccgcattatacatttataacgcgtatagaaaacaaaatatagcgcgcactagg
5 ataaaattatcgcgcggtgtcatctatgttacttagatcggttactcggttcactggccgtcgat
tttacaacgtcgtgactggaaaaccctggcgttacccacttaatcgccgtcagcac
atcccccttcgcagctggcgtaatagcgaagaggccgcaccgtcgcccttccaa
cagttgcgcagcctgaatggcgccgctcccttcgccttctcccttctcgcca
cgttcgccggcttccccgtcaagctctaattcggggctccctttagggttccgatt
10 agtgcttacggcacctcgacccaaaaacttgattgggtgatggttcacgttagtgg
gccatcgccctgatagacggtttcgccttgcgtggagtccacgttcttaata
gtggactttgttccaaactggaaacaacactcaaccstatctcggttattctttgat
ttataagggatttgcgatttcggaaaccaccatcaaacaggatttcgcctgctggg
caaaccagcgtggaccgcttgcactcaactcttcaggccaggcggtgaaggcaatca
15 gctgttgcgcgtctcaactggtaaaaagaaaaaccaccccgatcacattaaaacgtccgc
aatgtgttattaaagtgtctaagcgtcaattgttacaccacaatatacctgc

SEQ ID NO:49 is the nucleic acid sequence of pBI121-antisense-BnCPP.

Italicized sequences are the right and left border repeats. Underlined sequence is the 35S promoter. Sequence in bold is the BnCPP antisense sequence.

SEQ ID NO:50

gtttaccgc caata tata cctgtcaaac actgat gat ttaact gaaggc ggaa acga
caat ctgat catg agcggaga atta agg gatc acg ttat gacccccccgcatgacgcg
25 ggacaaggcgtttacg tttt ggaactg acaga accgca acgttgaaggagccactcagc
cgcg gggttctggat ttaatg agct aagcacatacgtcaga aaccattattgcgcgtt
caaa agtcgcctaagg tcaactatc agct agcaatatttctgtcaaaa atgctccact
gacgttccataaattcccctcggtatccaatttagagtctcatattcactctcaatccaa
ataatctgcaccggatctggatcg tttcgcatgattgaacaagatggattgcacgcagg
30 ttctccggccgcttgggtggagaggctattcggtatgactggcacaacagacaatcg
gctgctctgatgccg ccgtgttccggctgtcagcgcaggggcggccggttctttgtc
aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcgtg
gctggccacgacgggcgttcctgcgcagctgtcgtcagctgtcaactgaagcggaa
gggactggctgctattggcgaagtgcggggcagatctcctgtcatctcaccttgc

cctgccgagaaagtatccatcatggctgatgcaatgcggcggctgcatacgcttgc
 ggctacctgcccattcgaccaccaaggcgaaacatcgcatcgagcgacgtactcgga
 tggaaagccggcttgcgatcaggatgtggacgaagagcatcagggctcgccca
 gccaactgttcgcaggctcaaggcgcatgcccacggcgatgtctcgctgtgac
 5 ccatggcgatgcctgcttgcgaatatcatggtaaaaatggccgctttctggattca
 tcgactgtggccggctgggtgtggcggaccgctatcaggacatagcgttgctacccgt
 gatattgctgaagagcttggcggcgaatggctgaccgcctcgtgccttacggtat
 cgccgctcccgattcgcagcgcatgcctctatgccttgcgtacgagttctctgag
 cggactctggggttcgaaatgaccgaccaaggcgacgccccacctgcacgagatt
 10 tcgattccaccgcgccttatgaaaaggttggcttcggaatcgccccggacgcc
 ggctggatgatcctccagcgcgggatctcatgctggagttctcgcccacgggatctc
 tgcggAACAGGCGGTcgaagggtgccatATCATTACGACAGCAACGGCCGACAAGCACA
 acggccacgatcctgagcgacaatATGATCgggcccgggtccacatcaacggcgtcggc
 ggcactgcccaggcaagaccgagatgcaccgcgatATCTGCTGCGTGGATA
 15 cgtggagttcccgcacagacccggatgatccccgatcgttcaaACATTGGCAATAAA
 gtttcttaagattgaatcctgttgcggcttgcgtatgattatcatataattctgtt
 aattacgttaagcatgtataattacatgtaatgcgtacgttattatgagatgggt
 tttatgatttagagtccgcattatacattaaatcgcgatagaaaacaaaatata
 ggcgcacactaggataattatcgcgcgcggtgtcatctatgttactagatcggc
 20 tgtcaatgctggcggcggctctgggtggttctggggcggctctggggggct
 ctgagggtggcggtctgagggtggcggctctgaggggaggcggtccgggtggct
 ggttccgggtgatttgattatgaaaagatggcaaacgctaataaggggctatgacc
 aaatgcgcattaaacgcgcgtacagtctgacgctaaaggcaacttgattctgc
 ctgattacggtgctgtatcgatggttcattggtacggttccggccttgc
 25 taatggtgctactggtgatttgctggctctaattccaaatggctcaagtcgg
 tgataattcaccttaatgaataattccgtcaatattacctccctccctcaatcg
 ttgaatgtcgccctttgtcttggcccaatacgcaaacgcctctcccgcg
 ccgattcatTAATGCAGCTGGCACGACAGGTTCCCAGTGAAAGCGGG
 CAGTGA
 caacgcattaaatgtgagttagctcactcattaggcacccaggcttacactt
 30 ttccggctcgatgtgtggattgtgagcgataacaattcacacaggaaac
 agc
 tatgaccatgattacgccaagcttgcattgcgcaggagccatagatgc
aaactgaaatttctgcaagaatctcaaacacggagatctcaaagttgaaagaaaattt
atttctcgactcaaaacaaacttacgaaatttaggtagaacttatatacattatattg
taatttttgcataaaaaatgttttattattatagaatttactggtaaattaaa

aatgaatagaaaaggtaattaaaggagagaggagaaacatttcttattttt
 catatttcaggataaatttattgtaaaagttacaagattccatttgactagtgtaaa
 ttaggaatattctctagtaagatcatttcatctacttctttatcttaccagta
 gagaaataaacaatatttagctcattgtaaataacaattaaatttccttgcacatc
 5 attcaatttaatttacgtataaaataaaagatcatacctattagaacgattaaggag
 aaataacaattcgaatgagaaggatgtgccgttgtataataaacagccacacgacgta
 aacgtaaaatgaccacatgatggccaatagacatggaccgactactaataatagtaag
 ttacattttaggatgaaataatcataccgacatcagtttggaaagaaaaggaaaa
 aaagaaaaataaataaaagatatactaccgacatgagttccaaaagcaaaaaaaaaaag
 10 atcaagccgacacagacacgcgtagagagcaaaatgactttgacgtcacaccacgaaaa
 cagacgcttcatacgtgtcccttatctctcagtcctctataaacttagtgagacc
 ctccctgtttactcacaatatgcaaactagaaaacaatcatcaggaataaagggtt
 tgattacttctattggaaaggactctagaggatccatggcgattccttcatggaaacc
 gtcgttggtttatgatagtgtacgtttgagacgtattggatctgaggcaaca
 15 tactgctctcaagcttcccactctcccaaagactttgttggagtcatggcaagaga
 agtttgagaaatctcgagcttacagtcttgacaaaagccatttcactttgttcatgag
 tttgttactatacttatggactctgcgattctgttcttggatctgccttggtttg
 gaagatatctggcggcttctaccaatggggactcgatccagagaatgaaatcctgc
 acactcttcattcttggctggcttatgacatggcacagatcactgatttgcattt
 20 tctttgtactcaacttctgtatcgactctcgcatgggttcaacaaacaaacaatatg
 gatgttcattaggacatgatcaaaggataactcctctgtcatacctgccttccta
 tcgttgcgcattattgttagttcagaaaggaggtccttacctgcgcattctatctg
 tggcattcatgtttatcctgtcttagtgtatgactatataccctgtttgattgc
 acctctttcaacaagttcactccttgcgtatggagacctccggagaagattgaga
 25 aacttgcttctctaaagttcctctgaagaagctgtttgtcgatggatctaca
 aggtcaagccatagtaatgcttacatgtatgggtttctcaagaacaaaaggattgtct
 ttatgacacattgattcagcagtgcagaatgagaatgaaattgtggcggttattgcac
 acgagctggacactggaagctgaatcacactacatactcgatcgatgttcaaatc
 ctgccttcttgcataattggaggataactctgtcagaaactccactgatctttcag
 30 gagtttggtttgcatacacaaccagttctcattgggttgcataattcagcacactg
 taataccacttcaacacacccatgtaaagcttgcaccaacccattgttagtcagcgttgc
 tttcaggctgtatgtttgcagtgaatcttgggtatgcaaaaggatctacgtcctgcct
 agtgaagctacaggaagagaacttatcagcgatgaacacagacccattgtactcagctt
 atcactactcacaccctccttgcatacggatcgatggagaagacaag

aagacagataacccctcgaaattccccgatcg~~t~~caaacattggcaataaagtttct
taagattgaatcctgttgcgcgtctgcgtattatcatataattctgttgaattac
gttaaggcatgtataattaacatgtaatgc~~t~~gacgttatttatgagatgggttttat
gattagagtcccgcaattatacattaatacgc~~t~~gatagaaaacaaaatatacgcgcaaa

5 actaggataaattatcgcgcggtgtcatctatgttacttagatcg~~gg~~gattcactggc
cg~~t~~cg~~t~~ttacaacgtcg~~t~~gactggaaaacc~~t~~ggcgttaccc~~a~~c~~t~~taatcgcc~~t~~tg
cagcacatcccccttcg~~c~~cagctggcgtaatagc~~g~~aagaggcccgcaccgatcgccct
tcccaacagttgcgcagc~~t~~g~~a~~atggcgccg~~c~~ctc~~t~~tcg~~c~~ttcc~~c~~ttc~~c~~ttc
tcgcccacg~~t~~tcgcccgg~~c~~ttccccgtcaagctcta~~a~~atcg~~gg~~gg~~g~~ctcc~~c~~tttaggg~~t~~c

10 cgatttagtg~~c~~ttacggcac~~c~~tcgac~~cc~~aaaaacttgattgggtgatgg~~t~~cacg
tagtggccatcgcc~~c~~tgatagacgg~~t~~ttcgcc~~c~~ttgacgttg~~g~~ag~~t~~ccac~~g~~ttc
ttaatagtg~~g~~actcttg~~t~~ccaaactgg~~a~~acaacactcaacc~~c~~tatctcg~~g~~g~~c~~tattct
tttgatttataagg~~g~~atttgccgattcgg~~a~~accaccatcaa~~a~~acaggatttcg~~c~~ctg
ctggggcaaaccagcgtggaccg~~c~~ttgctgcaactctcagggccaggcgg~~t~~gaagg

15 caatcagctgttgc~~c~~ctcactgg~~t~~aaaagaaaaaccac~~cc~~c~~ag~~tacatta~~aa~~ac
gtccgc~~a~~atgtgttattaagttgtctaagcgtcaattt~~g~~ttacaccacaatatacct
gcca

SEQ ID NO:50 is the nucleic acid sequence of pRD29A-BnCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the RD29A promoter. Sequence in bold is the BnCPP sense sequence.

SEQ ID NO:51

25 gtttacccgccaatatacctgtcaaacactgatagttaaactgaaggcggaaacg
caatctgatcatgagcggagaattaagggagtacgttatgacccccgcccgtacgcg
ggacaagccgtttacgttggactgacagaaccgcaacgttgaaggagccactcagc
cgccgggttctggagttaatgagctaagcacatacgtcagaaaccattattgcgcgtt
caaaaagtgcgcctaaggtaactatcagctagcaaataatttctgtcaaaaatgctccact
gacgttccataaattccctcggtatccaatttagagtctcatattcactctcaatccaa
30 ataatctgcacccggatctggatcggttcgcattgttgcacgcagg
ttctccggccgcttgggtggagaggctattcggtatgactggcacaacagacaatcg
gctgctctgtatgccgcgtgttccggctgtcagcgcaggggcggccggttcttttgtc
aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcgtg
gctggccacgacggcgttccttgcgcagctgtgcacgttgcactgaagcggaa

gggactggctgctattggcgaagtgccggggcaggatctcctgtcatctcaccttgct
cctgccgagaaaagtatccatcatggctatgcataatgcggcggctgcatacgcttgc
ggctacctgcccattcgaccaccaagcgaaacatcgcatcgagcgagcacgtactcg
tggaaagccggctttgtcgatcaggatgtctggacgaagagcatcagggctcgcc
5 gccgaactgttcgccaggctcaaggcgcatgcccacggcgatgtatctcgatcgac
ccatggcgatgcctgcttgcgaatatcatggtaaaatggccgttttctggattca
tcgactgtggccggctgggtgtggcgaccgctatcaggacatagcgttgctacccgt
gatattgctgaagagcttggcgccgaatggctgaccgcttcctcgatcgagttttcg
10 cggactctgggttcgaaatgaccgaccaagcgacgccccaacctgcacgatc
tcgattccaccgcgccttatgaaagggtggcttcggatcgatcgatcgatcgatcg
ggctggatgatcctccagcgccggatctcatgctggagttctcgcccacggatctc
tgccgaacaggcggtcgaaagggtccgatattacacagcaacggccacaaggaca
acgcccacgatcctgagcgacaatatgatcggcccggtccacatcaacggcgatcg
15 ggcactgcccaggcaagaccgagatgcaccgcgatatttcgtcgatcgatattt
cgtggagttccgcacagaccggatgatccccgatcgatcgatcgatcgatcgatcg
gtttcttaagattgaatcctgttgcggcttgcgatgattatcatataattctgtt
aattacgttaagcatgtataattacatgtaatgcacgttattatgagatgggt
tttatgatttagagtccgcattatacatcgatcgatcgatcgatcgatcgatcg
20 gcgcaaactaggataaattatcgccgcggtgtcatctatgttactagatcgcc
tgtcaatgctggcgccggctctgggtggttctgggtggcgctctgggtggct
ctgagggtggcggtctgagggtggcgctctgggtggcggtccgggtggct
gggtccggtgatttgattatgaaaagatggcaaacgctaataaggggctatgacc
aaatgccatgaaaacgcgtacagtctgacgctaaaggcaaacttgcgtcgatcg
25 ctgattacggtgctgctatcgatggttcatggtgacgttccggcctgctaatt
aatggtgctactggtgatttgctggctctaattccaaatggctcaagtggatcg
tgataattcaccttaatgaataattccgtcaatattacctccctccctcaatcg
ttgaatgtcgccctttgtcttggcccaatacgcaaaccgcctctcccgcgatgg
ccgattcattaatgcagctggcacgacaggttccgactggaaagcgccgatcg
30 caacgcaattaatgtgagtttagctcactcattaggcacccaggcttacactt
ttccggctcgatgttgcggataacaattcacacaggaaacagc
tatgaccatgattacgccaagcttgcgtcgatcgatcgatcgatcgatcg
aaactgaaatttctgcaagaatctcaaacacggagatctcaaagtggaaagaaatt
atttcgactcaaaacaaacttacgaaatttaggtagaacttatacattatattg

taattttttaacaaaatgttttattattatagaattttactggttaaattaaa
 aatgaatagaaaaggtaatataagaggagagaggtaaacatttcttatTTT
 catatttcaggataaattattgtaaaagttaacatcattccatTTGactagtgtaaa
 tgaggaatattctcttagtaagatcattttcatctacttctttatTTTaccagta
 5 gaggaataaaacaatatttagctcTTGtaaatacaaaattaatttcTTGacatc
 attcaatttaattttacgtataaaaataaaagatcataacctattagaacgattaaggag
 aaatacaattcgaatgagaaggatgtGCCGTTGTTataataaacagccacacgacgta
 aacgtaaaatgaccacatgatggccaatagacatggaccgactactaataatagtaag
 ttacattttaggatgaaataatataccgacatcagTTGaaagaaaaggaaaa
 10 aaagaaaaataaaataaaagatatactaccgacatgagTTCCAAAAGcaaaaaaaag
 atcaagccgacacagacacgcgttagagagcaaaatgacttgcgtcacaccacgaaaa
 cagacgcttcatacgtgtccCTTatctctcagtctctataaacttagtgagacc
 ctccctgtttactcacaatatgcaaactagaaaacaatcatcaggaataaagggtt
 tgattacttctattgaaaggactctaga**ccagtgtcccagctcgTGTGcaataaccgc**
 15 **cacaatttcattctcattctggactgctgaatcaatgtgtcataaagaacaatcctt**
tgttcttgaagaaaccatacatgttaagcattactatggcttgcacTTGtagatccatcg
acaacaaacagcttccatcagaggaaacttagagaagaagcaagttctcaatcttc
ccggaggctccatcaggaagaggagtgaacttggaaaagaggtgcaatcaaaacag
ggtatatagtcatcatcactagagacaggataaacatgaatgcccacagatagatggcg
 20 **aggtaaggacctcTTCTGAactataacaataattgcggcaacgataggaggggcagg**
tatgacagagaggagtattcTTGatcatgtccctaattgtggatccatattgtttgtt
tgttgaaccatgcccagactcgatcacgaaagttagtgcataaaagaaaatggcaatca
gtgatctgtgaccatgtcataagaccagccaagaatgaaagagtgtgcaggatttcatt
ctctggatcgagtcaccattggtagaaaggatccccATCTACCGCTCGCGTCGGCA
 25 **TCCGGTCAGTGGCAGTGAAGGGCGAACAGTTCTGATTAACCACAAACCGTTCTACTTT**
ACTGGCTTGGTCGTATGAAGATGCCTGGACTTGCCTGGCAAAGGATTGATAACGTGCT
GATGGTGCACGACCACGCATTAATGGACTGGATTGGGGCAACTCCTACCGTACCTCGC
ATTACCCCTACGCTGAAGAGATGCTCGACTGGCAGATGAACATGGCATCGTGGTGATT
GATGAAACTGCTGCTGGCTTTCGCTCTTTAGGCATTGGTTCGAAGCGGGCAA
 30 **CAAGCCGAAAGAACTGTACAGCGAAGAGGCAGTCAACGGGAAACTCAGCAAGCGCACT**
TACAGGCGATTAAAGAGCTGATAGCGCGTGACAAAAACCACCCAAGCGTGGTGATGTGG
AGTATTGCCAACGAAACCGGATACCGTCCGCAAGGTGCACGGGAATATTCGCGCCACT
GGCGGAAGCAACGCGTAAACTCGACCCGACCGTCCGATCACCTGCGTCAATGTAATGT
TCTGCGACGCTCACACCGATACCATCAGCGATCTTTGATGTGCTGTGCCTGAACCGT

TATTACGGATGGTATGTCCAAAGCGGCGATTGGAAACGGCAGAGAAGGTACTGGAAAA
 AGAACTTCTGGCCTGGCAGGAGAAACTGTACACCGACATGTGGAGTGAAGAGTATCAGT
 GTGCATGGCTGGATATGTATCACCGCGTCTTGATCGCGTCAGCGCCGTCGTCGGTGAA
 CAGGTATGGAATTGCCGATTTGCACCTCGCAAGGCATATTGCGCGTTGGCGGTAA
 5 CAAGAAAGGGATCTTCACTCGCGACCGCAAACCGAAGTCGGCGCTTCTGCTGCAA
 AACGCTGGACTGGCATGAACCTCGGTGAAAACCGCAGCAGGGAGGCAAACAATGAATC
 AACAACTCTCCTGGCGACCATCGTCGGCTACAGCCTCGGAATTGCTACCGAGCTtc
ctaccaatggtggactcgatccagagaatgaaatcctgcacactttcattttggc
tggtcttatgacatggtcacagatcactgattgccatttcttgtactcaactttcg
 10 tgatcgagtctcgcatgggtcaacaaacaaatatggatgttcatagggacatg
atcaaaggaatactcctctgtcatacctgcccctctatcgttgcccaattattgt
tatagttcagaaaggaggtccttacctcgccatctatctgtggcattcatgtttatcc
tgtctctagtgtatgactatataccctgtttgattgcacctctttcaacaatgttcc
actcctttcctgtatggagacacctccggagaagattgagaaacttgcttctctaaa
 15 gtttcctctgaagaagctgtttgtcgatggatctacaaggtaagccatagtaatg
cttacatgtatggttcttcaagaacaaaggattgttctttagacacattgattcag
cagtgccagaatgagaatgaaattgtggcggttattgcacacgagctggacactggga
gctcgaattccccatcgtaaaacattggcaataaagttcttaagattgaatcct
gttgcggcttgcgtatcatataattctgttgaattacgttaagcatgtaat
 20 aattaacatgtatgcgttattatgagatgggtttatgattagagtccgc
aattatacattaatacgcgatagaaaacaaaatatagcgcgcaactaggataaatta
tcgcgcgcgtgtcatctatgttactagatcggaattcactggccgtcgccccat
gtcgtgactggaaaaccctggcgtaacccaacttaatcgccctgcagcacatccccct
ttcgccagctggcgtaatagcgaagaggcccgcaccgatcgccctccaacagttgcg
 25 cagcctgaatggcgccccgtcccttcgtttcccttccttcgtccacgtcgcc
ggcttccccgtcaagctctaaatcgaaaaacttgattgggtgatggttcacgttagtggccatcgc
cctgatagacggttttcgccttgcgttgcacgtggagtcacgtttaatagtggactc
ttgttccaaactggaacaacactcaaccatatctcggctatttttatttataagg
 30 gattttgccgatttcgaaaccaccatcaaacacaggatttcgcctgctggcaaaaccag
cgtggaccgcttgctgcaactctctcaggccaggcggtaagggcaatcagctgtgc
ccgtctcactggtaaaagaaaaaccaccccagtagcattaaaacgtccgcaatgtgtt
attaagttgtctaagcgtcaatttgcatttacaccacaatatacctgcca

SEQ ID NO:51 is the nucleic acid sequence of pRD29A-HP-BnCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the RD29A promoter. Sequence in bold is the BnCPP antisense sequence, bold and underlined sequence is BnCPP sense fragment and the upper case sequence represents the truncated GUS fragment.

SEQ ID NO:52

g~~tttacccggccaatatacctgtcaa~~acactgatagttaaactgaaggcggaaacga
 caatctgatcatgagcggagaattaaggagtcacgttatgacccccccggccatgacgcg
 ggacaaggcgtttacgttggaactgacagaaccgcaacgttgaaggagccactcagc
 cgcggtttctggagttaatgagcataacgtcagaaaccattattgcgcgtt
 caaaagtgcctaaggtcaactatcagctagcaaatatttcttgcaaaaatgctccact
 gacgttccataaattcccctcggtatccaatttagtctcatattcactctcaatccaa
 ataatctgcaccggatctggatcgttcgcatgattgaacaagatggattgcacgcagg
 ttctccggccgcttgggtggagaggctattcggtatgactgggcacaacagacaatcg
 gctgctctgatgccccgtgttccggctgtcagcgcaggggcgcccggttctttgtc
 aagaccgacctgtccggtgccctgaatgaactgcaggacgaggcagcgcggctatcg
 gctggccacgacgggcgttcttgccgacgtgtcactgaagcggaa
 gggactggctgctattggcgaagtgcgcggcaggatctcctgtcatctcaccttgc
 cctgccgagaaagtatccatcatggctgatgcaatgcggcggctgcatacgcttgc
 atccggctacctgcccattcgaccaccaagcgaaacatcgcatcgagcgcacgtactcg
 gatggaaagccggcttgcatcaggatgtctggacgaagagcatcaggggctcgccgcca
 gccgaactgttcgccaggctcaaggcgccatgcccacggcgatgtcgtcgtac
 ccatggcgatgcctgctgtccgaatatcatggttggaaaatggccgtttctggattca
 tcgactgtggccggctgggtgtggcggaccgctatcaggacatagcgttgctaccccgt
 gatattgctgaagagcttggcggcgaatgggtgaccgcttcctgtgctttacggat
 cgcccgtcccgattcgcagcgcatcgccttctatcgccttcttgacgagttcttctgag
 cgggactctgggttcgaaatgaccgaccaagcgacgcccacctgcccatccggaggatt
 tcgattccaccgccccttctatgaaaggttggcttcggaatcgttccggacgcc
 ggctggatgatcctccagcgcgggatctcatgctggagttctcgccacgggatctc
 tgcggAACAGGCGGTcgaaagggtgccgatatcattacgcacagcaacggccgacaagcaca
 acgcccacgatccttgagcgacaatatgatcggcccggtccacatcaacggcgtcggc
 ggcgactgcccaggcaagaccgagatgcacccgcgatatcttgctgcgttcgggatattt
 cgtggagttcccgcacagaccccggatgtccccgatcgttcaaacattggcaataaa

gtttcttaagattgaatcctgttgcggcttgcgatgattatcatataatttctgttg
aattacgttaagcatgtataataattaacatgtaatgcgtacgttatggatgggt
tttatgatttagagtcccgcaattatacattaatacgcatagaaaacaaaatatacg
gccaactaggataaattatcgccgcgggtgtcatctatgttactagatcgggcctcc
5 tgcgtcaatgcgtggcgccggctctgggtgggtctgggtggcggtctgagggtggct
ctgagggtggcggtctgagggtggcggtctgagggaggcggtccgggtggct
gggtccgggtgatttgattatgaaaagatggcaaacgctaataaggggctatgaccga
aaatgccatgaaaacgcgctacagtctgacgctaaaggcaaacttgattctgtcgcta
ctgattacggtgctgctatcgatggttcattggtgacggttcggccttgctaatttgt
10 aatggtgctactggtatttgctggctctaattccaaatggctcaagtcggtgacgg
tgataattcaccttaatgaataattccgtcaatattacccctccctcaatcggt
ttgaatgtcgccctttgtcttggccaaatacgcaaaccgcctctccccgcgttgg
ccgattcataatgcagctggcacgacaggttcccactggaaagcggcagtgagcg
caacgcaattaatgtgagttagctcactcattaggcaccccaggcttacactttatgc
15 ttccggctcgtatgtgtgtggatttgtgagcggtacaattcacacaggaaacagc
tatgaccatgattacgccaagcttgcattgcgcgtcaggagccatagatgcaattcaatc
aaactgaaatttctgcaagaatctcaaacacggagatctcaaagttgaaagaaaattt
atttctcgactcaaaacaaactacgaaatttaggtagaacttatatacattatattg
taattttttaacaaaatgttttattattatagaattttactggttaaattaaa
20 aatgaatagaaaaggtgaattaagaggagagaggtaaacatttcttattttt
catatttcaggataaatttattgtaaaagtttacaagattccatttgactgtgaaa
tgaggaatttctcttagtaagatcatttcatctacttctttatcttaccagta
gaggaataaacaatatttagtccttgtaaatacaaaattaatttccttgcacatc
attcaatttaatttacgtataaaataaaagatcatacctattagaacgattaaggag
25 aaatacaattcgaatgagaaggatgtgccgttgttataataaaacagccacacgacgta
aacgtaaaatgaccacatgatggccaatagacatggaccgactactaataatagtaag
ttacatttttaggatggaataaatatcataccgacatcagttgaaagaaaaggaaaa
aaagaaaaataaataaaagatatactaccgacatgagttccaaaaagcaaaaaaaaaag
atcaagccgacacagacacgcgtagagagcaaatgacttgcgtcacaccacgaaaa
30 cagacgcttcatacgtgtcccttattctctcagtctctataaaacttagtgagacc
ctcctctgtttactcacaatatgcaaactagaaaacaatcatcaggaataaagggtt
tgattacttctattggaaaggactctagaggatcc**ttaatctgtttctgtttctcc**
atcaatggctcgaagcctctacaagaggagggtgtgagtagtgataagctgagtaca
atgggtctgtgttcatcgctgataagttcttcctgttagcttcactagggcaggacgt

agatcctttgcataaccaagattcactgcaaaagcatcagcctgaaactcaaacgctcg
 actaacaagggttgggtcaaagcttacttaggtgttgaagtggattacagtgtgctgaa
 atatgatcaaaccaatgagaactggtgttatcaaaacccaaaactcctgaagagatca
 gtggagtttctgacaagagtgtatcctccaaattgcaagaaggcaaggattgaacagc
 5 aatgaacgagttatgttagtgtgattcagcttccagtgtcccagctcgtgtgcaataaccg
 ccacaatttcatttcattctggactgctgaatcaatgtgtcataaagaacaatcctt
 ttgttcttgaagaaaccatacatgtaagcattactatggcttgacctttagatccatc
 gacaacaaacagcttccatcaggaagaggagtgaacttggaaaagaggtgcaatcaaaaca
 10 gggtatatagtcatcatcactagagacaggataaacatgaatgcccacagatagatggc
 gaggttaaggaccccttctgaactataacaataattggcaacgataggaggggcag
 gtatgacagagaggagtattccttgcattatgtccctaatgaacatccatattgttgc
 ttgttgaacccatgccgagactcgatcacgaaagttgagttacaaagaaaatggcaaatc
 agtgatctgtgaccatgtcataagaccagccaagaatgaaagagtgtgcaggattcat
 15 tctctggatcgagtccaccattggtagaaagcccccagatattccaaaaccaaggc
 aagatccccaaagaacagaatcgagactccataagtatagtaacaaactcatgaacaaa
 gtgaaaatggctttgtcaagactgtaagctcgagattctcaaaacttcttggctaa
 tgactccaaccaaagtcttggagagtggaaagcttgcagagcagtatgtgcctcaga
 tccaaatacgtctaaaaacgtacatcactatcataaaaccaacgacgggttccatgaa
 20 ~~aggaatcgccat~~ccccctcgaaatttccccgatcggttcaaacattggcaataaagttct
 taagattgaatcctgttgcggcttgcgtatgattatcatataatttctgttgaattac
 gttaaggcatgtataattaacatgtaatgcgtatgcgttattatgagatgggtttat
 gattagagtcccgcaattatacatgtaatgcgtatgaaacaaaatagcgcgcaaa
 actaggataaattatcgccgcgggtgtcatctatgttacttagatcgggattcactggc
 25 cgtcgccccatcgactggaaaaccctggcggttacccacttaatcgcccttgc
 cagcacatcccccttgcggcttgcgtatagcgtatgcgtatggccgcaccgtcgccct
 tcccaacagttgcgtatgcgtatggccgcgttcccttgcgttcccttgcgttcccttgc
 tcgccccatcgactggaaaaccctggcggttacccacttaatcgcccttgcgttcccttgc
 cgatttagtgcgttacggcacctcgacccaaaaacttgattgggtgtggcgttccacg
 30 tagtggccatcgccctgatagacggtttgcgttgcgttgcgttccacgatcttgc
 ttaatagtggactcttgcgttccaaactggaaacaacactcaaccctatctggctattct
 ttgttgcgttccaaactggaaacaacactcaaccctatctggctattct
 ctggggcaaaaccagcgtggaccgcttgcgtcaactctctcaggccaggcggtaagg
 caatcagctgttgcgttccgtctcactggtaaaagaaaaaccacccactacattaaaac

gtccgc_aatgtgttattaa_gttctaa_gcgtcaatttgttacaccacaatatacct
gcca

SEQ ID NO:52 is the nucleic acid sequence of pRD29A-antisense-BnCPP.

- 5 Italicized sequences are the right and left border repeats. Underlined sequence is the RD29A promoter. Sequence in bold is the BnCPP antisense sequence.

SEQ ID NO:53

cgtggagttcccgccacagacccggatgatccccgatcgtaaaacattggcaataaa
gttcttaagattgaatcctgttgcggcttgcgttatcatataattctgttg
aattacgttaagcatgtataattacatgtatgcgtacgttatttatgagatgggt
tttatgatttagagtcccgaattatacatttataacgcgatagaaaacaaaatata
5 ggcgcaactaggataaattatcgcgccgggtcatctatgttactagatcgggcctcc
tgtcaatgctggcggcggctctgggtggttctggcggctctgagggtggct
ctgagggtggcggttctgagggtggcggctctgagggaggcggttccggtggtggct
ggttccggtgatttgattatgaaaagatggcaaacgctaataaggggctatgaccga
aaatgccatgaaaacgcgctacagtctgacgctaaaggcaaacttgattctgtcgcta
10 ctgattacggtgctgctatcgatggttcattggtacggttccggccttgcataatgg
aatggtgctactggtagttgtggctctaattccaaatggctcaagtgggtacgg
tgataattcaccttaatgaataattccgtcaatattacccctccctcaatcg
ttgaatgtcgcctttgtcttgcccaatacgcaaaccgcctctcccgccgtgg
ccgattcattaatgcagctggcacgacaggttcccactggaaagcggcagtgagcg
15 caacgcaattaatgtgagttagtcactcattaggcacccaggcttacacttatgc
ttccggctcgatgtgtggatttgtgagcggataacaatttcacacaggaaacagc
tatgaccatgattacgccaagctggaaatttcgccagttctaaatatccggaaacc
tcttggatgccattgcccattatgttaatttattgacgaaatagacgaaaaggaag
gtggctcctataaagcacatcattgcgataacagaaaggcattgttaagataacct
20 gctgacattggccccaaagtggaaagcaccacccatgaggagcaccgtggagtaagaag
acgttcgagccacgtcgaaaaagcaagtgtgtgatgtatctccattgacgtaagg
gatgacgcacaatccaactatccatcgcaagaccattgtctatataagaaagtata
tcatttcgagtggccacgctgaggggatccatggcgattccttcatggaaaccgtcg
ttggggatgtatgtatgtacgtttttgagacgtattggatctgaggcaacatact
25 gctctcaagcttccactctccaaagactttgggtggagtcattagccaagagaagtt
tgagaaatctcgagttacagtcttgcataaaaagccatttcacttgcattgtatgttt
ttactatacttatggactctgcgattctgtttggatctgccttggtttggaaag
atatctggccgcttctaccatggggactcgatccagagaatgaaatcctgcacac
tcttcattttggctgtttatgacatggcacagatcactgattgccattttctt
30 tgtactcaacttcgtatcgagtctcgcatgggttcaacaaacaaacaaatatggatg
ttcatttagggacatgatcaaaggaataactcctctgtcatacctgccttcatcg
tgccgcaattattgttagttcagaaaaggaggtcattacctcgccatctatctgtgg
cattcatgtttatcctgtcttagtgcgtatgcataaccctgtttgattgcac
ctttcaacaagttcactcccttgcattggagaccccgggagaagattgagaaact

tgcttcttctaaagttcctctgaagaagctgttgcgtggatctacaaggt
 caagccatagtaatgcttacatgtatggtttcaagaacaaaaggattgttcttat
 gacacattgattcagcagtgccagaatgagaatgaaattgtggcggttattgcacacga
 gctgggacactggaagctgaatcacactacatactcgttcattgctgtcaaattccttg
 5 **c**cttcttgcattggaggatacactcttgcagaaactccactgatctcttcaggagt
 tttggtttgatacacaaccagttctcattggttgatcatatttcagcacactgtaat
 accacttcaacacac tagtaagcttgcacctcaacctgttagtcgagcgtttagttc
 aggctgatgctttgcagtgaatcttggttatgcaaaggatctacgtcctgccttagtg
 aagctacaggaagagaacttatcagcgtatgaaacacagacccattgtactcagcttatca
 10 **c**tactcacaccctccttgcattgagaggcttcgagccattgatggagaagacaagaaga
cagattacccccgttgcattttccccgatcgatcattggcaataaagttcttaag
 attgaatcctgttgcggcttgcgtatgattatcatataatttctgttgaattacgtta
 agcatgtatataatttacatgtatgcattgcgttattatgagatgggtttatgatt
 agagtcccgcaattatacatttatgcgtatgaaaacaaaatatacgccgcaacta
 15 ggataaattatcgccgcgggtgtcatctatgttacttagatcggttattactcgccgtc
 gtttacaacgtcgtgactggaaaaccctggcgttacccaacttaatcgccctgcagc
 acatcccccttcgcccagctggcgtatagcgaagaggcccgcaccgatcgcccttccc
 aacagttgcgcagcctgaatggccgcgttcccttcgttcccttcgttccgc
 cacgttgcggcttccctgtcaagctctaattcggggctccctttagggttccgat
 20 tttagtgcttacggcacctcgacccaaaaacttgattgggtatgttacgt
 gggccatcgccctgatagacggtttgcgccttgcgttgcgttggagtcacgttctttaa
 tagtgactcttgttccaaacttggaaacaacactcaaccatatctcggttattcttgg
 atttataaggatttgcgatttgcgaccaccatcaaacaggatttgcgttgcgttgg
 ggc当地accagcgtggaccgcgttgcgtactctctcaggccaggcggtaaggcaat
 25 cagctgttgcggctctactggtaaaagaaaaaccacccaggatcataaaaacgtcc
 gcaatgttattaaagttgtctaagcgtcaatttgcgttacaccacaatatacctgcca

SEQ ID NO:53 is the nucleic acid sequence of MuA-BnCPP. Italicized sequences are the right and left border repeats. Underlined sequence is the MuA promoter.

30 Sequence in bold is the BnCPP sense sequence.

Example 5. Southern Analysis

Genomic Southern blot analysis of transgenic *Arabidopsis* was performed using standard techniques known to one skilled in the art. Typically, 10 μ g of DNA was

electrophoresed in a 0.8% agarose gel and transferred to an appropriate membrane such as Hybond N+ (Amersham Pharmacia Biotech). Pre-hybridization and hybridization conditions were as suggested by the membrane manufacturer, typically at 65°C. The final stringency wash was typically at 1XSSC and 0.1% SDS at 65°C. The NPTII coding 5 region was typically used as the radiolabeled probe in Southern blot analysis.

Thirty-seven *Arabidopsis* lines were selected as homozygous pBI121-AtCPP over-expression lines for further examination. Figure 3 shows a representative blot confirming the presence of the pBI121-AtCPP transgene. Lines were confirmed to be transgenic by PCR analysis using transgene specific primers in the PCR assays.

10 Thirty-three *Arabidopsis* lines were selected as homozygous pBI121-HP-AtCPP hair-pin down-regulation lines for further examination. Figure 4 shows a representative blot confirming the presence of the pBI121-HP-AtCPP hair-pin construct. All lines were confirmed to be transgenic by PCR analysis using transgene specific primers in the PCR assays.

15 *Arabidopsis* lines were selected as homozygous pRD29A-AtCPP over-expression lines for further examination. Figure 5 shows a representative blot confirming the presence of the pRD29A-AtCPP transgene. Lines were confirmed to be transgenic by PCR analysis using transgene specific primers in the PCR assays.

20 *Arabidopsis* lines were selected as homozygous pRD29A-HP-AtCPP lines for further examination. Figure 6 shows a representative blot confirming the presence of the pRD29A-HP-AtCPP transgene. Lines were confirmed to be transgenic by PCR analysis using transgene specific primers in the PCR assays.

Example 6: PCR analysis of transgenic plants

25 PCR was used as a method to confirm the presence of the transgene in all transgenic lines and every construct.. Typical PCR mixtures contained: 1X reaction buffer (10mM Tris-HCl pH 8.8, 1.5mM MgCl₂, 50mM KCl), dNTP's at 200μM, 1pM forward and reverse primer, 2.5U. *Taq* DNA polymerase, and template plus water to a final volume of 50μL. Reactions were run at 1 minute 94°C, 1 minute 60°C, 1 minute 72°C, for 30 cycles. Primers used in the analysis of pBI121-AtCPP and pBI121-HP- 30 AtCPP transgenic plants were as shown in Table 8. Primers used in the analysis of pRD29A-AtCPP were RD29AP1 (SEQ ID NO:66) and SEQ ID NO:7. Primers used in

the analysis of pRD29A-HP-AtCPP transgenic plants were those identified as RD29AP1 (SEQ ID NO:66), SEQ ID NO:8 and SEQ ID NO:8, Nosterm-RV (SEQ ID NO:67).

Table 8.

- 5 pBI121-AtCPP BamFW: 5'-GCCGACAGTGGTCCCAAAGATGG-3'
(SEQ ID NO:10)
- p35S-AtCPP SmaRV: 5'-AAACCCGGGTTAACATCTGTCTTCTTGTCTTCTCCA-3'
(SEQ ID NO:7)
- p35S-HP-AtCPP BamFW: 5'-CTGGAGCTCTTACCGAGGTTGGGCCTTGATCC-3'
10 (SEQ ID NO:8)
- p35S-HP-AtCPP SmaRV: 5'-GCAAGACCGGCAACAGGA-3'
(SEQ ID NO:13)
- pRD29AP1: 5'-TTTAAGCTTGGAGCCATAGATGCAATTCAA -3'
(SEQ ID NO:66)
- 15 pRD29AP1: 5'-TTTAAGCTTGGAGCCATAGATGCAATTCAA -3'
(SEQ ID NO:66)
- Nosterm-RV: 5'-GCAAGACCGGCAACAGGA-3'
(SEQ ID NO:67)

Example 7: Northern analysis of transgenic plants

20 Total RNA was isolated from developing leaf tissue of 27 35S-AtCPP *Arabidopsis* lines (T3 plants). Approximately 10 µg of total RNA was loaded into each lane. The Northern blot was first probed with P³² labeled, single-stranded antisense transcript of AtCPP which detects sense transcript, then stripped and re-probed with cDNA of β-tubulin that was used as a reference. The hybridizing bands of AtCPP and β-
25 tubulin were scanned and quantified using the UN-Scan-It programme (Silk Scientific, Utah, USA), and the ratio of the two hybridizing bands for each sample was obtained. The ratio of the wild type plants was set to 100%, and was compared with those of the transgenic lines. Twenty-one out of twenty-seven lines showed higher expression of AtCPP transcript as compared to the wild type. Values ranged from 104 % to 282 % of

wild type. The results of five lines (35, 84, 76, 136, and 156) of the 21 over-expressing lines is shown in Figure 7.

Example 8: Production of polyclonal antibodies against AtCPP

Anti-AtCPP antibodies were generated using AtCPP fusion protein over-expressed in *E. coli*. The over-expression vector, pMAL-p2, contains 1175 bp malE gene that is located upstream of AtCPP and encodes a 43 KDa maltose-binding protein (MBP). The 1275 bp *Bam*HI/*Sma*I DNA fragment of AtCPP was inserted into pMAL-p2 at *Bam*HI and *Sal*I sites. The *Sal*I site was converted into blunt end using Klenow fragment. The resulting fusion protein MBP-AtCPP was then over-expressed in DH5 α , and purified by one-step affinity for MBP as described by the manufacturer (New England Biolab). The soluble fraction of the crude bacterial extract containing the MBP-AtCPP fusion protein was loaded to a amylose column (1.5 cm x 10.0 cm), and the proteins were eluted with 10 mM maltose in column buffer (50 mM Tris-HCl, pH 7.5, 1 mM EDTA, and 200 mM NaCl). Fractions containing purified MBP-AtCPP fusion protein were pooled, and concentrated with a Centriprep-30 concentrator (Amicon). All purification steps were carried out at 4°C. To generate an antibody, the purified fusion protein was further separated by SDS-PAGE and the Coomassie stained band corresponding to the fusion protein was excised. The identity of the fusion protein was confirmed by Western analysis using anti-MBP antibodies (purchased from New England Biolab). The protein was eluted from the gel slice by electroelution and then emulsified in Ribi adjuvant (Ribi Immunochem) to a final volume of 1 ml. MBP-AtCPP protein was injected into a 3 kg New Zealand rabbit on day 1 and booster injections were given on day 21 and day 35 with 175 μ g of the protein each time. High-titer antisera were obtained one week after the final injection.

25 Example 9: Western blot analysis of 35S-AtCPP transgenic lines using Anti-AtCPP antibodies.

Western analysis was performed to examine expression level of AtCPP in the transgenic lines compared with that of wild type plants. Anti-Bip antibody, an ER luminal protein (Stressgen, Victoria, BC, Canada) was used as a reference. Total proteins were extracted from developing leaf tissue of five ABA^S lines and a wild type control.. The antigenic protein bands of AtCPP and Bip were scanned and quantified using the UN-Scan-It programme (Silk Scientific, Utah, USA) and the ratio of the two

protein bands for each sample was obtained. The ratio of the wild type plants was set to 100%, and was compared with those of the transgenic lines. Data is presented in Figure 7 indicating that the AtCPP protein level was increased in the transgenic lines compared to the wild type plants.

5 **Example 10: ABA sensitivity of transgenic seedlings.**

Approximately 100 seeds were assessed per line per 9 cm plate. Seeds were plated on minimal medium (1/2 MS) supplemented with no ABA or 1.0 µM ABA. Plates were chilled for 3 days at 4 °C in the dark, and incubated for up to 21 days at 22 °C with 24 hour continuous light. Plates were assessed for germination, cotyledon expansion, 10 true leaf development and seedling vigor. Seedlings were assessed for ABA sensitivity over 21 days of growth at which time sensitive seedlings were arrested at the cotyledon stage, lacked true leaves, and showed inhibition of root growth. Wild type control Columbia plants had two to three pairs of true leaves and a well developed root system. Lines were categorized as ABA sensitive (ABA^S) if less than 1% of plants looked like 15 control, moderately ABA sensitive (ABA^{MS}) if more than 1% but less than 50% of looked like control, or ABA insensitive (ABA^{Wt}) if greater than 50% looked like control.

For example, if a plate had 20 healthy seedlings and the control plate had 60 healthy seedlings, the line would be 33% of control and categorized as moderately ABA sensitive.

20 All four vector constructs (pBI121-AtCPP, pBI121Hp-AtCPP, pRD29AHp-AtCPP, pRD29A-ATCPP) have resulted in transgenic lines of *Arabidopsis* which have increased sensitivity to ABA which is indicative of stress tolerance. The data for all 4 constructs is shown in Figure 8. Of the lines transformed with the pBI121-AtCPP construct to over-express the AtCPP gene, 58% (21 out of 36) were classified as 25 sensitive and an added 30% (11 out of 36) were classified as moderately sensitive. These lines were tested again in T4 and T5 generations and their ABA sensitivity was still present indicating that ABA sensitivity is an inheritable trait. Of the lines transformed with the pBI121-HP-AtCPP construct to down-regulate the AtCPP gene by double stranded RNA-inhibition, 15% (7 out of 45) were classified as sensitive and 31% (14 out 30 of 45) were classified as moderately sensitive. To illustrate the increased sensitivity of transgenic lines to ABA, Figure 9 shows the results of germination and seedling development over a range of ABA concentrations. Wild type and pRD29A-HP-AtCPP

are compared. Of the lines transformed with pRD29AHp-AtCPP 70% (12 out of 17) showed high sensitivity and 24% (4 out of 17) showed moderate sensitivity to ABA. Of the lines transformed with pRD29A-AtCPP 29% (5 out of 17) showed high sensitivity and 12% (2 out of 17) moderate sensitivity to ABA. Clearly all 4 transgene constructs
5 are altering ABA sensitivity and ABA signal transduction.

Example 11: Drought Experiments

Arabidopsis plants were grown five plants per 4" or 3" pot, in a replicated water-stress experiment. All pots were filled with equal amounts of homogeneous premixed and wetted soil. Plants were grown under 16 hour daylight (150-200 $\mu\text{mol/m}^2/\text{s}$) at 22 °C
10 and 70% relative humidity. On the day that the first flower opened drought treatment was initiated. First soil water content in each pot was equalized on a weight basis and any further watering of plants was stopped. Daily measurements of soil water content were taken by recording total pot weight. At the end of the drought treatment (6 to 9 days for experiments in 4" pots and 4-5 days for experiments in 3" pots) plants were harvested
15 and shoot dry weights determined. Differences in plant growth were factored into the analysis by expressing water loss on a per gram shoot dry weight basis.

11a) pBI121-AtCPP, Drought stress screen:

Analysis of pBI121-AtCPP transgenic lines during water-stress treatment experiments of up to an eight day period, shows a strong trend towards increased soil
20 water content and reduced water loss per gram of shoot biomass. After three days of water-stress treatment most lines had increased soil water content relative to the wild type control with four out of twenty-four lines, 146, 149, 156 and 97, showing a statistically significant difference. The amount of water lost per gram of shoot biomass was lower for all lines except one (95), and thirteen of these lines were significantly
25 different from the wild type Columbia control (Figure 10). All of the lines showing a statistically significant lower water loss per gram shoot biomass also showed an increased ABA sensitivity. There is also a strong trend, for all but one line (95), which is ABA^{Wt}, towards greater shoot biomass at the end of the drought stress treatment. Seven of those lines 136, 146, 23, 46, 76, 84 and 9, were statistically significant from control at
30 a p=0.05 value.

11b) pBI121-AtCPP, Water loss per gram shoot biomass during water stress treatment:

Lines 35, 76, 95 and a wild type control were grown and placed under a water-stress treatment as above. Plants were harvested at 2 days, 4 days and 6 days of drought treatment. The ABA^S lines, 35 and 76, showed a statistically significant reduction in water-loss relative to shoot dry weight at all three time points (Table 9). Additionally, the two ABA^S transgenic lines had increased shoot biomass, due to increased leaf biomass, and maintained higher soil water contents during drought treatment.

Table 9. Water loss (g) per Shoot dry weight (g) after 2, 4 and 6 days of drought-stress treatment. Values in bold indicate statistically significant differences from Columbia.

Line	2 days		4 days		6 days	
	Mean	Std. Error	Mean	Std. Error	Mean	Std. Error
35	212.5	3.5	308.0	9.9	297.7	11.2
76	227.2	5.8	321.2	8.5	293.8	5.0
95	287.0	5.1	377.3	14.8	348.5	25.5
Columbia	265.3	11.8	408.2	7.7	345.9	6.7
Wild type						

11c) pBI121-AtCPP, Drought stress and shoot recovery:

Water-stress tolerance and determination of post drought-treatment recovery ability was assessed using 20 of the 24 pBI121-AtCPP transgenic lines. Drought treatment was imposed for 6 days after which the plants were watered and allowed to grow for 6 days. Recovered shoot fresh biomass was then determined. Soil water content of these plants was measured daily during the drought treatment and the results confirm previously seen trends. All ABA sensitive (ABA^S) lines that showed a statistically significantly reduction of water loss on a per gram dry weight basis in experiment 11a, continued to show a significant greater soil water content than control plants in this experiment (Table 10). Additionally, Table 10 shows that the recovered shoot fresh biomass after 6 days of drought treatment was significantly greater in all the ABAs lines than Columbia.

Table10. Soil water content on day 3 of drought treatment and recovered shoot fresh weight after 6 days of drought treatment (values in bold were significantly different from Columbia at p=0.05)

Line	ABA status	soil water content day 3		recovered shoot biomass	
		Mean (%) initial)	Std Error	Mean (g)	Std Error
136	ABA ^s	46.6	1.9	4.5	0.16
14	ABA ^s	50.25	0.7	4.1	0.12
146	ABA ^s	45.9	2.5	4.0	0.11
147	ABA ^s	45.1	1.7	4.0	0.15
149	ABA ^s	45.3	1.8	3.8	0.17
156	ABA ^s	47.1	1.9	4.0	0.134
23	ABA ^s	49	1.4	4.0	0.17
33	ABA ^s	46.9	1.6	4.3	0.14
35	ABA ^s	41.7	1.7	4.0	0.11
46	ABA ^s	44.8	1.7	3.8	0.09
63	ABA ^s	46.3	1.4	4.0	0.19
76	ABA ^s	47.8	1.0	3.9	0.17
79	ABA ^s	45.4	1.1	4.1	0.09
84	ABA ^s	46.8	1.9	4.1	0.16
85	ABA ^s	45.3	1.9	4.0	0.12
9	ABA ^s	45.2	2.1	3.9	0.12
93	ABA ^{wt}	43.5	1.2	2.8	0.07
94	ABA ^s	46.9	1.5	3.9	0.13
97	ABA ^s	53	1.2	3.8	0.16
95	ABA ^{wt}	41.9	1.2	2.7	0.06

Columbia	ABA ^{WT}	41.3	1.0	2.7	0.04
----------	-------------------	------	-----	-----	------

11d) pBI121-AtCPP, Seed yield after drought stress treatment:

Seed yield after drought stress during flowering was examined using ten pBI121-5 AtCPP transgenic lines, eight of which were ABA^S. Plants were grown one per 4" pot and were exposed to 9 days of drought treatment as described above. A second group of plants was grown and maintained under well watered conditions as the optimal group. After 9 days of drought treatment plants were re-watered and allowed to continue growth and seed set to maturity. After drought-treatment conditions all eight ABA^S lines had 10 increased yields relative to controls, which ranged from 109% to 126% of the Columbia (Table 11). Drought-treatment resulted in a reduction of yield in all lines, including 15 controls, relative to plants grown under optimal conditions. Expression of the seed yields obtained from drought-treated group relative to the same line under optimal conditions shows that the transgenics preserve a larger percentage of optimal seed yield than do wild type lines.

Table 11. Seed Yield following 9 days drought-treatment

Line	ABA status	Seed Yield (g per plant)		% Columbia	% Optimal
		Mean (g)	Std Error		
156	ABA ^S	0.735	0.044	126.2	83.7
63	ABA ^S	0.675	0.061	116.0	71.0
146	ABA ^S	0.666	0.053	114.4	72.9
94	ABA ^S	0.644	0.052	110.6	68.8
84	ABA ^S	0.642	0.049	110.4	61.8
76	ABA ^S	0.631	0.055	108.5	66.6
136	ABA ^S	0.630	0.051	108.3	74.1
35	ABA ^S	0.614	0.054	105.6	74.2

93	ABA ^{WT}	0.567	0.041	97.5	60.0
95	ABA ^{WT}	0.388	0.088	66.7	43.4
Columbia	ABA ^{WT}	0.582	0.060	100	53.8

11e) pBI121-AtCPP, Seed yield and growth under optimal water conditions:

The lines evaluated above and a number of additional lines were examined in a
5 growth and yield experiment under optimal, well-watered conditions. Results indicated
that the ABA^S lines were shorter at the stage of first open flower, had more rosette
leaves, however, by maturity there were no differences in plant height of transgenics and
Columbia. Moreover, the ABA^S transgenics showed similar or higher seed yields ranging
from 95% to 121% of the wild type control (Figure 11).

10 **11g) pRD29A-HP-AtCPP screen for drought tolerant phenotype:**

Analysis of 17 transgenic lines identified 7 candidate drought tolerant lines (12, 22, 23,
47, 82, 83, 90) on the basis of higher soil water content and lower water loss per g of
shoot dry weight (Table12). All 7 drought tolerant candidate lines showed strong ABA
sensitivity and lines that did not show drought tolerance did not show ABA sensitivity.

15 **Table 12.** Soil water content after 3 days of drought treatment and water lost per g shoot
dry weight. Values in bold are statistically different from those of Columbia wild type
(p=0.05)

	ABA status	soil water content day 2		water lost in 2days/g shootDW	
Line	ABA	Mean (% initial)	Std Error	Mean (g/g)	Std Error
10	ABA ^S	33.4	1.6	199.1	4.5
11	ABA ^S	34.6	3.3	173.1	1.6
12	ABA ^S	36.2	2.0	179.5	5.0
126	ABA ^{MS}	32.5	2.6	199.1	4.1

127	ABA ^{MS}	33.5	2.0	195.6	10.6
14	ABA ^S	32.7	1.2	203	4.9
17	ABA ^S	29.9	1.8	200.7	7.3
22	ABA ^S	39.3	2.1	170.0	3.0
23	ABA ^S	35.7	1.4	174.9	2.6
42	ABA ^{MS}	28	0.7	185.4	5.8
47	ABA ^S	35.9	2.2	181.2	7.7
7	ABA ^{WT}	35	1.3	201.8	5.1
82	ABA ^S	36.7	2.2	178.3	4.0
83	ABA ^S	40	1.4	180.7	6.9
9	ABA ^S	31.4	1.4	173.8	8.7
90	ABA ^S	38.2	1.3	177.6	6.2
93	ABA ^{WT}	30.7	1.8	175.3	4.6
Columbia	ABA ^{WT}	32.1	1.2	196.9	6.2

Example 12. Growth Analysis

The growth analysis of most promising constructs has been set up at 3 stages. Eight plants per line were grown in 3" pots with one plant per pot at 22C, 16hr light (150-200 µmol/m²/s) and 70% RH. Plants were harvested at vegetative growth stage (2 week old seedlings), bolting growth stage (at first open flower) and mid-flowering growth stage (5 to 7 days from first open flower). Also, in some growth experiments additional group of plants was grown in 4" pots (one per pot and 10 plants per line) to maturity for seed yield determinations.

10 **12a)** pBI121-AtCPP growth under optimal and biotic stress conditions

The growth and productivity of pBI121-AtCPP transgenic *Arabidopsis* lines was examined at several stages of development under optimal growth conditions. Although optimal growth conditions were maintained, plants were assessed to be under a degree of

stress that was later determined to be a result of the soil properties. Soil analysis found a fungal contaminant that was believed to be responsible for the biotic stress. This stress could be negated by sterilization of the soil prior to use. Eight ABA^S lines, two with normal ABA sensitivity (ABA^{Wt}) and a wild type Columbia control were analyzed.

5 Figure 12 presents the results of various growth (from mid-flowering stage) and yield parameters and each trait is expressed as a percentage of the Columbia control. The results strongly support an enhanced growth phenotype. This enhanced growth phenotype is present at all growth stages. At the vegetative stage, all ABA^S transgenic plants showed an increase in leaf number relative to that of the wild type with four of the
10 eight lines showing a statistically significant difference. The two ABA^{Wt} lines showed the same or fewer leaves relative to wild type.

15 At the bolting stage ABA^S transgenics showed an increase in leaf number but plants were shorter at this stage (first open flower) than controls. The shoot fresh weight of transgenics was significantly increased relative to that of controls, ranging from 80% to 342% of the wild type. The ABA^S transgenics displayed a delay in flowering from one to three days. The ABA^{Wt} transgenics did not show delayed flowering, increased shoot fresh weight or increased height.

20 At the flowering stage of development the enhanced growth phenotype is maintained (greater leaf number and fresh weight), however, there were no observable differences in plant height indicating that transgenics bolt shorter but reach same final plant height.

25 Of particular significance is the observation, that under these conditions (biotic stress due to presence of fungi in the soil) yields of the ABA^S transgenics were significantly higher, ranging from 120% to 229% of the wild type control. The ABA^{Wt} lines showed similar or slightly reduced yields relative to the Columbia control. This finding indicates that ABA^S transgenic lines are affected less by the biotic stress. This observation has been confirmed, where 5 of the drought tolerant lines were grown in contaminated soil to maturity. The seed yields of transgenic lines, even though greatly reduced relative to optimal conditions, were 2.5 to 4.5 fold higher than those of
30 Columbia wild type (Table 13).

Table 13. Seed yield of pBI121-AtCPP lines grown in contaminated soil. Values in bold indicate statistical differences at p=0.05

Line	ABA sensitivity	Seed Yield per plant (g)	% of Columbia
156	ABA ^S	0.33 ± 0.04	316%
23	ABA ^S	0.35 ± 0.05	336%
76	ABA ^S	0.31 ± 0.04	296%
84	ABA ^S	0.25 ± 0.33	237%
9	ABA ^S	0.48 ± 0.05	455%
Columbia	ABA ^{WT}	0.11 ± 0.03	

12b) pBI121-AtCPP early seedling growth:

Four ABA^S and one ABA^{WT} line plus Columbia were examined for early seedling growth on agar plates. Twenty seeds were plated in a line on agar plates containing 50% MS with 1% sucrose and vitamins and 6 plates per line were used. Plates were placed on slants, which allowed roots to grow downwards. Root length was measured on 7-day old seedlings and shoot and root biomass determined on 11-day old seedlings. Two of the ABA^S transgenic lines had significantly longer roots and all 4 ABA^S lines had shoot dry weights 114% to 123% of controls and root dry weights of 116% to 151% of controls. As a result, the shoot biomass to rootbiomass ratios were slightly reduced in transgenics. These results indicate that enhanced growth of these transgenics is evident in the early growth stage, shortly after germination, and the root growth is more enhanced relative to shoot growth. In a different experiment seedlings were pulled out of agar and roots were stained with toluidine blue to show their structure. Figure 13 shows that transgenic lines had more extensive lateral root system, which would account for greater root biomass.

12c) pRD29A-HP-AtCPP optimal growth characteristics

An optimal growth study has been conducted with 10 lines as described before. Vegetative growth data showed that two of the lines (12 and 9) had significantly more leaves and seven of the lines (12, 22, 23, 47, 82, 9) had significantly greater shoot biomass. Bolting data showed that eight of the lines (12, 22, 23, 47, 82, 9, 90, 93) were significantly delayed in flowering by one to two days, and seven of the lines were significantly shorter than Columbia at first open flower. All of the lines except 42 and 7 had significantly greater number of rosette leaves and shoot FW and this trend is

maintained into the mid-flowering harvest (Figure 14). The plant height, however, by mid-flowering harvest was not significantly different between the transgenic lines and control. All the lines that showed this enhanced growth also showed drought tolerance and ABA sensitivity.

5

Example 13. Ultrastructure pBI121-AtCPP

Two of the drought tolerant and ABA^S lines (35 and 76) plus Wt Columbia were used to examine stem and root cross-sections for any differences in ultrastructure. Free hand sections of mature stems (plants flowering for 10 days) were obtained from above 10 the first node, stained with toluidine blue and preserved with glycerol. The stems of transgenic plants appeared to have more dense cellular structure and contain one or two more vascular bundles than those of Columbia Wt indicating more enhanced water and nutrient transport system.

Leaf disks were taken and fresh weights determined. Transgenic leaf disks were 15 significantly heavier, 20-24% greater than corresponding wild type controls. This increase is believed to be as a result of a thicker leaf.

Example 14. Cold stress experiment pBI121-AtCPP

Four drought tolerant, ABA^S lines (156, 23, 35, 76) and one ABA^{Wt}(95) line plus wild type Columbia were included in a cold stress study. Plants were grown in 3" pots 20 (one per pot) with 10 replicate pots per line at 22C for 10 days (7 days on agar plates and 4 in soil). The cold stress group was moved into 7°C for 5 days while the optimal group was left at 22C. After 5 days in the cold both cold stress group and the optimal group were harvested for shoot biomass determination. ABA^S and drought tolerant lines had significantly greater shoot biomass than Columbia in both optimal (25 to 39% greater 25 shoot fresh weight) and cold stress groups (18 to 44% greater shoot DW) (Table 14).

Results of an eight-day cold stress showed that differences between the transgenic lines and Columbia were even more pronounced (53 to 61% greater shoot fresh weight). This result indicates greater plant vigor and better ability of transgenics to cope with cold stress.

30

Table 14. Shoot fresh weight of optimal and cold stressed (5C for 5d) pBI121-AtCPP. Values in bold indicate statistical difference at p=0.05

Line	ABA sensitivity	Optimal shoot FW		Cold stress shoot FW	
		mg	% of Columbia	mg	% of Columbia
156	ABA ^S	95.4 ± 3.7	137%	23.1 0.7	118%
23	ABA ^S	96.3 ± 3.9	139%	28.3 1.5	144%
35	ABA ^S	87.0 ± 1.7	125%	25.3 1.4	130%
76	ABA ^S	94.7 ± 2.2	136%	27.3 1.5	140%
95	ABA ^{Wt}	67 ± 2.4	96%	21.4 1.0	109%
Columbia	ABA ^{Wt}	69 ± 1.9		19.6 1.1	

Example 15. Drought stress under high temperature pBI121-AtCPP

A drought stress experiment was conducted as described above except that day temperature of 32°C (16hr) and night temperature of 22°C (8hr) was maintained. These temperatures were achieved daily over a 2hr ramping period. Four ABA^S and one ABA^{Wt} line plus Columbia were included. Plants were monitored daily for water loss and soil water content and after 5 days of drought treatment half of the plants were harvested and the other half was re-watered and allowed to recover for four days. Shoots were harvested and shoot fresh weight determined. The results (Table 15) of this experiment showed that previously identified drought tolerant lines maintained their drought tolerant phenotype at high temperature and were able to recover well from the drought stress at high temperature

Table 15. Soil water content on day 2 and water lost in 2 days/final shoot dry weight plus recovery shoot FW after 5days of drought stress at 32C day and 22C night temperatures. Values in bold indicate significant differences from the Columbia control.

line	ABA sensitivity	soil water content day 2	water lost in 2d/shoot DW	recovered shoot FW (g)
136	ABA ^S	50.4 ± 1.1	485.7 ± 18.5	1.30 ± 0.04
146	ABA ^S	52.1 ± 1.0	504.5 ± 7.9	1.15 ± 0.04
35	ABA ^S	52.2 ± 0.8	502.8 ± 15.8	1.19 ± 0.02
76	ABA ^S	52.1 ± 0.6	435.6 ± 10.5	1.11 ± 0.03

95	ABA Wt	50.0 ± 0.9	518.2 ± 13.0	0.86 ± 0.03
Columbia	ABA Wt	48.6 ± 0.6	559.7 ± 19.0	0.84 ± 0.03

Example 16. Heat stress and seed yield pBI121-AtCPP

Two ABA^S lines and one ABA^{Wt} line plus Columbia were examined for the effect of heat stress during flowering on the final seed yield. Plants were grown in 4 inch pots (one/pot) as described above and 9 days from first open flower the temperature was ramped from 22 C to 43C over 2 hours and plants were kept at 43C for 2hr. Temperature was then ramped back to 22C over 2 hours and plants were grown under optimal conditions until maturity. The seed yields from this experiment are shown in Table 16. One of the drought tolerant lines (35) had significantly greater yield than Columbia.

10

Table 16. Seed yield of pBI121-AtCPP lines after two hour 43C heat stress 9 days from first open flower. Values in bold are statistically significant from Columbia.

line	ABA sensitivity	seed yield (g/plant)	seed yield (% of col.)
35	ABA ^S	0.55 ± 0.05	347%
76	ABA ^S	0.24 ± 0.03	148%
95	ABA Wt	0.11 ± 0.02	69%
Columbia	ABA Wt	0.16 ± 0.03	

The effect of heat shock on lines of pBI121-AtCPP at the early flowering stage was assessed. Three ABA^S lines (76, 136, 97) a ABA^{Wt} line (95) and a Columbia wild type control were seeded in 128 cell flats, one flat per line. At the early flowering stage flats were exposed to a temperature of 46.8°C for 50 minutes and then returned to normal growth conditions. Lack of continued growth from main meristems was defined as main meristem death and scored for each line. Data is shown in Table 17.

20

Table 17. Meristem death due to heat shock

Line	Wt	95	76	136	97
% Death	91	97	79	59	18

Example 17. Stomata density determinations pBI121AtCPP

Two ABA^S lines (76 and 35) plus Columbia were examined for stomata density on the upper and lower leaf surface. Nail polish imprints of the upper and lower epidermis were obtained from a fully expanded leaf #5. These imprints were analyzed under the microscope and the number of stomata per $8.7 \times 10^{-8} \text{ m}^2$ were counted. There were no significant differences found between transgenics and Columbia in the stomata of the upper or lower epidermis (Table 18). The increases seen in drought tolerance and reduced water loss is not attributable to a reduced number of leaf stomata.

10

Table 18. Stomata numbers per $8.7 \times 10^{-8} \text{ m}^2$ of abaxial and adaxial epidermis of fully expanded leaf #5 in pBI121AtCPP.

line	ABA sensitivity	stomata on upper epidermis	stomata on lower epidermis
35	ABA ^S	68 ± 5	103 ± 7
76	ABA ^S	58 ± 6	120 ± 16
Columbia	ABA Wt	57 ± 6	116 ± 11

Example 18. CPP Consensus Sequences

15 Also included in the invention is the CPP consensus sequences. The consensus sequences were generated by alignment of the CPP polypeptide and nucleic acid sequences as well as sequences homologous using the program BioEdit.

The “x” in the consensus sequence represents any amino acid or nucleotide. Preferably “x” a conservative amino acid or nucleotide substitution. More preferably, 20 “x” is the most amino acid or nucleotide most prevalent at a given position. For example, the amino acid at position 145 of SEQ ID NO: 73 is a proline as it occurs 66% of the time.

Table 19. ClustalW Analysis of BASF Nucleic Acids

- 1) **BASF_AT1** (SEQ ID NO:21)
- 2) **BASF_AT2** (SEQ ID NO:23)
- 3) **BASF-Corn** (SEQ ID NO:25)
- 4) **BASF-Soy** (SEQ ID NO:27)

5) Consensus (SEQ ID NO:68)

BASF_AT1	TGAAATACTGCATACT	CTTTCAATTCTTGGTGTATGACATGGTCACACATCACTGA	368
BASF_AT2	TGAAATACTGCATACT	CTTTCAATTCTTGGTGTATGACATGGTCACAGATCACTGA	368
BASF-Corn	TGAGATAATACACACCCTTGCTTTCTTAAGCTGGTTCATGGTTGGCAGATTACAGA	80	
BASF-Soy	TGAAATACTGCATAACCCCTTGCCCTTCTTAGCAGGGCTGATGATTGGTCACAGATAACAGA	600	
Consensus	TGAXATACTXCAACXCTTXCXTTCTTGCGXGGXXXATGXXXTGCTXCAAXATXACXGA	600	
	610 620 630 640 650 660		
BASF_AT1	TTGCCATTTCCTTGTACTCAACTT	CGTATGAGTCCTGGCATGGGTCACACAAACA	428
BASF_AT2	TTGCCATTTCCTTGTACTCAACTT	CGTATGAGTCCTGGCATGGGTCACACAAACA	428
BASF-Corn	CTTGGCTTCTCTCTATTCACAACTT	GTTATAGAGGCTGACATGGTTAAACAAGCA	140
BASF-Soy	TTGCCCTTTCTCTACTCAACTT	CGTATGAGGCCGTCATGGTTATAAAGCA	660
Consensus	XTTGCCXTTCTCTACTCAACTT	XGTXATXAGGXCGXCATGGXTTAXAXAA	660
	670 680 690 700 710 720		
BASF_AT1	AACAATATGGATGTCATTAGGACATGATCAAAGGA	CATTCCCTCTGTCTACTAGG	488
BASF_AT2	AACAATATGGATGTCATTAGGACATGATCAAAGGA	CATTCCCTCTGTCTACTAGG	488
BASF-Corn	AACATATGGCTTCTCATTAAGGA	TATGATCAAAGGAATTACTATCCATGATATTGGG	200
BASF-Soy	AACACCATGGTATTCTTAGGACATECTAAAGGAATTTCCTTCCATAATAATTGG	720	
Consensus	AACXXXATGGXTTCTAGGAXATGTXAAGGAAXXXTCXCTXXATAXTXGG	720	
	730 740 750 760 770 780		
BASF_AT1	CCCACCCATTGTGGCGATAATTTCATAGT	CCAGAAAGGAGGTCTTATCTTGCCAT	548
BASF_AT2	CCCACCCATTGTGGCGATAATTTCATAGT	CCAGAAAGGAGGTCTTATCTTGCCAT	548
BASF-Corn	GCACCAATCTGGCTGCTATCAT	CTACATGACAGATTGGAGAACCTTACCTGGCTAT	260
BASF-Soy	TCCACCTATTGTGGCTGCAATTATTGT	ATAGTACAGAAAGGAGGTGATACTTGGCCAT	780
Consensus	XCCACCACTXGTXGCGXATXATXXXXATAGTXCAGAXXGGAGGXCGXTAXXXTXGCA	780	
	790 800 810 820 830 840		
BASF_AT1	CTATCTGGGGCATTCATGTTATCCTGTCCTAGTGTGATGACTATATACCCGGCTT	608	
BASF_AT2	CTATCTGGGGCATTCATGTTATCCTGTCCTAGTGTGATGACTATATACCCGGCTT	608	
BASF-Corn	ATATCTCTGGGTTTATGTTGTTGATTAGTCCTACTGTGATGACAAATATACCCATTGT	320	
BASF-Soy	CTATCTTGGGTTTACGTTGGCTTCCTATTGTTGATGACCCTTATCCAGTACT	840	
Consensus	XTATCTXTGGGXXTTXAXGTTXXXXTXXCTXTXXTGATGATGACXXTAXCCXXTXX	840	
	850 860 870 880 890 900		
BASF_AT1	GATAGCACCGCTCTCAACAAAGTTCACTCCTCTCCAGATGGAGACCTCGGGAGAAAGAT	668	
BASF_AT2	GATAGCACCGCTCTCAACAAAGTTCACTCCTCTCCAGATGGAGACCTCGGGAGAAAGAT	668	
BASF-Corn	GATAGCTCCTCTGGTACAAAGTTCACTCCTCTCCAGATGGAGACCTCGGGAGAAAGAT	380	
BASF-Soy	AATAGCTCCACTCTCAATAAGTTCACTCCACTTCCAGATGGTCACTCAGGGAGAAAAT	900	
Consensus	XATAGCXCCXCTXTCAAXAAGTTCACTCCCTTCCXGAXGGXXXCTXGGAXAA	900	
	910 920 930 940 950 960		
BASF_AT1	TGAGAAACTTGCTCTCTAAAGTTCTTGAAGAAGCTGTTGTTGTCGATGGATC	728	
BASF_AT2	TGAGAAACTTGCTCTCTAAAGTTCTTGAAGAAGCTGTTGTTGTCGATGGATC	728	
BASF-Corn	AGAGAAAGCTGGCAGCTCCCTCAAGTTCTTGAAGAAGCTTCTGTCGATGGATC	440	
BASF-Soy	CGAGAAAATTGCTCTCCCTCAACTACCGTTAAAGAAAGCTTCTGTCGATGGATC	960	
Consensus	XGAGAACTXGCGXXCTXCTXAAXTXTCCTTAAAXCTXTGTXGATGGXTC	960	
	970 980 990 1000 1010 1020		
BASF_AT1	TACAAGGTCAAGCCAATGCAATGCTTACATGTATGGTTCTTAAAGAACAAAAGGATTGT	788	
BASF_AT2	TACAAGGTCAAGCCAATGCAATGCTTACATGTATGGTTCTTAAAGAACAAAAGGATTGT	788	
BASF-Corn	TACCAAGATCAAGCCACAGTAATGCCCTACATGTATGGTTTTCAAGAACAGCGCATAGT	500	
BASF-Soy	CACAAGATCAAGTCAAGCAATGCCATAATGTATGGATTCTTCAGAACAAAGAGGATTGT	1020	
Consensus	XACXAGXTCAAGXCAAGXATGCTXATGTATGGTTXXTAAGAACAAAGGXATXGT	1020	
	1030 1040 1050 1060 1070 1080		
BASF_AT1	TCTTTATGATACGTTGATTCACTGAGCTGCAACATGGAGATGAAATTGTCGGGTTATTGC	848	
BASF_AT2	TCTTTATGATACGTTGATTCACTGAGCTGCAACATGGAGATGAAATTGTCGGGTTATTGC	848	
BASF-Corn	ACTCTATGACACATTGATTCACTGAGCTGAGCAATGGAGATGAGATAGTTCTGTTATAGC	560	
BASF-Soy	CCTTTATGACACATTGATTCAACAGTGCAAGAGACGATGAGGAATTGTCGTGTTATTGC	1080	
Consensus	XTCTXTATGAXACXTTXATTCACTGAGXAXXXAGXAGXAGXATXGTXCXXGTTATXGC	1080	
	1090 1100 1110 1120 1130 1140		
BASF_AT1	ACACGAGCTGGACATTGGAAACTGAATCAACACTACATACTCGTTCACTGCAATTCAAA	908	
BASF_AT2	ACACGAGCTGGACATTGGAAACTGAATCAACACTACATACTCGTTCACTGCAATTCAAA	908	
BASF-Corn	ACATGAACCTGGACACTGGAAACTCAATCATACTGTCCTATTCTCTTCAAGCTGTCAGCT	620	
BASF-Soy	CCATGAGTTGGACACTGGAAAGCTCAACCATACTGTCACACATTCTTGTATGCAAGAT	1140	
Consensus	XCAAGAXXXTGGACATGGAAACTXAACTXACACTXXXTAXXCTTXXTGCGXXTXCAAX	1140	

	1150	1160	1170	1180	1190	1200	
BASF_AT1	CCTTGCCCTTACAATTGGAGGATA	CACTCTTGTCA	GAGAAACTCCACTGATCTC	TTCAG			968
BASF_AT2	CCTTGCCCTTACAATTGGAGGATA	CACTCTTGTCA	GAGAAACTCCACTGATCTC	TTCAG			968
BASF-Corn	GCTTATGGTTCTCAATTGGAGGATA	CACTCTA	GAGAGCTCCAAAGATCTATT	TGG			680
BASF-Soy	TCTTACACTTACAATTGGAGGATA	CACTCTA	GAGAGCTCCAAAGATCTATT	TGG			1200
Consensus	XCTTXXXXTXXX	XCAATTGGAGGATA	XACXTGTXGXXXX	XGATCTX	TXXXG		1200
	1210	1220	1230	1240	1250	1260	
BASF_AT1	GAGTTTCGGATTGATACACAGCCTGT	TCTCATTGGTTGATCATATT	TTCAGCACACTGT				1028
BASF_AT2	GAGTTTCGGATTGATACACAGCCTGT	TCTCATTGGTTGATCATATT	TTCAGCACACTGT				1028
BASF-Corn	AAGTTTGGCTCAAGGACACAGCCAGTA	AAATTGGATTGATCATTT	CCCGCACACC	CAT			740
BASF-Soy	AAGCTTGGGTTGATACGCAGCCAGT	CCTCATGGGCT	CATCATATT	TTCAGCATACTGT			1260
Consensus	XAGXTTXGGXTXXX	XAGCCXGTXXX	XATTGGXX	XATCATXT	XGCA	XACXX	1260
	1270	1280	1290	1300	1310	1320	
BASF_AT1	AAATACCACTGCAACATCCAGTA	AAAGCTTGGCT	CAACCTTGT	TACTTCGAGCGTTGAGTT			1088
BASF_AT2	AAATACCACTGCAACATCTAGTA	AAAGCTTGGCT	GAACCTCGTT	TACTTCGAGCGTTGAGTT			1088
BASF-Corn	AAATACCCATTCACCAACACCT	TCTGAGCTT	TCCTGAAACCTTGT	TCACAGAGCATTTGAATT			800
BASF-Soy	AAATCCCACATTCACCAATTG	TGTGAGCTTGGCT	TGTCAACCTAGTCACCGCAT	TGTATTGAATT			1320
Consensus	ATXCCXXTXCA	XCAXXXXXX	XAGCTT	XGTXAACCTXGTX	XACXX	XAXCXTT	1320
	1330	1340	1350	1360	1370	1380	
BASF_AT1	TCAGGCTGATGCTTTGCTGT	GAAGCTTGGCT	TATGCAAAAGATCT	TTCGTCCTACTCTAGT			1148
BASF_AT2	TCAGGCTGATGCTTTGCTGT	GAAGCTTGGCT	TATGCAAAAGATCT	TTCGTCCTGCTCTAGT			1148
BASF-Corn	TCAGGCTGATGCTTTGCCAAGAA	CCTTGGAT	TATGCCCTCTAGCTCC	GAGCCCTTGT			860
BASF-Soy	TCAGGCTGATGCTTTGCCAAGA	AGCTTGGAT	TATGCACTCTGGAT	TACCCGGTGGCTTGT			1380
Consensus	TCAGGCTGATGXXXX	TGXXXGAA	XCTTGGXT	TATGCCXXXXXXXXXX	XGXXXXXX	CTXGT	1380
	1390	1400	1410	1420	1430	1440	
BASF_AT1	GAAACTACAGGAAGAGAACTT	ATCAGCAATGA	TACTTGATCCATTG	TACTCAGCTTATCA			1208
BASF_AT2	GAAACTACAGGAAGAGAACTT	ATCAGCAATGA	AAAAGATCT	TGTACTCAGCTTATCA			1208
BASF-Corn	TAACACTCAGGAAGAGAACTT	GTC	TGCAGTAA	CACCGATCTTGT	ATTCGGCATATCA		920
BASF-Soy	GAAACTACAGGAAGAGAACTT	GTC	CAGTATGA	TACAGATCTTGT	CTCGTGTGCG	---	1434
Consensus	XAAACTACAGGA	XGAGAAXXX	XTCXGCA	XATGAAX	XACXGATCXX	XGXXXXXXXXXXXXXX	1440
	1450	1460	1470	1480	1490	1500	
BASF_AT1	CTACTCACATCCTCTTGTGAA	AGGCTTGCAGCATTGAT	GGAGAAGAC	AGAACAGAAG			1268
BASF_AT2	CTACTCACATCCTCTTGTGAA	AGGCTTGCAGCATTGAT	GGAGAAGAC	AGAACAGAAG			1268
BASF-Corn	CTACTCCCCACCCACCACT	CGTCGAGAGGCT	GAAGCTT	TGGAAGATT	CAGACGACA	AAA	980
BASF-Soy	-----	-----	-----	-----	-----	-----	1434
Consensus	XXXXXXXXXXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	1500
	1510	1520	1530	1540	1550	1560	
BASF_AT1	AGATTAA	-----	-----	-----	-----	-----	1275
BASF_AT2	AGATTAA	-----	-----	-----	-----	-----	1275
BASF-Corn	AGAAGATTAGTCGATCCTGT	TATGAGTTACATATGG	ATTTCCCTGCC	CACATGCACA	1040		
BASF-Soy	-----	-----	-----	-----	-----	-----	1434
Consensus	XXXXXXXXXXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	1560
	1570	1580	1590	1600	1610	1620	
BASF_AT1	-----	-----	-----	-----	-----	-----	1275
BASF_AT2	-----	-----	-----	-----	-----	-----	1275
BASF-Corn	CCGATTCA	GTC	TGGATGGT	GAGGGTTTGACATAGGAGTGT	CAAAGCTT	AGAGT	1100
BASF-Soy	-----	-----	-----	-----	-----	-----	1434
Consensus	XXXXXXXXXXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	1620
	1630	1640	1650	1660	1670	1680	
BASF_AT1	-----	-----	-----	-----	-----	-----	1275
BASF_AT2	-----	-----	-----	-----	-----	-----	1275
BASF-Corn	GCATCTT	CGTCAGGT	GCAACAGC	CTTCGGTCATTGAGAC	ATATAAGCGA	ATTAGCTA	1160
BASF-Soy	-----	-----	-----	-----	-----	-----	1434
Consensus	XXXXXXXXXXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	XXXXXXXXXXXXXX	1680
	1690	1700	1710	1720	1730	1740	
BASF_AT1	-----	-----	-----	-----	-----	-----	1275
BASF_AT2	-----	-----	-----	-----	-----	-----	1275

BASF-Corn	TTAAAAAAAAACAGAACTGTTGCATCAAAAAAAAAAAAAAGAAAACAAAAA	1220
BASF-Soy	-----	1434
Consensus	XX	1740
	1750 1760 1770 1780 1790 1800	
BASF_AT1	-----	1275
BASF_AT2	-----	1275
BASF-Corn	AAAAAAAAAAAAGAAAAAAGAAAAAGTGCCTCTGCCTGTTACACTGCTTG	1280
BASF-Soy	-----	1434
Consensus	XX	1800
	1810 1820	
BASF_AT1	-----	1275
BASF_AT2	-----	1275
BASF-Corn	CCCTATAGTGTATCGTACAGA	1301
BASF-Soy	-----	1434
Consensus	XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX	1821

Table 20. ClustalW Analysis of BASF Amino Acids

- 1) **BASF_AT1** (SEQ ID NO:22)
- 2) **BASF_AT2** (SEQ ID NO:24)
- 3) **BASF-Corn** (SEQ ID NO:26)
- 4) **BASF-Soy** (SEQ ID NO:28)
- 5) **consensus** (SEQ ID NO:69)

	10 20 30 40 50 60	
BASF_AT1	MAIP E METVVGFMI V MYIFETYLD E RQLTALKLPTLPKT I GVISQEKF E KSRAYSLDKS	60
BASF_AT2	MAIP E METVVGFMI V MYIFETYLD E RQLTALKLPTLPKT I GVISQEKF E KSRAYSLDKS	60
BASF-Corn	-----	1
BASF-Soy	MAFP P YMEA V VGFMI V YIFETYLD V RQH A LKLPTLPKT I GVISQEKF E KSRAYSLDKS	60
Consensus	BASF XXX	60
	70 80 90 100 110 120	
BASF_AT1	Y FHFVHEFTI T MDMSAILE F GILPWFWRMSGAV E PRLGLD P ENEILHTLSFLAGV Y WTWSH	120
BASF_AT2	Y FHFVHEFTI T MDMSAILE F GILPWFWRMSGAV E PRLGLD P ENEILHTLSFLAGV N WTWS	120
BASF-Corn	-----	24
BASF-Soy	H FHFVHEFTI T YDSTI L YFC G LPWFWRKSD F MTIA C FN A ENEILHTLAFLAGSMVWSQ	120
Consensus	BASF XXX	120
	130 140 150 160 170 180	
BASF_AT1	ITDLPFLSYSTFVI E SRHGFNQQT I W M FRIDMIKG T FLSVILG P PIVA A II E IVQKGGPY	180
BASF_AT2	ITDLPFLSYSTFVI E SRHGFNQQT I W M FRIDMIKG T FLSVILG P PIVA A II E IVQKGGPY	180
BASF-Corn	ITDLPFLSYSTFVI E ARHGFnQQT I W M FRIDMIKG T LLS M ILG P PIVA A II E IVC I GGPY	84
BASF-Soy	ITDLPFLSYSTFVI E ARHGFnQQT P W M FRIDMIKG I FLSVI E G P PIVA A II E IVVQKGGPY	180
Consensus	BASF ITDLPFLSYSTFVI E XRHGFnQQT X W M FRDMXKGXXLS X IXG P PIVA A II E IVC X GGPY	180
	190 200 210 220 230 240	
BASF_AT1	LAIYLWAFM E DSLVM M TIY P V L IAPLFNK T PLPDG D LREKIEKLASSLK F PLKKLFVV	240
BASF_AT2	LAIYLWAFM E DSLVM M TIY P V L IAPLFNK T PLPDG D LREKIEKLASSLK F PLKKLFVV	240
BASF-Corn	LAIYLWGFM M LA L MM M TIY P IAPLFNK T PLP G GV L REKIEKLASSLK F PLKKLFVV	144
BASF-Soy	LAIYLWVFT E GLS M MM M TIY P V L IAPLFNK T PLPDG D OLREKIEKLASSLN Y PLKKLFVV	240
Consensus	BASF LAIYLAXE E EXLXXXMM M TXYP X IAPLFNK T PLPXG X REKIEKLAXSIXXPLKKLFVV	240
	250 260 270 280 290 300	
BASF_AT1	DGSTRSSHSNAYMYGFFKNKRIVLYDTL I QQCKNEDEIVAVIAHELGHWKL N H T TYSF E A	300
BASF_AT2	DGSTRSSHSNAYMYGFFKNKRIVLYDTL I QQCKNEDEIVAVIAHELGHWKL N H T TYSF E A	300
BASF-Corn	DGSTRSSHSNAYMYGFFKNKRIVLYDTL I QQCSNEDEIV S VIAHELGHWKL N H T VYSF M A	204
BASF-Soy	DGSTRSSHSNAYMYGFFKNKRIVLYDTL I QQCK D DEEIVAVIAHELGHWKL N H T VYTF V A	300
Consensus	BASF DGSTRSSHSNAYMYGFFKNKRIVLYDTL I QQCX XXX EIV X VIAHELGHWKL N H T XY X A	300
	310 320 330 340 350 360	
BASF_AT1	VQILAFLQFGGGYTLVRNSTDLFRSFGFDTQPVLIGLI I FOHTV I PLQH P V S FGLNLVSRA	360

BASF_AT1	VQILALFLQFGGYTLVRNSTDLFRSGFDFDTQPVLIGLIIIFQHTVPLQLHLVSFGGLNLVSR	360
BASF_Corn	VCLMLMFLQFGGYTLVRNSKDLFGSFGFKDQPVIGLIIFPHTIPIHLSFRFLNLVSR	264
BASF_Soy	MQILITLLQFGGYTLVRNSADLYRSFGFDTQPVLIGLIIIFQHTVPLQLOLVSFGGLNLVSP	360
Consensus BASF	XQXLXXLQFGGYTLVPSXDLXXSFGFXXOPVXIGLIIFXHTXIPXXXXXXSFXXLNLVSRX	360
	370 380 390 400 410 420	
BASF_AT1	FEFQADAFAVKLGYAKDLPPTLVKLQUEENLSAMNTDPLYSAYHYSHPPLVERLRAIDGED	420
BASF_AT2	FEFQADAFAVKLGYAKDLPPALVKLQUEENLSAMRTDLYISAYHYSHPPLVERLRAIDGED	420
BASF_Corn	FEFQADAFAKNLGYAPQLRAALVKLQUEENLSAMNTDPWYSAHYSHPPLVERLQALEDSD	324
BASF_Soy	FEFQADGFAKKLGYASCLPFGGLVKLQUEENLSAMNTDPCSC-----	400
Consensus BASF	FEFQADXFAXXLGYAXXLPXXLVKLQUEENLSAMXTXXXXXXXXXXXXXXXXXXXXXX	420
	430 440 450 460 470 480	
BASF_AT1	KPTD-----	424
BASF_AT2	KPTD-----	424
BASF_Corn	DPKEDSILVGLHMDFSLPHAHRFSAWMVRVLTECCQSFRVHLSVRCNSLSVIETYKRISY	384
BASF_Soy	XX	400
Consensus BASF	XX	480
	490 500 510 520 530 540	
BASF_AT1	-----	424
BASF_AT2	-----	424
BASF_Corn	KKQNCCCIKKKKKETKKKKKKKKKKKKVLCVTTACPIVIVS-----	429
BASF_Soy	-----	400
Consensus BASF	XX	525
	550 560 570 580 590 600	
BASF_AT1	-----	424
BASF_AT2	-----	424
BASF_Corn	-----	429
BASF_Soy	-----	400
Consensus BASF	-----	525
	610 620 630 640 650 660	
BASF_AT1	-----	424
BASF_AT2	-----	424
BASF_Corn	-----	429
BASF_Soy	-----	400
Consensus BASF	-----	525
	670 680 690 700 710 720	
BASF_AT1	-----	424
BASF_AT2	-----	424
BASF_Corn	-----	429
BASF_Soy	-----	400
Consensus BASF	-----	525
	730 740 750 760 770 780	
BASF_AT1	-----	424
BASF_AT2	-----	424
BASF_Corn	-----	429
BASF_Soy	-----	400
Consensus BASF	-----	525
	790 800 810 820 830 840	
BASF_AT1	-----	424
BASF_AT2	-----	424
BASF_Corn	-----	429
BASF_Soy	-----	400
Consensus BASF	-----	525
	850 860 870 880 890 900	
BASF_AT1	-----	424
BASF_AT2	-----	424
BASF_Corn	-----	429
BASF_Soy	-----	400
Consensus BASF	-----	525

Table 21. ClustalW Analysis of Generic Nucleic Acids

- 1) **afc1** (SEQ ID NO:29)
 - 2) **AT4g01320** (SEQ ID NO:31)
 - 3) **AF007269** (SEQ ID NO:33)
 - 4) **Consensus** (SEQ ID NO:70)

afcl	TCCTTTGAAGAAGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATG-----	751
AT4g01320	TCCTTTGAAGAAGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATG-----	772
AF007269	TCCTTTGAAGAAGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATGTGAG	2040
Consensus	TCCTTTGAAGAAGCTGTTGTCGATGGATCTACAAGGTCAAGCCATAGCAATGXXXX	2040
	2050 2060 2070 2080 2090 2100	
afcl	751
AT4g01320	----- ----- ----- ----- ----- ----- ----- ----- -----	772
AF007269	AAGCTTGAGATCTTCTACCTACTTACTCTAGTTTACCAATTAGAAGCTTACGTATCT	2100
Consensus	XX 2100	
	2110 2120 2130 2140 2150 2160	
afcl	795
AT4g01320	----- ----- ----- ----- ----- ----- ----- ----- -----	816
AF007269	TGTTACATCATACAGGCTTACATGTATGGTTCTTAAGAACAAAAGGATTGTTCTTAT	2160
Consensus	XXXXXXXXXXXXXXCTTACATGTATGGTTCTTAAGAACAAAAGGATTGTTCTTAT	2160
	2170 2180 2190 2200 2210 2220	
afcl	GATACTGTTGATTCAGCAG----- ----- ----- ----- ----- -----	813
AT4g01320	GATACTGTTGATTCAGCAG----- ----- ----- ----- ----- -----	834
AF007269	GATACTGTTGATTCAGCAGGTACTGTGACTCTTGATGCTTCAACAGAGCTATACTCACATT	2220
Consensus	GATACTGTTGATTCAGCAGXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX 2220	
	2230 2240 2250 2260 2270 2280	
afcl	829
AT4g01320	----- ----- ----- ----- ----- ----- ----- ----- -----	850
AF007269	TCTGTTTCTGGTTCTGAAACATAACATAATCTTCTATTGTGCGATGCAAGAACATGAGGATG	2280
Consensus	XXTGCAAGAACATGAGGATG	2280
	2290 2300 2310 2320 2330 2340	
afcl	889
AT4g01320	AAATTGTGGCGGTTATTGCACACAGAGCTGGACATTGAAAAGTGAATCACACTACACT	910
AF007269	AAATTGTGGCGGTTATTGCACACAGAGCTGGACATTGAAAAGTGAATCACACTACACT	2340
Consensus	AAATTGTGGCGGTTATTGCACACAGAGCTGGACATTGAAAAGTGAATCACACTACACT	2340
	2350 2360 2370 2380 2390 2400	
afcl	CGTTCATTGCGATTCAA----- ----- ----- ----- ----- -----	906
AT4g01320	CGTTCATTGCGATTCAA----- ----- ----- ----- ----- -----	927
AF007269	CGTTCATTGCGATTCAAGTGAGGCTCAACCGACAGTCAAAAAACTTACTCACATCTACAT	2400
Consensus	CGTTCATTGCGATTCAAXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX 2400	
	2410 2420 2430 2440 2450 2460	
afcl	915
AT4g01320	----- ----- ----- ----- ----- ----- ----- ----- -----	936
AF007269	TTCACCTAACGAAATCATGTCTTATGACCCCTCTCAATGTTTGCTTCAGATCCTTGCC	2460
Consensus	XXATCCTTGCC	2460
	2470 2480 2490 2500 2510 2520	
afcl	975
AT4g01320	TTCTTACAATTGGAGGATACACTCTTGTCTCAGAAACTCCACTGATCTCTCAGGAGTTTC	996
AF007269	TTCTTACAATTGGAGGATACACTCTTGTCTCAGAAACTCCACTGATCTCTCAGGAGTTTC	2520
Consensus	TTCTTACAATTGGAGGATACACTCTTGTCTCAGAAACTCCACTGATCTCTCAGGAGTTTC	2520
	2530 2540 2550 2560 2570 2580	
afcl	1020
AT4g01320	GGATTTGATACACAGCCTGTTCTCATTGGTTTGATCATATTTCAG----- -----	1041
AF007269	GGATTTGATACACAGCCTGTTCTCATTGGTTTGATCATATTTCAGGTTGTTATTTTGC	2580
Consensus	GGATTTGATACACAGCCTGTTCTCATTGGTTTGATCATATTTCAGXXXXXXXXXXXXXX	2580
	2590 2600 2610 2620 2630 2640	
afcl	1020
AT4g01320	----- ----- ----- ----- ----- ----- ----- ----- -----	1041
AF007269	CTTTGACACTAACATCAATGAATCAAGGATGGATTAGAAGAAAAAAACTCTAACACCTTG	2640
Consensus	XX 2640	
	2650 2660 2670 2680 2690 2700	

afcl	CACACTGTAATACCACTGCAACATCTAGTAAGC	1053
AT4g01320	CACACTGTAATACCACTGCAACATCTAGTAAGC	1074
AF007269	GTTATATCCTCTGCTGATTATCACAG	2700
Consensus	XXXXXXXXXXXXXXXXXXXXXXXXXXXXXX	2700
	2710 2720 2730 2740 2750 2760	
afcl	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG	1093
AT4g01320	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG	1114
AF007269	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG	2760
Consensus	TTTGGCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG	2760
	2770 2780 2790 2800 2810 2820	
afcl	1093
AT4g01320	1114
AF007269	AGATCCAACCATAAGTTCTTTATTGCAATGGCAGCCTCATCTACTAATCTGAGTTAACGT	2820
Consensus	XX	2820
	2830 2840 2850 2860 2870 2880	
afcl	1141
AT4g01320	CTGATGCTTTGCCTGTAAGCTTGGCTATGCAAAGATCTCGTCTG	1141
AF007269	CTGATGCTTTGCTGTGAAGCTTGGCTATGCAAAGATCTCGTCTG	1162
Consensus	TCCCTTTGCAGGCTGATGCTTTGCTGTGAAGCTTGGCTATGCAAAGATCTCGTCTG	2880
	XXXXXXXXXXXXCTGATGCTTTGCCTGTAAGCTTGGCTATGCAAAGATCTCGTCTG	2880
	2890 2900 2910 2920 2930 2940	
afcl	CTCTAGTAACTACAGG	1159
AT4g01320	CTCTAGTAACTACAGGTCAAGAGAGATAACAACAGAACACAAACTGTTACCTCAATT	1222
AF007269	CTCTAGTAACTACAGGTCAAGAGAGATAACAACAGAACACAAACTGTTACCTCAATT	2940
Consensus	CTCTAGTAACTACAGGXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX	2940
	2950 2960 2970 2980 2990 3000	
afcl	1177
AT4g01320	GTGTCACACACTTAATGGATTTTGTTGGGATTTGCGAGGAAGAGAACTTATCAGCAA	1282
AF007269	GTGTCACACACTTAATGGATTTTGTTGGGATTTGCGAGGAAGAGAACTTATCAGCAA	3000
Consensus	XXAAGAGAACTTATCAGCAA	3000
	3010 3020 3030 3040 3050 3060	
afcl	1237
AT4g01320	TGAACACTGATCCATTGCACTCAGCTTATCACTACTCACATCCTCCTCTGTTGAAAGGC	1342
AF007269	TGAACACTGATCCATTGACTCAGCTTATCACTACTCACATCCTCCTCTGTTGAAAGGC	3060
Consensus	TGAACACTGATCCATTGACTCAGCTTATCACTACTCACATCCTCCTCTGTTGAAAGGC	3060
	3070 3080 3090	
afcl	1275
AT4g01320	TTCGAGCCATTGATGGAGAAGACAAGAAGACAGATTAA	1380
AF007269	TTCGAGCCATTGATGGAGAAGACAAGAAGACAGATTAA	3098
Consensus	TTCGAGCCATTGATGGAGAAGACAAGAAGACAGATTAA	3098

Table 22. ClustalW Analysis of Generic Amino Acids

- 1) **afcl** (SEQ ID NO:30)
 - 2) **AT4g01320** (SEQ ID NO:32)
 - 3) **AF007269** (SEQ ID NO:34)
 - 4) **Consensus** (SEQ ID NO:71)

Sequence alignment showing conservation of amino acids across different accessions:

	10	20	30	40	50	60	70	80	90	100	110	120				
afcl				
AT4g01320	MA	IP	FMET	VVG	FMIV	MYI	FETY	LLR	RQL	TALK	LPTLP	KTLVG	VISQEK	FEKSRAY	RDI	58
AF007269	MA	IP	FMET	VVG	FMIV	MYI	FETY	LLR	RQL	TALK	LPTLP	KTLVG	VISQEK	FEKSRAY	RDI	60
Consensus Publi	MA	IP	FMET	VVG	FMIV	MYI	FETY	LLR	RQL	TALK	LPTLP	KTLVG	VISQEK	FEKSRAY	RDI	60

Note: The sequence ends at position 60 for AF007269 and AT4g01320.

afcl	K-----SYFHFVHEFTILMDSAILFFGILPWFWKMSGAVLPRLGLDPENEILHHTLSFLA	113
AT4g01320	ENFNICSYFHFVHEFTILMDSAILFFGILPWFWKMSGAVLPRLGLDPENEILHHTLSFLA	120
AF007269	-----T-----	42
Consensus Publi	XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX XXXXXXXXXXXXXXXXXXXXXX	120
	130 140 150 160 170 180	
afcl	
AT4g01320	GVMNTWSQITDLPFLSYLSTFVIESRHFNFNKQTIWFMFIRDMIKGTFLSVILGPPIVAAIFI	173
AF007269	GVMNTWSQITDLPFLSYLSTFVIESRHFNFNKQTIWFMFIRDMIKGTFLSVILGPPIVAAIFI	180
Consensus Publi	-----DLPFLSYLSTFVIESRHFNFNKQTIWFMFIRDMIKGTFLSVILGPPIVAAIFI	93
	XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX XXXXXXXXXXXXXXXXXXXXXX	180
	190 200 210 220 230 240	
afcl	
AT4g01320	VQKGGPYLAIYLWAFMFILSLVMMTIYPVLIAPLFNKFTPLPDGDLREKIEKLASSLKFP	233
AF007269	VQKGGPYLAIYLWAFMFILSLVMMTIYPVLIAPLFNKFTPLPDGDLREKIEKLASSLKFP	240
Consensus Publi	VQKGGPYLAIYLWAFMFILSLVMMTIYPVLIAPLFNKFTPLPDGDLREKIEKLASSLKFP	153
	XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX XXXXXXXXXXXXXXXXXXXXXX	240
	250 260 270 280 290 300	
afcl	
AT4g01320	LKKLFVVDGSTRSSHSNAYMYGFFKNKRIVLYDTLIQQCKNEDEIVAVIAHELGHWKLNH	293
AF007269	LKKLFVVDGSTRSSHSNAYMYGFFKNKRIVLYDTLIQQCKNEDEIVAVIAHELGHWKLNH	300
Consensus Publi	LKKLFVVDGSTRSSHSNAYMYGFFKNKRIVLYDTLIQQCKNEDEIVAVIAHELGHWKLNH	213
	XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX XXXXXXXXXXXXXXXXXXXXXX	300
	310 320 330 340 350 360	
afcl	
AT4g01320	TTYSFIAVQILAFLQFGGYTLVRNSTDLFRSGFDTQPVLIGLIIIFQHTVIPLQHLVSFG	353
AF007269	TTYSFIAVQILAFLQFGGYTLVRNSTDLFRSGFDTQPVLIGLIIIFQHTVIPLQHLVSFG	360
Consensus Publi	TTYSFIAV-----QHTVIPLQHLVSFG	235
	XXXXXXXXXXXXXXXXXXXXXXXXXXXXXX XXXXXXXXXXXXXXXXXXXXXX XXXXXXXXXXXXXXLVSXX	360
	370 380 390 400 410 420	
afcl	
AT4g01320	LNLVSRAFEFQADAFAVKLGYAKDLRPAVLKLC-----	386
AF007269	LNLVSRAFEFQADAFAVKLGYAKDLRPAVLKLVQREDNNRTQT-----	420
Consensus Publi	LNLVSRAFEFQADAFAVKLGYAKDLRPAVLKLVQREDNNRTQT-----	278
	XXXXXXXXAFXXXXXXXXXXXXX XXXXXXXXXXXXXXXXXXXXXX XXXXXXXXXXXXXXGXX	420
	430 440 450 460 470 480	
afcl	
AT4g01320	-EENLSAMNTDPLHSAYHYSHPPLVERLRAIDGEDKKTD-----	424
AF007269	QEENLSAMNTDPLHSAYHYSHPPLVERLRAIDGEDKKTD-----	459
Consensus Publi	-EENLSAMNTDPLHSAYHYSHPPLVERLRAIDGEDKKTD-----	316
	XXXXLSAMNTDPLHSAYHYSHPPLVERLRAIDGEDKKTD-----	480

Table 23. ClustalW Analysis of PPI Nucleic Acids

- 1) **PPI-AtCPP** (SEQ ID NO:1)
- 2) **PPI-BnCPP** (SEQ ID NO:14)
- 3) **PPI-SoyCPP** (SEQ ID NO:17)
- 4) **Consensus** (SEQ ID NO:72)

	10 20 30 40 50 60	
PPI-AtCPP	
PPI-BnCPP	ATGGCGATTCTCTTCATGGAAACCGTCGTGGTTTATGATAGTGTACATTGGAG	60
PPI-SoyCPP	ATGGCGATTCTCTTCATGGAAACCGTCGTGGTTTATGATAGTGTACATTGGAG	60
Consensus	ATGGCGATTCTCTTCATGGAAACCGTCGTGGTTTATGATAATGTACATTGGAG	60
	
PPI-AtCPP	
PPI-BnCPP	ACGTATTTGGATCTGAGGCCACTCACTGCTCTCAAGCTTCCAACTCTCCCAGAAACCTTG	120
PPI-SoyCPP	ACGTATTTGGATCTGAGGCCACATACTGCTCTCAAGCTTCCAACTCTCCCAGAAACCTTG	120
Consensus	ACTTACCTTGGATCTGCGACAACATAGGCCCTCAACTTCCACTCTTCCAAAGACTTTA	120
	ACXTAXTTGGATXTGXGXCAACXXXXGXCTCAAXCTTCCAXCTCTXXCAAXACXTTX	120
	130 140 150 160 170 180	
PPI-AtCPP	
PPI-BnCPP	GTTGGTGTATTAGCCAAGAGAAGTTGAGAAATCACGAGCATACAGTCTTGACAAAAGC	180
PPI-SoyCPP	GTTGGTGTATTAGCCAAGAGAAGTTGAGAAATCTCGAGCTTACAGTCTTGACAAAAGC	180
	GAGGGTGTATCAGCCAAGAGAAATTGAGAAATCTAGAGCCTATAGCTTGATAAAAGC	180

Consensus	GXXGCGXGTAXATXAGCCAAGAGAAATTGGAGAAATCXXGAGCXATAAGTCTTGAXAAAAGC	180
190.....200.....210.....220.....230.....240	
PPI-AtCPP	TATTTTCACTTTGTCATGAGTTGTAACCTATACTTATGGACTCTGCACATTGTTCTTT	240
PPI-BnCPP	CATTTTCACTTTGTCATGAGTTGTTACCTATACTTATGGACTCTGCACATTGTTCTTT	240
PPI-SoyCPP	CACTTCCATTTGTCACGAGTTGTCACAGACTCTACAAATTGTTACTTT	240
Consensus	XAXTTXCAAXTTGTCAXGAGTTGTXACXATAXTAXXXGACTCTXXATTXGTGXCTTT	240
250.....260.....270.....280.....290.....300	
PPI-AtCPP	GGGATCTTGCCCTGGTTTGGAAAGATGTCTGGAGCTCTTACCGAGGTTGGGCTTGAT	300
PPI-BnCPP	GGGATCTTGCCCTGGTTTGGAAAGATATCTGGCGGCTTCTACCAATGGTGGGACTCGAT	300
PPI-SoyCPP	GGGGTATTGCCCTGGTTTGGAAAGAACATCAGGAGATTATGACAATAGCTGTTTCAAT	300
Consensus	GGGXXXTTGTGCOXGGTTTGGAAAGAXTCXGGXGXXXTTTXXXXXAXXXXXGXXXTXXAT	300
310.....320.....330.....340.....350.....360	
PPI-AtCPP	CCGGAGAATGAAATACTGCATACTCTTCATTCTTGGCTGGTGTATGACATGGTCACAG	360
PPI-BnCPP	CCAGAGAATGAAATCCTGCACACTCTTCATTCTTGGCTGGTCTTATGACATGGTCACAG	360
PPI-SoyCPP	GCTGAGAATGAAATACTGCATAACCTTGCCTCTTAGCAGGGCTGATGATTGGTCACAG	360
Consensus	XCGAGAATGAAATXCTGCAACXXCTTXXCTTXXGCGXGGXXATGAXXTGGTCACAG	360
370.....380.....390.....400.....410.....420	
PPI-AtCPP	ATCACTGATTGCACTTTCTTGTACTCAACTTTCGTGATCGAGTCGGCATGGGTTTC	420
PPI-BnCPP	ATCACTGATTGCACTTTCTTGTACTCAACTTTCGTGATCGAGTCGGCATGGGTTTC	420
PPI-SoyCPP	ATAACAGATTGCGCTTTTCTGTGACTCAACTTTGTGATTGAGGCCGTCATGGTTT	420
Consensus	ATXACXGATTGCGCTTTTCTGTGACTCAACTTTGTGATXGAGXXCGXCATGGXTTX	420
430.....440.....450.....460.....470.....480	
PPI-AtCPP	AACAAACAAACAAATGGATGTTCATAGGGACATGATCAAAGGAACATTCCCTCTGTC	480
PPI-BnCPP	AACAAACAAACAAATGGATGTTCATAGGGACATGATCAAAGGAATACCTCCCTCTGTC	480
PPI-SoyCPP	AAATAACAAACACCATTGGTTATTCCTTGTGACTCAACTTTGTGATTGAGGCCGTCATGGTTT	480
Consensus	AAAXAAXCAAACAXXATGGXXTTCCTTGTGACTCAACTTTGTGATXGAGXXCGXCATGGXTTX	480
490.....500.....510.....520.....530.....540	
PPI-AtCPP	ATACTAGGCCACCCATTGTTGCTGCCATAATTTCATAGTCAGAAAGGAGGTCTTAT	540
PPI-BnCPP	ATACCTGCCCTCCCTATCGTTGCCAAATTATTGTTATAGTTTCAGAAAGGAGGTCTTAC	540
PPI-SoyCPP	ATAATTGGTCCACCTATTGTGCTGCAATCATTGTAATAGTACAGAAAGGAGGTCCATAC	540
Consensus	ATAXXXGXXCCXXATXGTGCGXGXXATXATTXTXATAGTIXCAGAAAGGAGGTCTXTG	540
550.....560.....570.....580.....590.....600	
PPI-AtCPP	CTTGGCATCTATCTGTGGGCATTCATGTTTATCCTGTCCTAGTGTGATGACTTATAC	600
PPI-BnCPP	CTTGGCATCTATCTGTGGGCATTCATGTTTATCCTGTCCTAGTGTGATGACTTATAC	600
PPI-SoyCPP	TTGGGCATCTATCTTGGGTTTTACCTTGGCTTCTTGTGATGATGACCTTTAT	600
Consensus	XTXGCCATCTATCTXTGGGXXTIXXAGTTXXXCTXTCTXTGTGATGATGACXXXTAX	600
610.....620.....630.....640.....650.....660	
PPI-AtCPP	CCGGTCTTGTATAGCACCGCTCTCAACAAATTCACTCTCTCCAGATGGAGACCTCCGG	660
PPI-BnCPP	CCTGTTTGTATTCACCTCTTCAACAAAGTTCACTCTCTCCGTATGGAGACCTCCGG	660
PPI-SoyCPP	CCAGTACTAATAGCTCCACTCTTCAATAAGTCACTCCACTTCCAGATGGTCACACTCAGG	660
Consensus	CCXGTXXXATXGCGXXCTXTCAAXAATTCACTCCCTCCXGATGGXXXCTOXGG	660
670.....680.....690.....700.....710.....720	
PPI-AtCPP	GAGAAGATTGAGAAACTTGCTCTCCCTAAAGTTCCCTTGAGAAGAGCTGTTGTC	720
PPI-BnCPP	GAGAAGATTGAGAAACTTGCTCTCTCTAAAGTTCCCTTGAGAAGAGCTGTTGTC	720
PPI-SoyCPP	GAGAAATAGAGAAACTTGCTCTCCCTCAACTATGCGTTAAAGAAACTATTGTC	720
Consensus	GAGAAAXATXGAGAAACTTGCTCTXGCTXAAXTXTCXXXTXAGAAACTXTTGTGTC	720
730.....740.....750.....760.....770.....780	
PPI-AtCPP	GATGGATCTACAAGGTCAAGCCATAGCAATGCTTACATGTTAGGTTCTTAAGAACAA	780
PPI-BnCPP	GATGGATCTACAAGGTCAAGCCATAGTAATGCTTACATGTTAGGTTCTTCAGAACAA	780
PPI-SoyCPP	GATGGATCTACAAGGTCAAGCCATAGTAATGTTAGGATCTTCAGAACAA	780
Consensus	GATGGATCTACAAGGTCAAGXXATGCTXAGXAAATGCTXAGTGTATGGXTTCTTXAGAACAA	780
790.....800.....810.....820.....830.....840	
PPI-AtCPP	AGGATTGTTCTTATGATACGTTGATTCAGCAGTGCAGAAGAATGAGGATGAAATTGTC	840
PPI-BnCPP	AGGATTGTTCTTATGACACATTGATTCAGCAGTGCAGAATGAGAATGAAATTGTC	840
PPI-SoyCPP	AGGATTGTCCTTATGACACATTGATTCAGCAGTGCAGAAGAATGAGGAAATTGTC	840

Table 24. ClustalW Analysis of PPI Amino Acids

- 1) PPI-AtCPP (SEQ ID NO:2)
 - 2) PPI-BnCPP (SEQ ID NO:15)
 - 3) PPI-SoyCPP (SEQ ID NO:18)
 - 4) Consensus (SEQ ID NO:73)

PPI-AtCPP	HFHFVHEFVTIVTDSTILYFGVLPWFWKKSGDFMTIAGFNAENEILHTLAFLAGLMIWSQ	120
PPI-BnCPP	HFHFVHEFVTILEMDSAILEFGCILPWFWKISGGFLPMVGLDEEENILHTLSFLAGLMIWSC	120
PPI-SoyCPP	HFHFVHEFVTIVTDSTILYFGVLPWFWKKSGDFMTIAGFNAENEILHTLAFLAGLMIWSQ	120
Consensus PPI	HFHFVHEFVTIXXDSXILXFGXLWPWFWXSGXXXXXXXXXXXXENEILHTLXFLAGLMXWSQ	120
	130 140 150 160 170 180	
PPI-AtCPP	ITDLPFSLYSTFVIEARHGFNKQTIPWLFERDMLKGIFLSVIIGPPIVAAIIIVIVVKGGPY	180
PPI-BnCPP	ITDLPFSLYSTFVIESTRHGFNKQTILWFLIRDMKGIILSVIPAPPIVAAIIIVIVVKGGPY	180
PPI-SoyCPP	ITDLPFSLYSTFVIEARHGFNKQTIPWLFERDMLKGIFLSVIIGPPIVAAIIIVIVVKGGPY	180
Consensus PPI	ITDLPFSLYSTFVIEXRHGFNKQTXXWFXRDMXKGIXLSVIXXPPIVAAIIIVIVVKGGPY	180
	190 200 210 220 230 240	
PPI-AtCPP	LAIYLWVFTFGLSIVMMTLYPVLIAPLFNKFTPPLPDGQLREKIEKLASSLNYPPLKKLFVV	240
PPI-BnCPP	LAIYLWFLMFIILSIVMMTLYPVLIAPLFNKFTPPLPDGDLREKIEKLASSLKPLKKLFVV	240
PPI-SoyCPP	LAIYLWVFTFGLSIVMMTLYPVLIAPLFNKFTPPLPDGQLREKIEKLASSLNYPPLKKLFVV	240
Consensus PPI	LAIYLWFLMFIILSIVMMTLYPVLIAPLFNKFTPPLPDGDLREKIEKLASSLKPLKKLFVV	240
	250 260 270 280 290 300	
PPI-AtCPP	DGSTRSSHSNAYMYGFFKNKRIVPYDTLIQQCKDDEEIVAVIAHELGHWKLHNHTVYTFVA	300
PPI-BnCPP	DGSTRSSHSNAYMYGFFKNKRIVYDTLIQQCQNEEIVAVIAHELGHWKLHNHTYSEAIA	300
PPI-SoyCPP	DGSTRSSHSNAYMYGFFKNKRIVYDTLIQQCKDDEEIVAVIAHELGHWKLHNHTVYTFVA	300
Consensus PPI	DGSTRSSHSNAYMYGFFKNKRIVYDTLIQQCXXXXEIVAVIAHELGHWKLHNHTYSEAIA	300
	310 320 330 340 350 360	
PPI-AtCPP	MQILTLLQFGGYTLVRNSADLYRSFGFDTQPVLIGLIIIFQHTVIPPLQQLVSFGNLNVRS	360
PPI-BnCPP	MQILAFQFGGYTLVRNSTDLERSFGFDTQPVLIGLIIIFQHTVIPPLQQLVSFGNLNVRS	360
PPI-SoyCPP	MQILTLLQFGGYTLVRNSADLYRSFGFDTQPVLIGLIIIFQHTVIPPLQQLVSFGNLNVRS	360
Consensus PPI	XQILXXQFGGYTLVRNSXDLERSFGFDTQPVLIGLIIIFQHTVIPPLQXLVSFXNLNVRS	360
	370 380 390 400 410 420	
PPI-AtCPP	FEFQADGFAKKLGYASGLRGGLVQLQEENLSAMNTDPWYSAYHYSHPPLVERLAALDEPD	420
PPI-BnCPP	FEFQADFAVNLYGAKDLFPALVQLQEENLSAMNTDPWYSAYHYSHPPLVERLRAALGED	420
PPI-SoyCPP	FEFQADGFAKKLGYASGLRGGLVQLQEENLSAMNTDPWYSAYHYSHPPLVERLAALDEPD	420
Consensus PPI	FEFQADFAXXXLYGAXXLFXXLVQLQEENLSAMNTDPWYSAYHYSHPPLVERLXXDXD	420
	430 440 450 460 470 480	
PPI-AtCPP	KKED-----	424
PPI-BnCPP	KKTD-----	424
PPI-SoyCPP	KKED-----	424
Consensus PPI	KKXD-----	480

Table XX. ClustalW Analysis of PPI/Generic Nucleic Acids

- 1) **PPI-AtCPP** (SEQ ID NO:1)
- 2) **PPI-BnCPP** (SEQ ID NO:14)
- 3) **PPI-SoyCPP** (SEQ ID NO:17)
- 4) **afc1** (SEQ ID NO:29)
- 5) **AT4g01320** (SEQ ID NO:31)
- 6) **AF007269** (SEQ ID NO:33)
- 6) **Consensus** (SEQ ID NO:75)

	10 20 30 40 50 60 70	
PPI-AtCPP NA	-----	
PPI-BnCPP	-----	
PPI-SoyCPP	-----	
afc1	-----	
AT4g01320	-----	

PPI-SoyCPP	TTTCCTTCTGTAATAATTGGTCCACCTATTGGCTGCAA-----TCATGGTAA-TAGTACAGAAAGG
afc1	-----
AT4g01320	-----
AF007269	CTCTTCACTGCTCCAATGTTCCATCAGTAGTCAGCACAAAGAGATCTTTATCTGGTGATCAAAA
Consensus	TTCCCTCTGTATA-----G--CC-CCTATTGTG-CTGCAA-----T-ATTGTA--TAGT-CAGAAAGG
	640 650 660 670 680 690 700
PPI-AtCPP NA	AGGTCCATA--CTTGGCCATCTATCTTGGGTTTACGTTGGCTTCTATTGTGATGATGACCCTT
PPI-BnCPP	-----
PPI-SoyCPP	AGGTCCATA--CTTGGCCATCTATCTTGGGTTTACGTTGGCTTCTATTGTGATGATGACCCTT
afc1	-----
AT4g01320	-----
AF007269	AAGTAGATGATGTTATTGAATTTCAGTGTGGAGATCTGTGTTGGCATTAGAGTAGATTGAT
Consensus	AGGTCC----TATG-CCATCTATCTGGG-----TTAGTTCTCTGTGATGATGACC--TT
	710 720 730 740 750 760 770
PPI-AtCPP NA	ATCCACTACTA-ATAGCTCACTCTCAATAAGTTCACTCCACT--TCCAGATGGTCAACTCAGGGAGAA
PPI-BnCPP	GGCGATTCCT--TTCATGGAAACCGTCGTTGGTTTATGATAGTGTACGTTTGAGACGTATTG
PPI-SoyCPP	ATCCACTACTA-ATAGCTCACTCTCAATAAGTTCACTCCACT--TCCAGATGGTCAACTCAGGGAGAA
afc1	GGCGATTCCT--TTCATGGAAACCGTCGTTGGTTTATGATAGTGTACGTTTGAGACGTATTG
AT4g01320	GGCGATTCCT--TTCATGGAAACCGTCGTTGGTTTATGATAGTGTACGTTTGAGACGTATTG
AF007269	TTCATCTCTGTTTATTCTTACAGTTTATGATAGTGTACGTTTGAGACGTATTG
Consensus	A-CC-----GTTATGCCCTCTCAA-AAGTTCACTCC-CT--TCCGATGG--ACTC-GGGAGAA
	780 790 800 810 820 830 840
PPI-AtCPP NA	AATCGAGA-AACTTGCTTCCCTCCT-CAACTATCCGTTAAAGAAACTATTGTTGTCGATGGATCCACAA
PPI-BnCPP	ATCTGAGGCAACATACTGCTCTCAAGCTTCCCACCTCCCCAAAGACT-TTGGTTGAG-TCATTAGCCAA
PPI-SoyCPP	AATCGAGA-AACTTGCTTCCCTCCT-CAACTATCCGTTAAAGAAACTATTGTTGTCGATGGATCCACAA
afc1	ATCTGAGGCAACATACTGCTCTCAAGCTTCCCACCTCCCCAAAGACC-TTGGTTGAG-TCATTAGCCAA
AT4g01320	ATCTGAGGCAACACTCACTGCTCTCAAGCTTCCCACCTCCCCAAAGACC-TTGGTTGAG-TCATTAGCCAA
AF007269	ATCTGAGGCAACACTCACTGCTCTCAAGCTTCCCACCTCCCCAAAGACC-TTGGTTGAG-TCATTAGCCAA
Consensus	A---TGAGA-AACTTGCTTCTCCT----A---ATCCTTAAG--AACTATTGTTGTCGATGGATC-ACAA
	850 860 870 880 890 900 910
PPI-AtCPP NA	GATCAAGTCACAGCAATGCCTATATGATGGATTCTCAAGAACAGAGGATTGTC--CTTATGACAC
PPI-BnCPP	G-AGAACTTGGAGAAATCTCGA-GCTTACAG
PPI-SoyCPP	GATCAAGTCACAGCAATGCCTATATGATGGATTCTCAAGAACAGAGGATTGTC--CTTATGACAC
afc1	G-AGAACTTGGAGAAATCACGA-GCATACAG
AT4g01320	G-AGAACTTGGAGAAATCACGA-GCATACAG
AF007269	G-AGAACTTGGAGAAATCACGA-GCATACAG
Consensus	G-TCAAG-CATAG-AATGCTAA-TGTATGG--TTCTTAAGAACAA-AGGATTGTC---TTATGACAC
	920 930 940 950 960 970 980
PPI-AtCPP NA	ATTAATTC-----AACAGTGCAGAACAC-GATGAGGAAATTGTTG-CCTTATTGCCCATGAG
PPI-BnCPP	-----TCTTGACAAA--AGCCATTTCACTTG
PPI-SoyCPP	ATTAATTC-----AACAGTGCAGAACAC-GATGAGGAAATTGTTG-CCTTATTGCCCATGAG
afc1	-----TCTTGACAAA--AGCTATTTCACTTG
AT4g01320	-----GGATATCATCACTGAGACCTTAAATATGCAACCTTTTCACTTTG
AF007269	TTTAGTTTTATAATTGCCAGGGATATCATCACTGAGACCTTAAATATGCAACCTTTTCACTTTG
Consensus	ATTATTC-----ACAGTGCAGA-----GAAGAAATTGTTG--CGTTATTGCC---AGA
	990 1000 1010 1020 1030 1040 1050
PPI-AtCPP NA	TTGGGACACTGGAAAGCTCAACCAACTGTGTACACATTGTTGCTATGCAGATCTTACACTTCTACAAAT
PPI-BnCPP	TTCATGAGTTGTTACTATA-CTTATGGACTCTGGAT-TCTGTTCTTGGGATCTTGC-----CTTGGTT
PPI-SoyCPP	TTGGGACACTGGAAAGCTCAACCAACTGTGTACACATTGTTGCTATGCAGATCTTACACTTCTACAAAT
afc1	TTCATGAGTTGTTACTATA-CTTATGGACTCTGGAT-TCTGTTCTTGGGATCTTGC-----CTTGGTT
AT4g01320	TTCATGAGTTGTTACTATA-CTTATGGACTCTGGAT-TCTGTTCTTGGGATCTTGC-----CTTGGTT
AF007269	GTGGGACACTGGAA-----CTAACACTTACACATT-ATTGTTTC-----AATCTT-----CTTTACAAT
Consensus	
	1060 1070 1080 1090 1100 1110 1120
PPI-AtCPP NA	TTGGAGGATATACACTAGTGCAGAACATTGCTGATCTGTATCGAAGCTTGGGTTGATACGCAGCCAGT
PPI-BnCPP	TTGGAAAG-
PPI-SoyCPP	TTGGAGGATATACACTAGTGCAGAACATTGCTGATCTGTATCGAAGCTTGGGTTGATACGCAGCCAGT
afc1	TTGGAAAG-
AT4g01320	TTGGAAAGGACATATCTGGTTTCGGTATACAGTATCT-CATTGAAATATAGAGTTGACATTACAA--
AF007269	TTGGAGGATACAC-CTAGTG--AAATCC---TGATCT---TGAG---TTGGTTGATAC-CAGCCG--
Consensus	
	1130 1140 1150 1160 1170 1180 1190

	1200	1210	1220	1230	1240	1250	1260
PPI-AtCPP NA	AACCTAGTCAGCCGATCATTTGAATTTCAGGCTGATGGCCTTGCCAAAGAAGCTGGATATGCATCTGGAT	AATGAAAATCCTGCACACTCTTCAATTCTGGCTGGTC-TTATGACATGGTCACAG	AACCTAGTCAGCCGATCATTTGAATTTCAGGCTGATGGCCTTGCCAAAGAAGCTGGATATGCATCTGGAT	AATGAAAATCCTGCACACTCTTCAATTCTGGCTGGTG-TTATGACATGGTCACAG	AATGAAAATCCTGCACACTCTTCAATTCTGGCTGGTG-TTATGACATGGTCACAG	AACCTG-----TAGCGACTTTGATTTCAGGCTGATG-CITTCG-----GAAGCTGG-TATGCAGTCGG-----	
PPI-BnCPP							
PPI-SoyCPP							
afc1							
AT4g01320							
AF007269							
Consensus							
	1270	1280	1290	1300	1310	1320	1330
PPI-AtCPP NA	TACGCGGTG--GTCTTGTGAAACTACAGGAGGAGAACCTGTCAGCT--ATGAATACAGATCCTGGTA						
PPI-BnCPP							
PPI-SoyCPP							
afc1							
AT4g01320							
AF007269							
Consensus	CCCTTCATATAGTCTATACTCGTTAGCATCAAATATCTATTTCTTAAGATAATAATTCTTTATA	--T-----GTCTAGTGAA-CTACAGGAGAGAA---TGTCA-----ATGAA-ACAGATCCTTG-TA					
	1340	1350	1360	1370	1380	1390	1400
PPI-AtCPP NA	CTCT---GCTTATCACTATCTCATCCCTCCCT-----TGTTGAAAGATTGCCGCGCTGGACGAA						
PPI-BnCPP	--ATCACTGATTGCCATTTCCTTGT-----TACTCAACTTTCG-----TGATCGAG-----						
PPI-SoyCPP	CTCT---GCTTATCACTATCTCATCCCTCCCT-----TGTTGAAAGATTGCCGCGCTGGACGAA						
afc1	--ATCACTGATTGCCATTTCCTTGT-----TACTCAACTTTCG-----TGATCGAG-----						
AT4g01320	--ATCACTGATTGCCATTTCCTTGT-----TACTCAACTTTCG-----TGATCGAG-----						
AF007269	--TCTGATGCAGATCACTGATTGCCATTTCCTTGT-----TACTCAACTTTCG-----TGATCGAG-----						
Consensus	CTC---GCTTATCACTATCCACCTCCCTTGAAAGATGCTGAGAGAAAGAGATAATCTAAATTCT						
	1410	1420	1430	1440	1450	1460	1470
PPI-AtCPP NA	--CCGGATAAGAGGAAGACTAAE						
PPI-BnCPP	--TCTCGGCATGGGTTCAACAAA						
PPI-SoyCPP	--CCGGATAAGAGGAAGACTAA						
afc1	--TCTCGGCATGGGTTCAACAAA						
AT4g01320	--TCTCGGCATGGGTTCAACAAA						
AF007269	--TCTCGGCATGGGTTCAACAAA						
Consensus	TTCCCTTTTCATGGAGGTAACAAA						
	1480	1490	1500	1510	1520	1530	1540
PPI-AtCPP NA							
PPI-BnCPP							
PPI-SoyCPP							
afc1							
AT4g01320							
AF007269							
Consensus	GAGATGTGGATTAAATTGCTTCTAAATTCTGTTGACAG						
	GAGATGTGGATTAAATTGCTTCTAAATTCTGTTGACAG						
	GAGATGTGGATTAAATTGCTTCTAAATTCTGTTGACAG						
	1550	1560	1570	1580	1590	1600	1610
PPI-AtCPP NA	TCAAAGGAATCTCCTCTGTCACTACCTGCCCTCCTATCGTTGCCGAATTATTGTTATAGTCAG-----						
PPI-BnCPP							
PPI-SoyCPP							
afc1							
AT4g01320	TCAAAGGAATCTCCTCTGTCACTACCTGCCCTCCTATCGTTGCCGAATTATTGTTATAGTCAG-----						
AF007269	TCAAAGGAATCTCCTCTGTCACTACCTGCCCTCCTATCGTTGCCGAATTATTGTTATAGTCAG-----						
Consensus	TCAAAGGAATCTCCTCTGTCACTACCTGCCCTCCTATCGTTGCCGAATTATTGTTATAGTCAG-----						
	1620	1630	1640	1650	1660	1670	1680
PPI-AtCPP NA							
PPI-BnCPP							
PPI-SoyCPP							
afc1							
AT4g01320							
AF007269							
Consensus	TTGATGATTCTGGATTCACTTATTTCTGAGTTTCACTGGATGACTATTCTCCATTGAGTGTGAGCT						

PPI-AtCPP NA	AGTAAGCTTGACCTCACACCTGTTAGTCGAGCGTTGAGTTTCAGG-----
PPI-BnCPP	-----
PPI-SoyCPP	-----
afc1	AGTAAGCTTGCCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----
AT4g01320	AGTAAGCTTGCCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----
AF007269	AGTAAGCTTGCCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----TACCATCTTACAATCCCTCAAGA
Consensus	AGTAAGCTTGCCCTGAACCTCGTTAGTCGAGCGTTGAGTTTCAGG-----TACCATCTTACAATCCCTCAAGA
	2810 2820 2830 2840 2850 2860 2870
PPI-AtCPP NA
PPI-BnCPP	-----
PPI-SoyCPP	-----
afc1	-----
AT4g01320	-----
AF007269	TCCAACCATAGTTCTTATTGCAATGGCAGCCTCATCTACTAATCTGAGTTAACGTTCTTTGCAGGC-----
Consensus	TCCAACCATAGTTCTTATTGCAATGGCAGCCTCATCTACTAATCTGAGTTAACGTTCTTTGCAGGC-----
	2880 2890 2900 2910 2920 2930 2940
PPI-AtCPP NA
PPI-BnCPP	TGATGCTTTGCAGTGAATCTTGGT-----
PPI-SoyCPP	TATGCAAAGGATCTACGTCCTGCCCTAGTGAAGCTACAGG-----
afc1	TGATGCTTTGCAGTGAAGCTTGGCTATGAAAAGATCTTCGTCCTGCTCTAGTGAAGACTACAGG-----
AT4g01320	TGATGCTTTGCAGTGAAGCTTGGCTATGAAAAGATCTTCGTCCTGCTCTAGTGAAGACTACAGGTCAGA-----
AF007269	TGATGCTTTGCAGTGAAGCTTGGCTATGAAAAGATCTTCGTCCTGCTCTAGTGAAGACTACAGGTCAAGA-----
Consensus	TGATGCTTTGCAGTGAAGCTTGGCTATGAAAAGATCTTCGTCCTGCTCTAGTGAAGACTACAGGTCAGA-----
	2950 2960 2970 2980 2990 3000 3010
PPI-AtCPP NA
PPI-BnCPP	-----
PPI-SoyCPP	-----
afc1	-----
AT4g01320	GAAGATAACAACAGAACACAAACTGTTACCTCAATTGAGTCACACACTTAAATGGATTTGTTGGGA-----
AF007269	GAAGATAACAACAGAACACAAACTGTTACCTCAATTGAGTCACACACTTAAATGGATTTGTTGGGA-----
Consensus	GAAGATAACAACAGAACACAAACTGTTACCTCAATTGAGTCACACACTTAAATGGATTTGTTGGGA-----
	3020 3030 3040 3050 3060 3070 3080
PPI-AtCPP NA
PPI-BnCPP	-----A-----
PPI-SoyCPP	-----AGAGAACTTATCAGG-----
afc1	-----AGAGAACTTATCAGG-----A-----GATGAACACAGACCCATTG-----TACTCAGCTTATCACTACTCACACCCC-----
AT4g01320	-----A-----AGAGAACTTATCAGG-----A-----GATCCATTG-----C-----ACTCAGCTTATCACTACTCACATCC-----
AF007269	-----TTTGCAAGGA-----AGAGAACTTATCAGG-----A-----GATCCATTG-----T-----ACTCAGCTTATCACTACTCACATCC-----
Consensus	-----TTTGCAAGGA-----AGAGAACTTATCAGG-----A-----GATCCATTG-----T-----ACTCAGCTTATCACTACTCACATCC-----
	3090 3100 3110 3120 3130
PPI-AtCPP NA
PPI-BnCPP	TCCTCTTGTAGAGAGGCTTCGAGCATTGATGGAGAAGACAAGAAGACAGATTAA-----
PPI-SoyCPP	-----
afc1	TCCTCTTGTGAAAGGCTTCGAGCATTGATGGAGAAGACAAGAAGACAGATTAA-----
AT4g01320	TCCTCTTGTGAAAGGCTTCGAGCATTGATGGAGAAGACAAGAAGACAGATTAA-----
AF007269	TCCTCTTGTGAAAGGCTTCGAGCATTGATGGAGAAGACAAGAAGACAGATTAA-----
Consensus	TCCTCTTGTGAAAGGCTTCGAGCATTGATGGAGAAGACAAGAAGACAGATTAA-----

Table XX. ClustalW Analysis of PPI/Generic Nucleic Acids

- 1) **PPI-AtCPP** (SEQ ID NO:1)
- 2) **PPI-BnCPP** (SEQ ID NO:14)
- 3) **PPI-SoyCPP** (SEQ ID NO:17)
- 4) **afc1** (SEQ ID NO:29)
- 5) **AT4g01320** (SEQ ID NO:31)
- 6) **AF007269** (SEQ ID NO:33)
- 6) **Consensus** (SEQ ID NO:75)

PPI-AtCPP NA
PPI-BnCPP	-----
PPI-SoyCPP	-----
afc1	-----
AT4g01320	-----
AF007269	ATGGCGATTCTTTCATGGAAACCGCTGGTAAGCTTCAAAACCTTTCTGAGACATTTACTATCC-----

Table 26. ClustalW Analysis of PPI/Generic Amino Acids

1) PPI-AtCPP	(SEQ ID NO:2)
2) PPI-BnCPP	(SEQ ID NO:15)
3) PPI-SoyCPP	(SEQ ID NO:18)
4) afc1	(SEQ ID NO:30)
5) AT4g01320	(SEQ ID NO:32)
6) AF007269	(SEQ ID NO:34)
7) Consensus Gener	(SEQ ID NO:74)
	10 20 30 40 50 60
PPI-AtCPP	MAIPFVMEAVVGFMI MYI FETYLD V RQHRAKLPLTPKTL E GVISQEKF E KSRAYS--LD
PPI-BnCPP	MAIPFMETVVVGFMIVM Y I F ETYLDLRLC H TALKLPLTPKTL V GVISQEKF E KSRAYS--LD
PPI-SoyCPP	MAIPFVMEAVVGFMI VMYI FETYLDLRLC H TALKLPLTPKTL V GVISQEKF E KSRAYS--LD
afc1	MAIPFMETVVVGFMIVM YI FETYLDLRLC H TALKLPLTPKTL V GVISQEKF E KSRAYS--LD
AT4g01320	MAIPFMETVVVGFMIVM YI FETYLDLRLC H TALKLPLTPKTL V GVISQEKF E KSRAYRDI T
AF007269	MAIPFMETVVVGFMIVM YI FETYLDLRLC H TALKLPLTPKTL V GVISQEKF E KSRAYRDI T
Consensus Gener	MAIPFVMEAVVGFMI MYI FETYLD X RQXXALKLPLTPKTL V XXXXXXXXXXXXXXXXXXXXXX 60
	70 80 90 100 110 120
PPI-AtCPP	KS-----HFHFVHEFVTI V DST T ILYFGVLPWFWRKSGDFMTIA G FAENEILHTLAFLA
PPI-BnCPP	KS-----HFHFVHEFVTI M DS A ILEFG G ILPWFWR I SCGF L PMVG L DPENEILHTLSFLA
PPI-SoyCPP	KS-----HFHFVHEFVTI V DST T ILYFGVLPWFWRKSGDFMTIA G FAENEILHTLAFLA
afc1	KS-----YFHFVHEFVTI M DS A ILEFG G ILPWFWRMSGAV P RLGLDPENEILHTLSFLA
AT4g01320	ENFNICSYFHFVHEFVTI M DS A ILEFG G ILPWFWRMSGAV P RLGLDPENEILHTLSFLA
AF007269	-----TDLPLS Y STFVIESRHF N KQT I WMFIRDMIKGT F LSVI G PP I VAAIFI
Consensus Gener	XX 120
	130 140 150 160 170 180
PPI-AtCPP	GENIWSQITDLPFLS Y STFVIE R HGF N KQT P W G F F RDN M KG I FLSVI G PP I VAAIFI
PPI-BnCPP	GENIWSQITDLPFLS Y STFVIE S RHF G FK N KQT I WMFIRDMIKG I LLS V I P APP I VAAIFI
PPI-SoyCPP	GENIWSQITDLPFLS Y STFVIE R HGF N KQT P W G F F RDN M KG I FLSVI G PP I VAAIFI
afc1	GW I WSQITDLPFLS Y STFVIE S RHF G FK N KQT I WMFIRDMIKGT F LSVI G PP I VAAIFI
AT4g01320	GW I WSQITDLPFLS Y STFVIE S RHF G FK N KQT I WMFIRDMIKGT F LSVI G PP I VAAIFI
AF007269	-----TDLPLS Y STFVIESRHF N KQT I WMFIRDMIKGT F LSVI G PP I VAAIFI
Consensus Gener	XXXXXXXXTDLPLS Y STFVIE R HGF N KQT X W X X RDN M KG XX LS V I X X PP I VAAIFI 180
	190 200 210 220 230 240
PPI-AtCPP	VQKGGPYLAIYLWVFT F GLS E VMM T Y P V L I A PL F N K FTPLPDG Q LREKIEK K ASSLN Y P
PPI-BnCPP	VQKGGPYLAIYLWAFMF I LSLVMM T Y P V L I A PL F N K FTPLPDG Q LREKIEK K ASSL K FP
PPI-SoyCPP	VQKGGPYLAIYLWAFMF I LSLVMM T Y P V L I A PL F N K FTPLPDG Q LREKIEK K ASSL N Y P
afc1	VQKGGPYLAIYLWAFMF I LSLVMM T Y P V L I A PL F N K FTPLPDG Q LREKIEK K ASSL K FP
AT4g01320	VQKGGPYLAIYLWAFMF I LSLVMM T Y P V L I A PL F N K FTPLPDG Q LREKIEK K ASSL K FP
AF007269	VQKGGPYLAIYLWAFMF I LSLVMM T Y P V L I A PL F N K FTPLPDG Q LREKIEK K ASSL K FP
Consensus Gener	VQKGGPYLAIYLW X FX X LS X VMM T Y P V L I A PL F N K FTPLPDG Q LREKIEK K ASSL XX P
	250 260 270 280 290 300
PPI-AtCPP	LKKLFVVDGSTRSSHSNAYMGFFKNKRIV P YDTL Q QQC DDE EIVAVIAHELGHWKLNH
PPI-BnCPP	LKKLFVVDGSTRSSHSNAYMGFFKNKRIV L YDTL Q QQC EN EIVAVIAHELGHWKLNH
PPI-SoyCPP	LKKLFVVDGSTRSSHSNAYMGFFKNKRIV P YDTL Q QQC DDE EIVAVIAHELGHWKLNH
afc1	LKKLFVVDGSTRSSHSNAYMGFFKNKRIV L YDTL Q QQC NE EIVAVIAHELGHWKLNH
AT4g01320	LKKLFVVDGSTRSSHSNAYMGFFKNKRIV L YDTL Q QQC DE EIVAVIAHELGHWKLNH
AF007269	LKKLFVVDGSTRSSHSNAYMGFFKNKRIV L YDTL Q QQC ND EIVAVIAHELGHWKLNH
Consensus Gener	LKKLFVVDGSTRSSHSNAYMGFFKNKRIV X YDTL Q QQC XXXX EIVAVIAHELGHWKLNH 300
	310 320 330 340 350 360
PPI-AtCPP	TTYSFIA Q IL L TLQF G GGY T LRV N SAD L YRSFG F DTQPVLIGLI I FQHTV I PLQ C LVS F G
PPI-BnCPP	TTYSFIA Q IL L TLQF G GGY T LRV N STD L YRSFG F DTQPVLIGLI I FQHTV I PLQ H LVS F D
PPI-SoyCPP	TTYSFIA Q IL L TLQF G GGY T LRV N STD L YRSFG F DTQPVLIGLI I FQHTV I PLQ C LVS F G
afc1	TTYSFIA Q IL L TLQF G GGY T LRV N STD L YRSFG F DTQPVLIGLI I FQHTV I PLQ H LVS F G
AT4g01320	TTYSFIA Q IL L TLQF G GGY T LRV N STD L YRSFG F DTQPVLIGLI I FQHTV I PLQ H LVS F G
AF007269	TTYSFIA Q IL L TLQF G GGY T LRV N STD L YRSFG F DTQPVLIGLI I FQHTV I PLQ H LVS F G
Consensus Gener	TTYSFIA Q IL L TLQF G GGY T LRV N STD L YRSFG F DTQPVLIGLI I FQHTV I PLQ H LVS F G

What is claimed is:

1. A method of producing a transgenic plant, comprising introducing into a plant cell a compound that increases prenyl protease expression or activity to generate a transgenic cell; and regenerating a transgenic plant from said transgenic cell.
2. The method of claim 1, wherein said plant has an altered phenotype selected from the group consisting of increased tolerance to stress, delayed senescence, increased ABA sensitivity, increased yield, increased productivity and increased biomass compared to a wild type plant.
3. The method of claim 1, wherein said compound comprises a nucleic acid sequence encoding prenyl protease.
4. The method of claim 3, wherein said nucleic acid comprises SEQ ID NO: 1, 14, 17, 68, 70, 72, 74, 21, 23, 25, 27, 29, 31, or 33.
5. The method of claim 3, wherein said nucleic acid is operably linked to a promotor.
6. The method of claim 5, wherein said promoter is selected from the group consisting of a constitutive promoter, an ABA inducible promoter, tissue specific promoters or a guard cell-specific promoter
7. The method of claim 1, wherein said compound is a prenyl protease polypeptide or fragment thereof.
8. The method of claim 7, wherein said prenyl protease polypeptide comprises the amino acid sequence of SEQ ID NO: 2, 15, 18, 22, 24, 26, 28, 30, 32, 34, 69, 71, 73, or 75.
9. The transgenic plant produced by claim 1.

10. The seed produced by the transgenic plant of claim 9, wherein said seed produces a plant that has an altered phenotype selected from the group consisting of increased tolerance to stress, delayed senescence, increased ABA sensitivity, increased yield, increased productivity and increased biomass compared to a wild type plant.
11. A method of producing a transgenic plant, comprising introducing into a plant cell a nucleic acid that inhibits prenyl protease expression or activity to generate a transgenic cell; and regenerating a transgenic plant from said transgenic cell.
12. The method of claim 11, wherein said plant has an altered phenotype selected from the group consisting of increased tolerance to stress, delayed senescence, increased ABA sensitivity, increased yield, increased productivity and increased biomass compared to a wild type plant.
13. The method of claim 11, wherein said nucleic acid comprises an antisense nucleic acid sequence encoding prenyl protease.
14. The method of claim 13, wherein said antisense nucleic acid comprises 20 or more consecutive nucleic acids complementary to SEQ ID NO: 1, 14, 17, 21, 23, 25, 27, 29, 31, 33, 68, 70, 72, or 74.
15. The method of claim 13, wherein said antisense nucleic acid comprises SEQ ID NO: 16, 19, 20, 5, 35, 37, 38, 42, 43, 45, 46, 48, 49, 51, or 52.
16. The method of claim 11, wherein said nucleic acid is operably linked to a promotor.
17. The method of claim 16, wherein said promoter is selected from the group consisting of a constitutive promoter, an ABA inducible promoter, tissue specific promoters or a guard cell-specific promoter
18. The method of claim 11, wherein the nucleic acid is an inhibitor of farnesylation.

- 19 The transgenic plant produced by any one of the methods of claims 11.
20. The seed produced by the transgenic plant of claim 19, wherein said seed produces a plant that has an altered phenotype selected from the group consisting of increased tolerance to stress, delayed senescence, increased ABA sensitivity, increased yield, increased productivity and increased biomass compared to a wild type plant.
21. A method of producing a transgenic plant, comprising introducing into a plant cell a nucleic acid selected from the group consisting of SEQ ID NO: 16, 19, 20, 5, 35, 37, 38, 42, 43, 45 , 46, 48, 49, 51, and 52 to generate a transgenic cell; and regenerating a transgenic plant from said transgenic cell.
22. An isolated polypeptide comprising the mature form of an amino acid sequenced selected from the group consisting of SEQ ID NO: 2, 15, 18, 69, 71, 73 and 75.
- 23 An isolated polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 2, 15, 18, 69, 71, 73 and 75.
24. An isolated polypeptide comprising an amino acid sequence which is at least 96% identical to an amino acid sequence selected from the group consisting of SEQ ID NO: 15
25. An isolated polypeptide comprising an amino acid sequence which is at least 99% identical to an amino acid sequence selected from the group consisting of SEQ ID NO: 2, 15, 18
- 26 The polypeptide of claim 24, wherein said polypeptide has prenyl pretease activity.
- 27 An isolated polypeptide, wherein the polypeptide comprises an amino acid sequence comprising one or more conservative substitutions in the amino acid sequence selected from the group consisting of SEQ ID NO: 2, 15, and 18.

28. The polypeptide of claim 23, wherein said polypeptide is naturally occurring.
29. An isolated nucleic acid molecule comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO: 1, 14, 17, 20, 16, 19, 68, 70, 72, and 74.
30. The nucleic acid molecule of claim 29, wherein the nucleic acid molecule is naturally occurring.
31. An isolated nucleic acid molecule encoding the mature form of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO: SEQ ID NO: 2, 15, and 18.
32. An isolated nucleic acid molecule, wherein said nucleic acid molecule hybridizes under stringent conditions to the nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 14, 17, 20, 16, 19, 68, 70, 72, and 74.
33. An isolated nucleic acid molecule comprising a nucleotide sequence which is at least 94% identical to the nucleotide sequence selected from the group consisting of SEQ ID NO: 17 ,18 and 19.
34. An isolated nucleic acid molecule comprising a nucleotide sequence which is at least 99% identical to the nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 14, 17, 20, 16, 19.
35. A vector comprising the nucleic acid molecule of claim 29.
36. The vector of claim 35, further comprising a promoter operably linked to said nucleic acid molecule.
37. A cell comprising the vector of claim 36.

38. An antibody that immunospecifically binds to the polypeptide of claim 22.
39. The antibody of claim 38, wherein the antibody is a monoclonal antibody.
40. The antibody of claim 37, wherein the antibody is a polyclonal antibody.
41. A method of identifying an agent that binds to the polypeptide of claim 27, the method comprising:
 - (a) introducing said polypeptide to said agent; and
 - (b) determining whether said agent binds to said polypeptide.
42. The method of claim 41, wherein the agent is a farnesylation inhibitor.
43. A method for identifying farnesylation modulator, the method comprising:
 - (a) providing a cell expressing the polypeptide of claim 22;
 - (b) contacting the cell with a candidate substance; and
 - (c) determining whether the substance alters farnesylation activity;
whereby, if an alteration observed in the presence of the substance is not observed when the cell is contacted with a composition in the absence of the substance, the substance is identified as a farnesylation modulator.
44. A method for identifying an interacting gene of prenyl protease, the method comprising:
 - a) providing the transgenic plant of claim 1;
 - b) creating a library of mutagenized plants from (a);
 - c) determining whether the mutagenized plant contains an altered phenotype;

whereby, the mutagenized plant has altered the function of an interacting gene of prenyl protease which results in an altered phenotype from the transgenic plant of (a) to that of a wild type non-transgenic plant.

45. A plant, wherein a mutation has been introduced in the gene encoding prenyl protease, resulting in said plant displaying altered prenyl protease activity and an increased tolerance to stress as compared to a wild type plant.

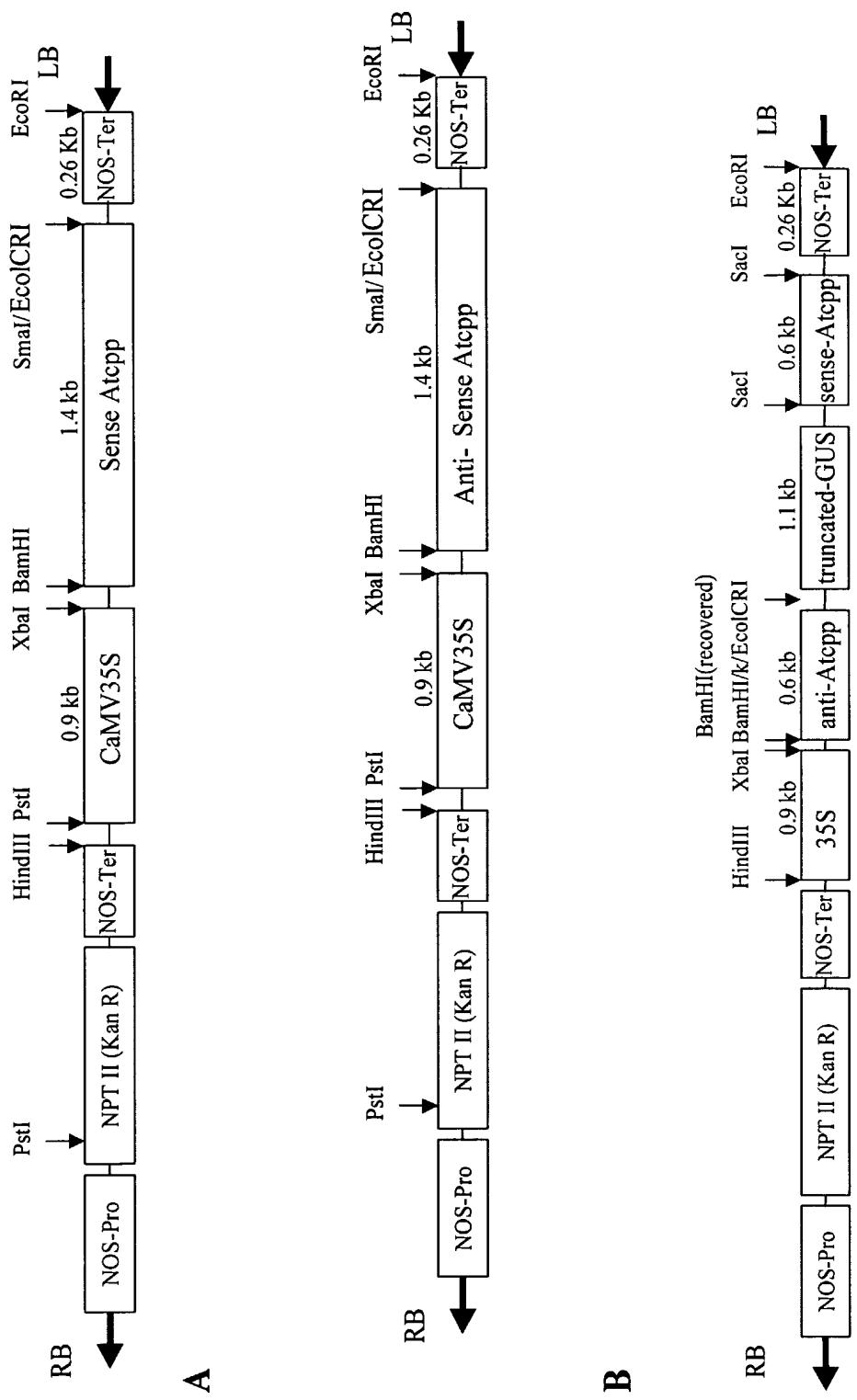


Figure 1.

A

Nucleic Acid	PPI-AtCPP	PPI-BnCPP	PPI-SoyCPP	BASF-AT1	BASF-AT2	BASF-Corn	BASF-Soy	AFC1	AT4g01320	AF007269
PPI-AtCPP	X									
PPI-BnCPP	92	X								
PPI-SoyCPP	76	77	X							
BASF-AT1	98	93	76	X						
BASF-AT2	99	93	76	99	X					
BASF-Corn	57	57	57	57	57	X				
BASF-Soy	72	72	93	72	72	52	X			
AFC1	99	93	77	99	99	57	72	X		
AT4g01320	99	92	70	99	99	50	64	99	X	
AF007269	97	91	10	97	97	13	8	97	97	X

B

Amino Acid	PPI-AtCPP	PPI-BnCPP	PPI-SoyCPP	BASF-AT1	BASF-AT2	BASF-Corn	BASF-Soy	AFC1	AT4g01320	AF007269
PPI-AtCPP	X									
PPI-BnCPP	94	X								
PPI-SoyCPP	83	83	X							
BASF-AT1	98	95	83	X						
BASF-AT2	99	95	83	99	X					
BASF-Corn	82	82	79	82	82	X				
BASF-Soy	83	83	99	83	83	73	X			
AFC1	98	95	83	99	99	82	83	X		
AT4g01320	95	93	82	96	96	72	76	96	X	
AF007269	98	94	82	98	99	82	82	98	100	X

Figure 2

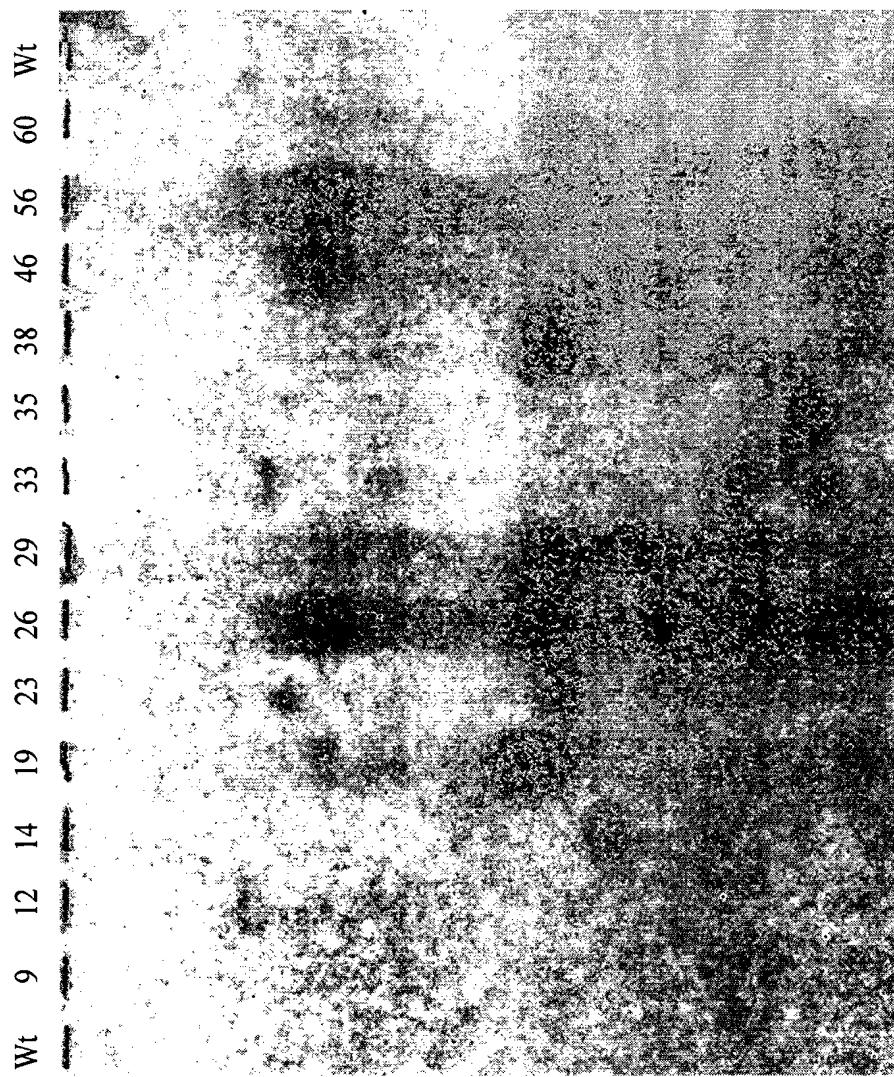


Figure 3.

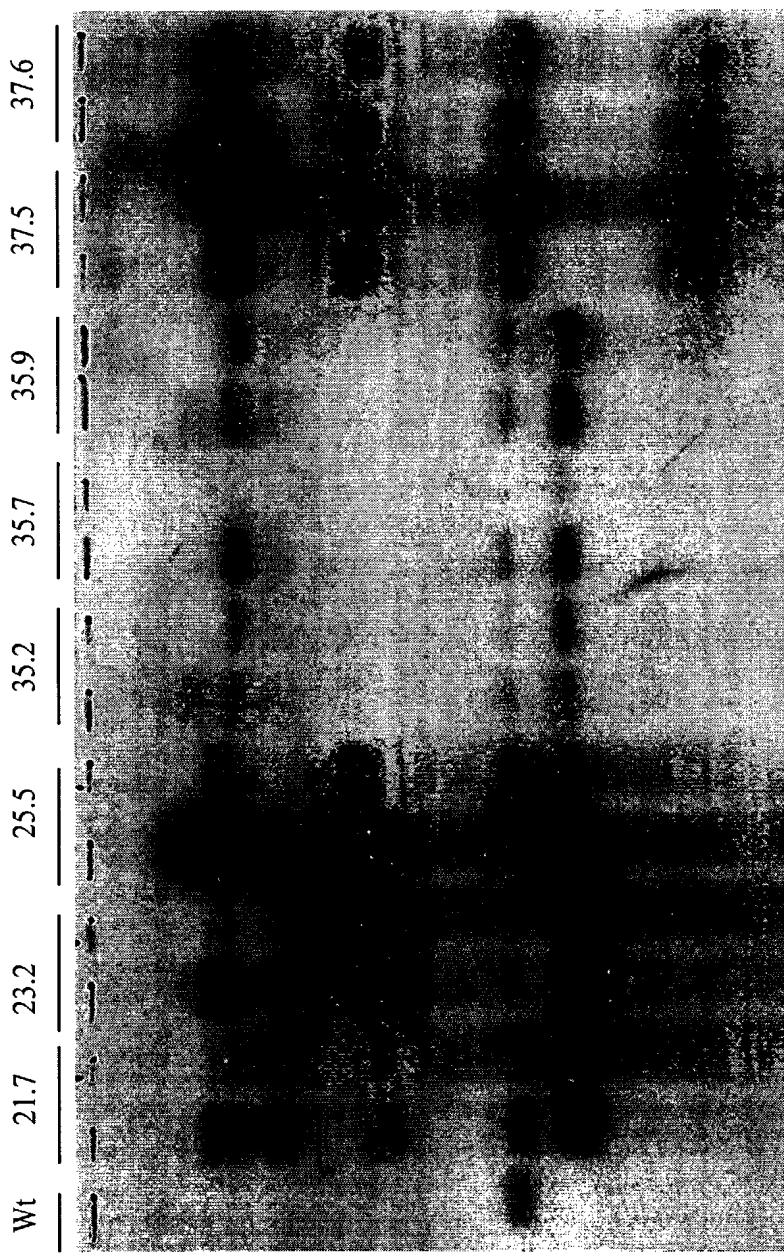


Figure 4.

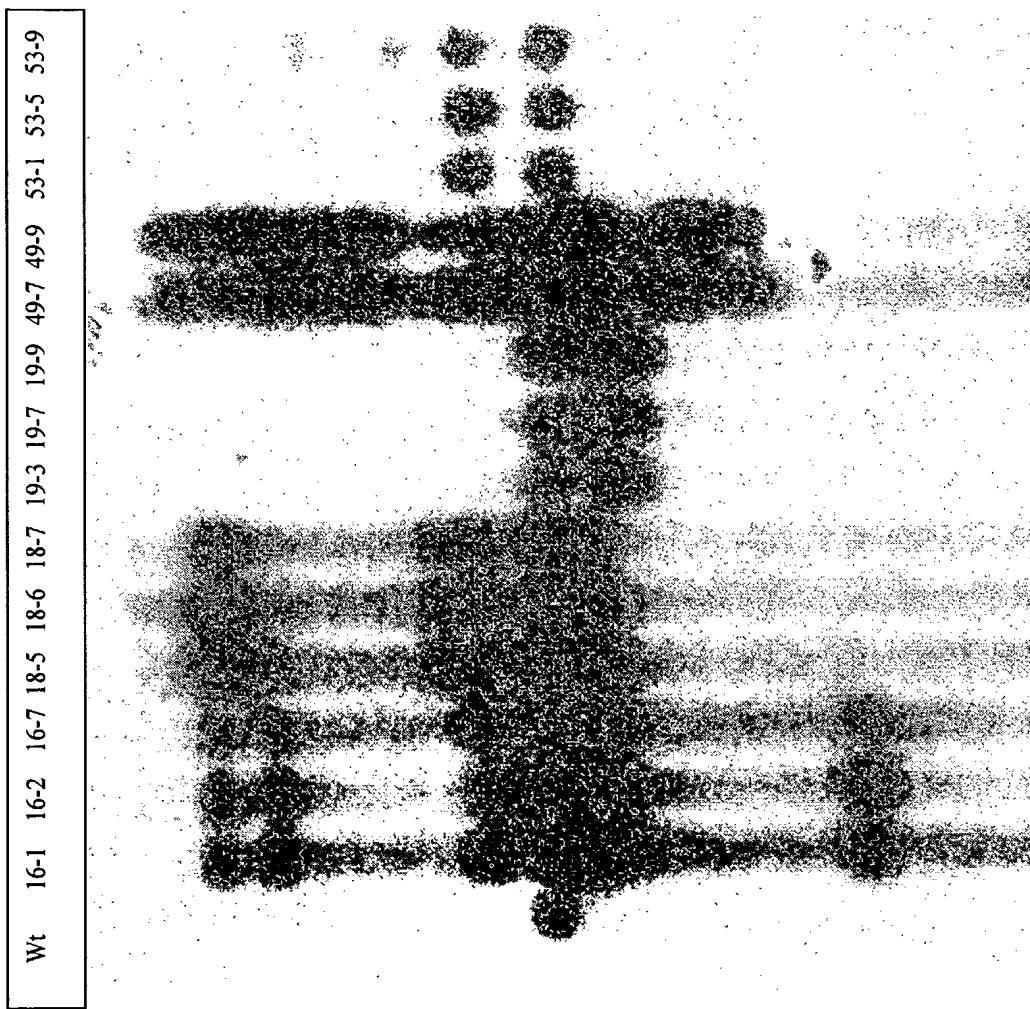


Figure 5

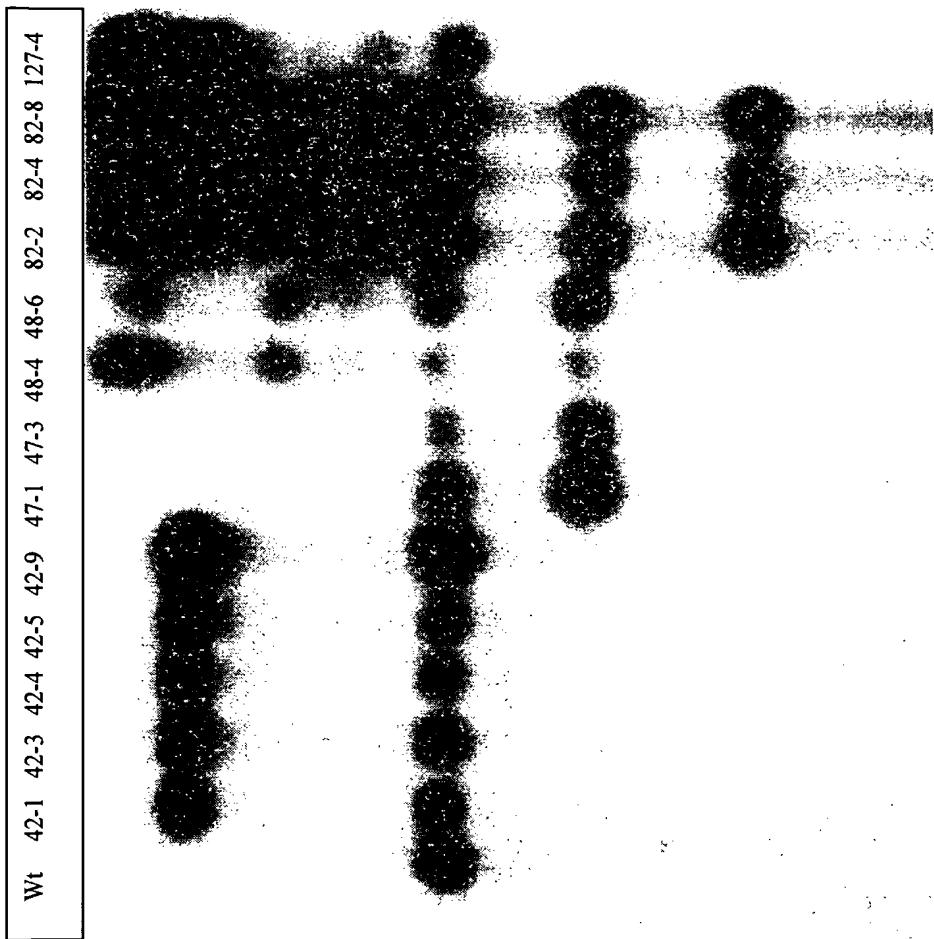


Figure 6

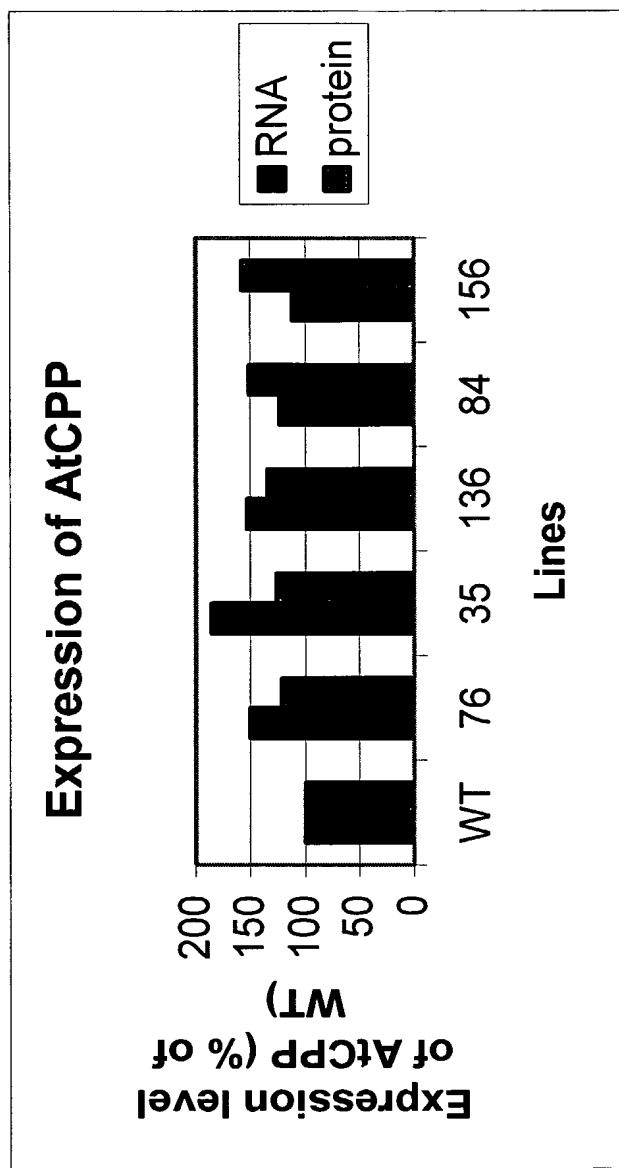


Figure 7

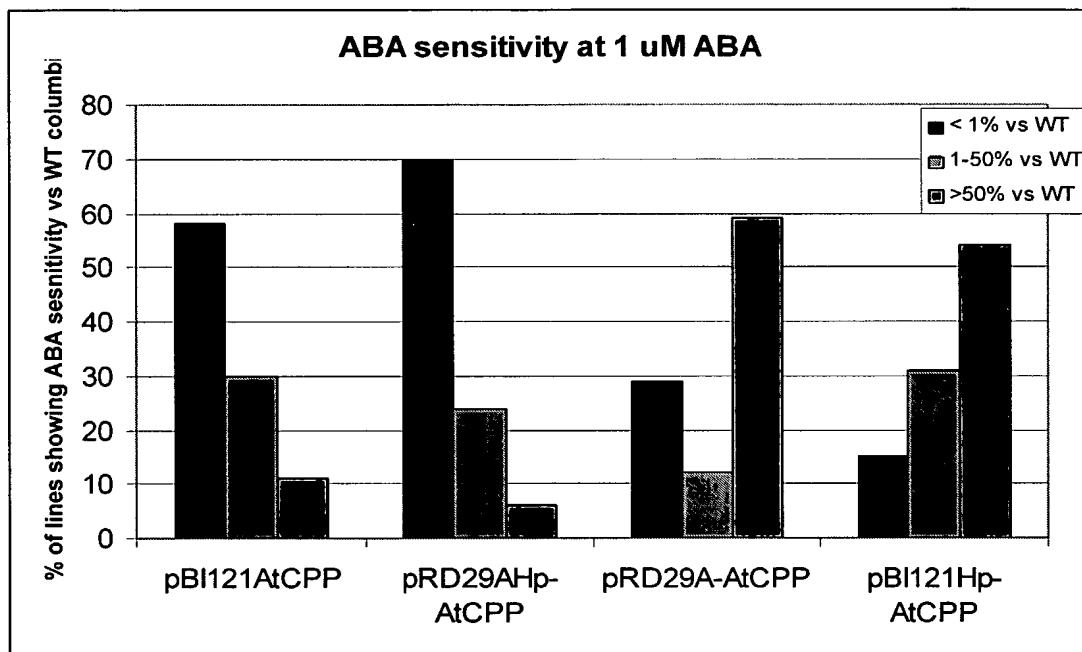


Figure 8

2 weeks old seedling on different [ABA]

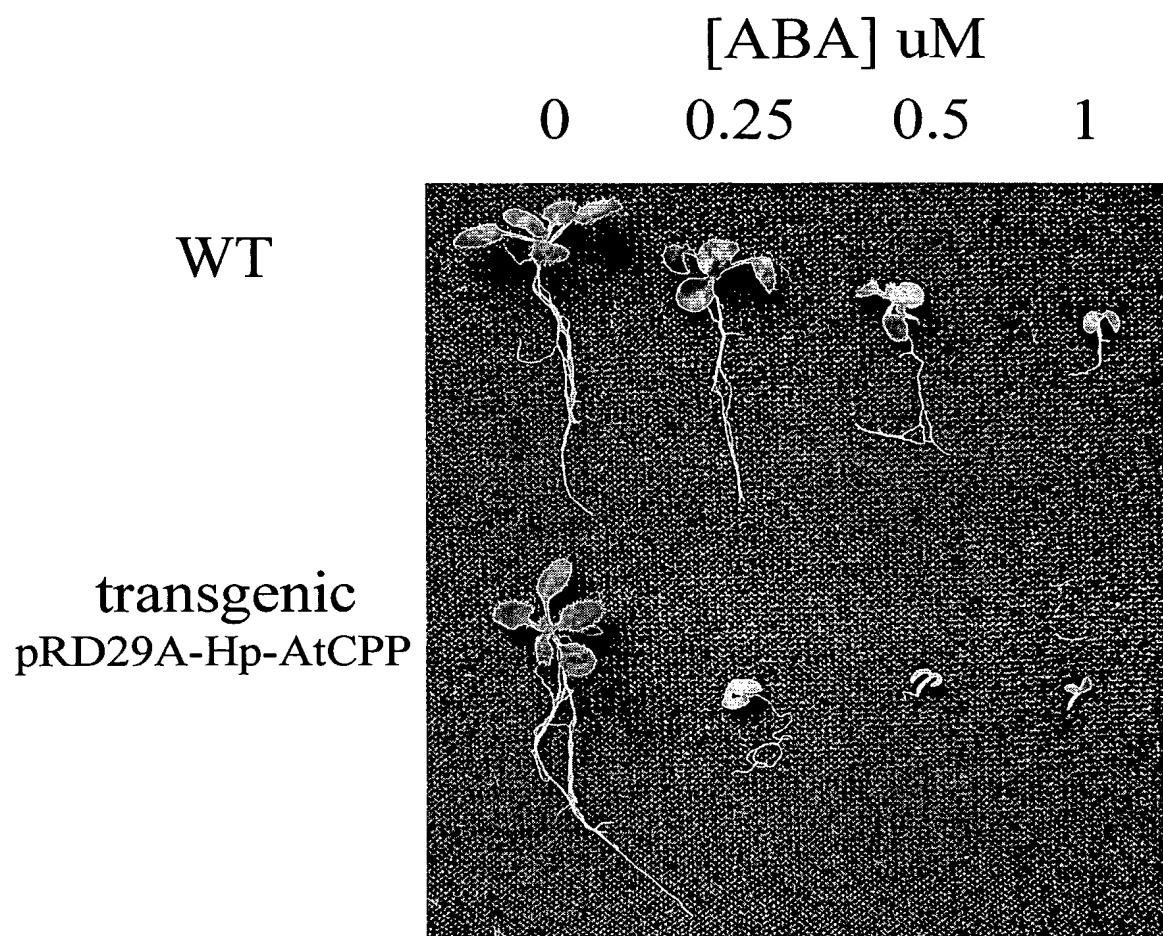


Figure 9

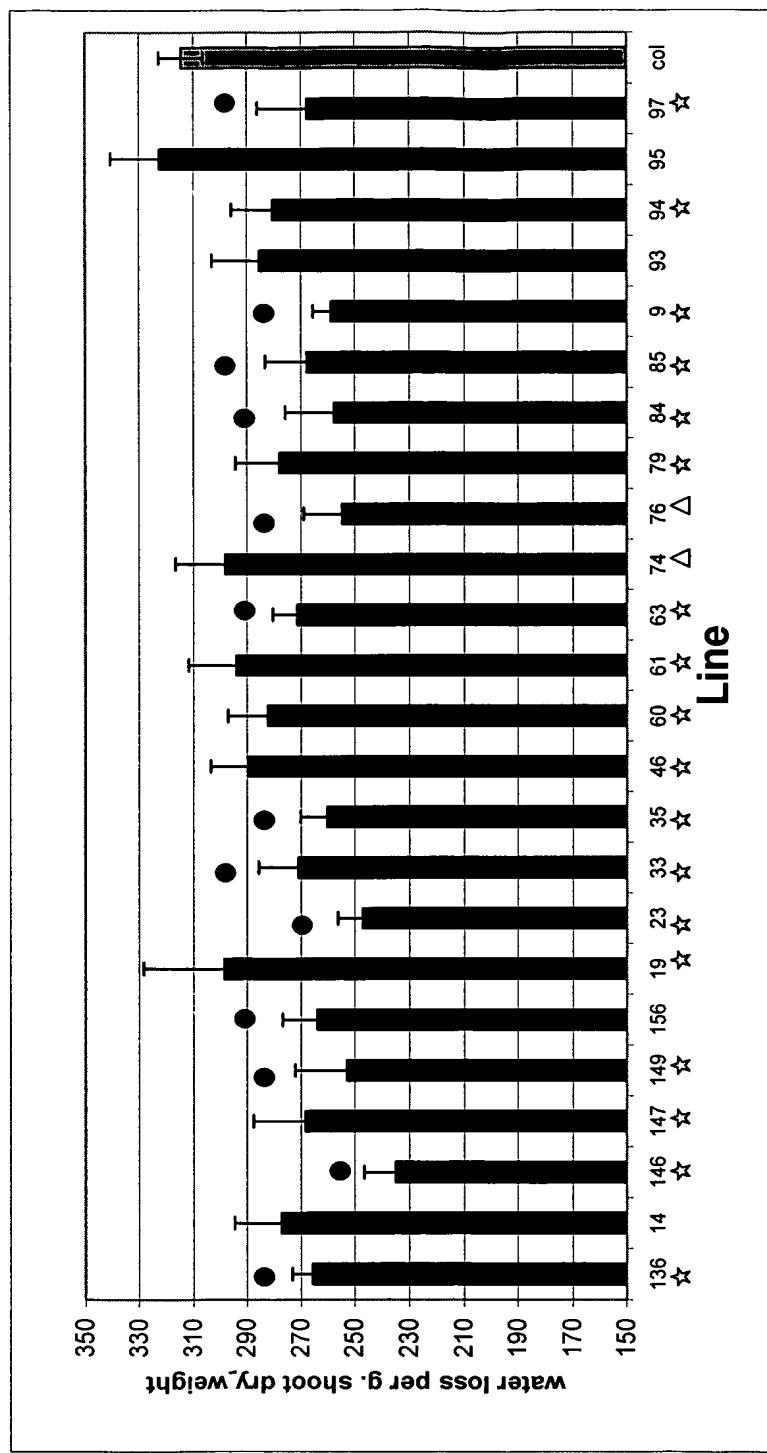


Figure 10.

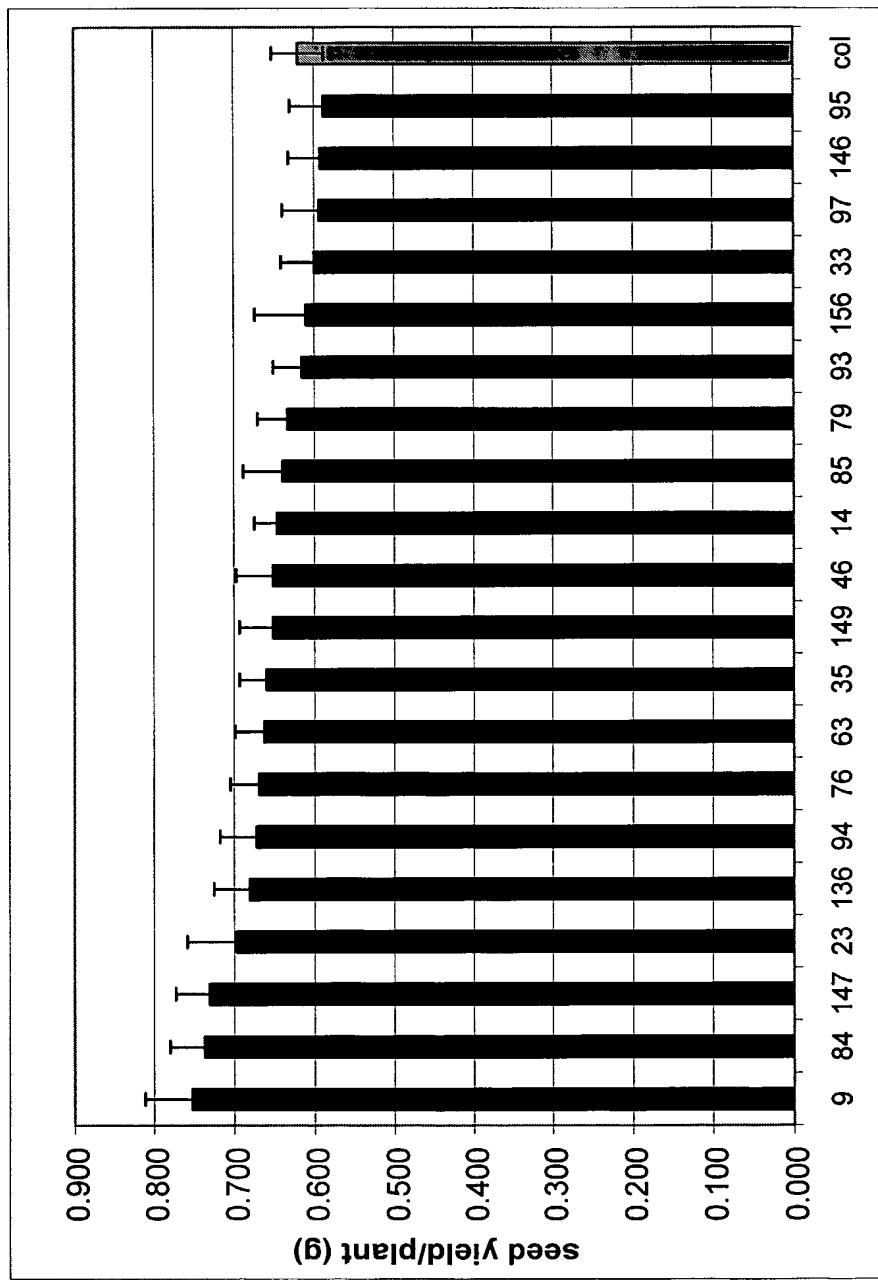


Figure 11

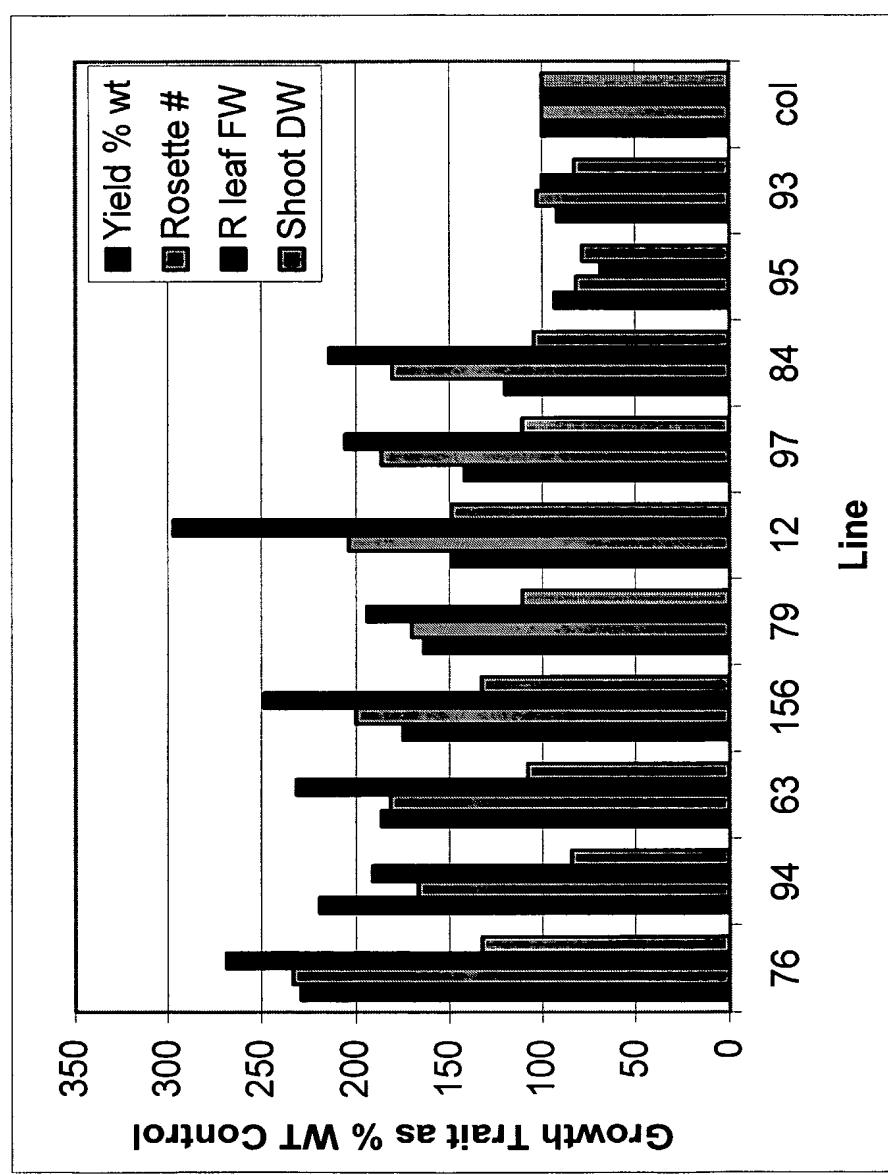


Figure 12

35S-AtCPP 12 days old seedlings - toluidin blue

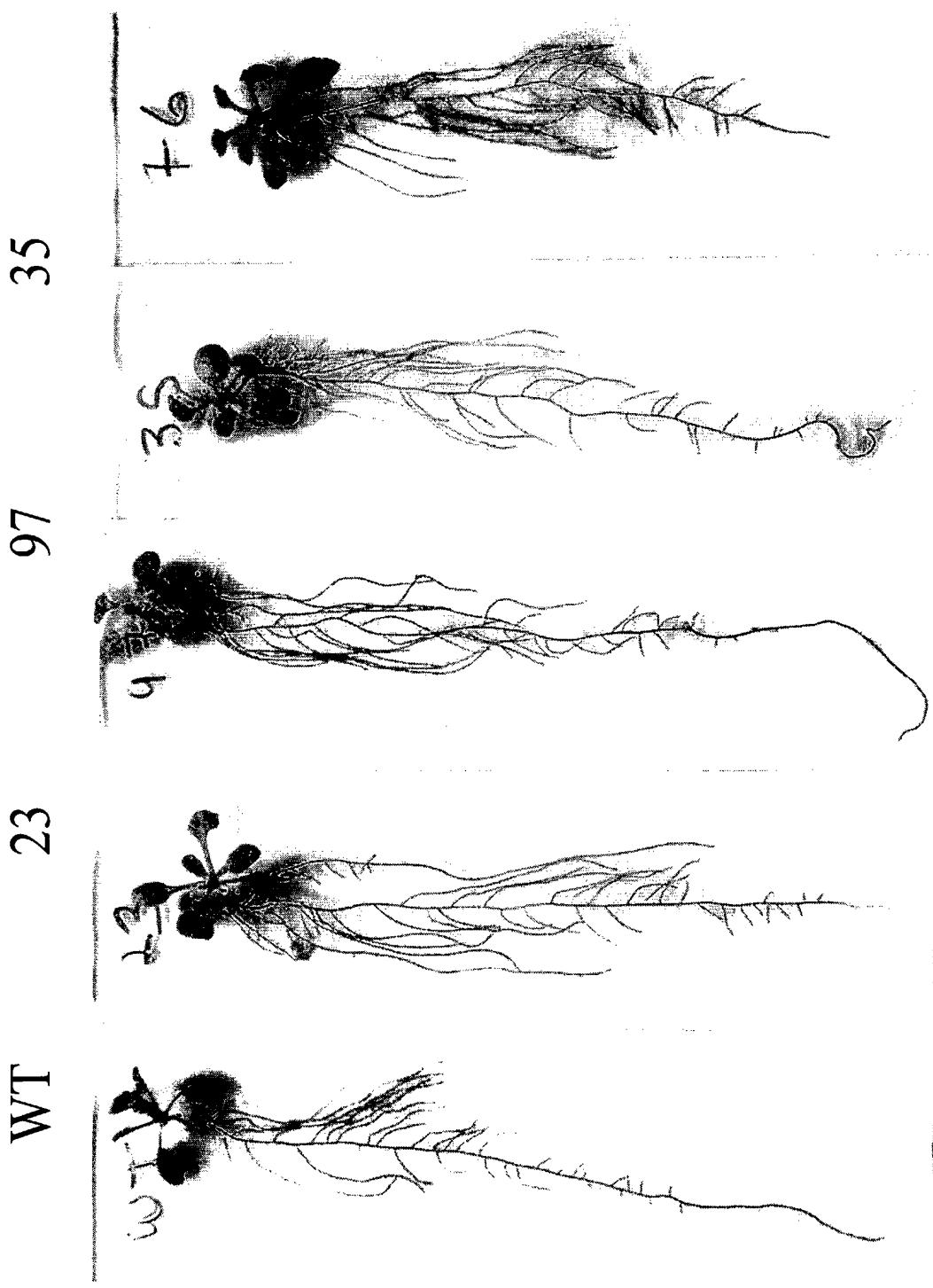


Figure 13

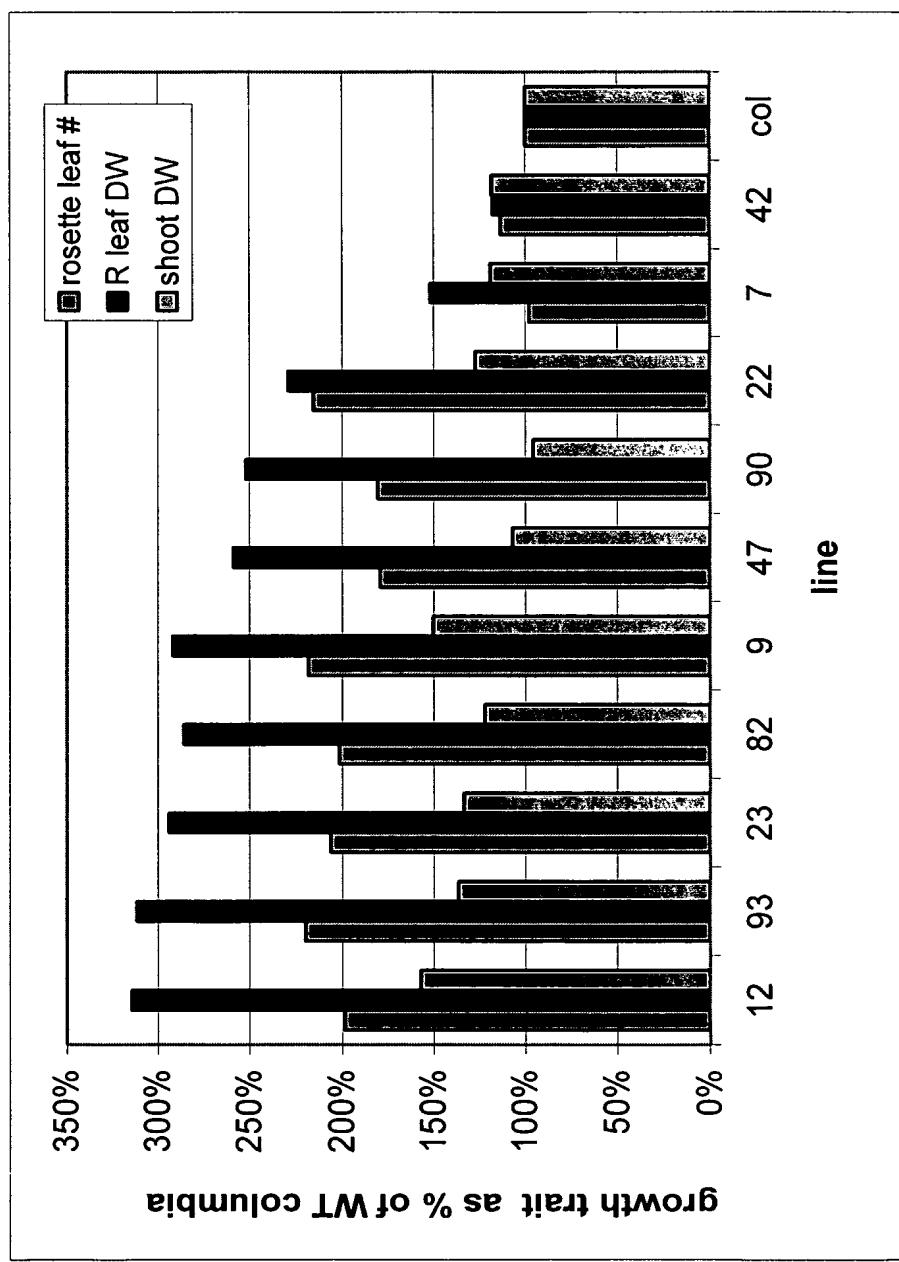


Figure 14